

Multivariate Normal Log-likelihoods in the **mvtnorm** Package ¹

Torsten Hothorn

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Chapter 1

Introduction

This document describes an implementation of [Genz \(1992\)](#) and, partially, of [Genz and Bretz \(2002\)](#), for the evaluation of N multivariate J -dimensional normal probabilities

$$p_i(\mathbf{C}_i \mid \mathbf{a}_i, \mathbf{b}_i) = \mathbb{P}(\mathbf{a}_i < \mathbf{Y}_i \leq \mathbf{b}_i \mid \mathbf{C}_i) = (2\pi)^{-\frac{J}{2}} \det(\mathbf{C}_i)^{-1} \int_{\mathbf{a}_i}^{\mathbf{b}_i} \exp\left(-\frac{1}{2} \mathbf{y}^\top \mathbf{C}_i^{-1} \mathbf{y}\right) d\mathbf{y} \quad (1.1)$$

where $\mathbf{a}_i = (a_1^{(i)}, \dots, a_J^{(i)})^\top \in \mathbb{R}^J$ and $\mathbf{b}_i = (b_1^{(i)}, \dots, b_J^{(i)})^\top \in \mathbb{R}^J$ are integration limits, $\mathbf{C}_i = (c_{jj}^{(i)}) \in \mathbb{R}^{J \times J}$ is a lower triangular matrix with $c_{jj}^{(i)} = 0$ for $1 \leq j < J < J$, and thus $\mathbf{Y}_i \sim \mathbb{N}_J(\mathbf{0}_J, \mathbf{C}_i \mathbf{C}_i^\top)$ for $i = 1, \dots, N$.

One application of these integrals is the estimation of the Cholesky factor \mathbf{C} of a J -dimensional normal distribution based on N interval-censored observations $\mathbf{Y}_1, \dots, \mathbf{Y}_N$ (encoded by \mathbf{a} and \mathbf{b}) via maximum-likelihood

$$\hat{\mathbf{C}} = \operatorname{argmax}_{\mathbf{C}} \sum_{i=1}^N \log(p_i(\mathbf{C} \mid \mathbf{a}_i, \mathbf{b}_i)).$$

In other applications, the Cholesky factor might also depend on i in some structured way.

Function `pmvnorm` in package `mvtnorm` computes p_i based on the covariance matrix $\mathbf{C}_i \mathbf{C}_i^\top$. However, the Cholesky factor \mathbf{C}_i of the given covariance matrix is computed in FORTRAN first each time this function is called. Function `pmvnorm` is not vectorised over $i = 1, \dots, N$ and thus separate calls to this function are necessary in order to compute likelihood contributions.

The implementation described here is a re-implementation (in R and C) of Alan Genz' original FORTRAN code, focusing on efficient computation of the log-likelihood $\sum_{i=1}^N \log(p_i)$ and the corresponding score function.

The document first describes a class and some useful methods for dealing with multiple lower triangular matrices $\mathbf{C}_i, i = 1, \dots, N$ in Chapter 2. The multivariate normal log-likelihood, and the corresponding score function, is implemented as outlined in Chapter 3. An example demonstrating maximum-likelihood estimation of Cholesky factors in the presence of interval-censored observations is discussed in Chapter 4. We use the technology developed here to implement the log-likelihood and score function for situations where some variables have been observed exactly and others only in form of interval-censoring in Chapter 5 and for nonparametric maximum-likelihood estimation in unstructured Gaussian copulae in Chapter 6. An attempt to provide useRs with a simple and (hopefully) bullet proof interface is documented in Chapter 8.

The development of this infrastructure was motivated by the necessity to evaluate probabilities (1.1) arising in the likelihood of multivariate conditional transformation models ([Klein et al., 2022](#)) for discrete or censored observations. Some forms of the likelihood for such nonparanormal models are discussed in [Hothorn \(2024\)](#).

Chapter 2

Lower Triangular Matrices

"ltMatrices.R" 2≡

- ⟨ *R Header 145* ⟩
- ⟨ *ltMatrices 6a* ⟩
- ⟨ *syMatrices 6b* ⟩
- ⟨ *dim ltMatrices 6c* ⟩
- ⟨ *dimnames ltMatrices 7a* ⟩
- ⟨ *names ltMatrices 7b* ⟩
- ⟨ *is.ltMatrices 7c* ⟩
- ⟨ *as.ltMatrices 119b* ⟩
- ⟨ *print ltMatrices 11* ⟩
- ⟨ *reorder ltMatrices 12* ⟩
- ⟨ *subset ltMatrices 14* ⟩
- ⟨ *lower triangular elements 17* ⟩
- ⟨ *diagonals ltMatrices 19* ⟩
- ⟨ *diagonal matrix 22* ⟩
- ⟨ *mult ltMatrices 23a* ⟩
- ⟨ *mult syMatrices 27* ⟩
- ⟨ *solve ltMatrices 31* ⟩
- ⟨ *logdet ltMatrices 33b* ⟩
- ⟨ *tcrossprod ltMatrices 37* ⟩
- ⟨ *crossprod ltMatrices 38* ⟩
- ⟨ *crossprod tcrossprod methods 39* ⟩
- ⟨ *chol syMatrices 41* ⟩
- ⟨ *add diagonal elements 20* ⟩
- ⟨ *assign diagonal elements 21* ⟩
- ⟨ *kronecker vec trick 46* ⟩
- ⟨ *convenience functions 50* ⟩
- ⟨ *aperm 53a, ...* ⟩
- ⟨ *marginal 54b* ⟩
- ⟨ *conditional 57* ⟩
- ⟨ *check obs 59b* ⟩
- ⟨ *colSumsdnorm ltMatrices 60b* ⟩

◇

```

"ltMatrices.c" 3≡

  < C Header 146 >
  #ifndef USE_FC_LEN_T
  # define USE_FC_LEN_T
  #endif
  #include <Rconfig.h>
  #include <R_ext/Lapack.h> /* for dtptri */
  #ifndef FCONE
  # define FCONE
  #endif
  #include <R.h>
  #include <Rmath.h>
  #include <Rinternals.h>
  #include <Rdefines.h>
  < colSumsdnorm 60a >
  < solve 29 >
  < solve C 30 >
  < logdet 33a >
  < tcrossprod 36 >
  < mult 24b >
  < mult transpose 26 >
  < chol 42 >
  < vec trick 44a >
  ◇

```

We first define and implement infrastructure for dealing with multiple lower triangular matrices $\mathbf{C}_i \in \mathbb{R}^{J \times J}$ for $i = 1, \dots, N$. We note that each such matrix \mathbf{C} can be stored in a vector of length $J(J+1)/2$. If all diagonal elements are one (that is, $c_{jj}^{(i)} \equiv 1, j = 1, \dots, J$), the length of this vector is $J(J-1)/2$.

2.1 Multiple Lower Triangular Matrices

We can store N such matrices in an $J(J+1)/2 \times N$ matrix (`diag = TRUE`) or, for `diag = FALSE`, in an $J(J-1)/2 \times N$ matrix.

Each vector might define the corresponding lower triangular matrix either in row or column-major order:

$$\begin{aligned}
\mathbf{C} &= \begin{pmatrix} c_{11} & & & 0 \\ c_{21} & c_{22} & & \\ c_{31} & c_{32} & c_{33} & \\ \vdots & \vdots & & \ddots \\ c_{J1} & c_{J2} & \dots & c_{JJ} \end{pmatrix} \text{matrix indexing} \\
&= \begin{pmatrix} c_1 & & & 0 \\ c_2 & c_{J+1} & & \\ c_3 & c_{J+2} & c_{2J} & \\ \vdots & \vdots & & \ddots \\ c_J & c_{2J-1} & \dots & c_{J(J+1)/2} \end{pmatrix} \text{column-major, byrow = FALSE} \\
&= \begin{pmatrix} & c_1 & & & & 0 \\ & c_2 & & c_3 & & \\ & c_4 & & c_5 & c_6 & \\ & \vdots & & \vdots & & \ddots \\ c_{J((J+1)/2-1)+1} & c_{J((J+1)/2-1)+2} & \dots & & & c_{J(J+1)/2} \end{pmatrix} \text{row-major, byrow = TRUE}
\end{aligned}$$

Based on some matrix object, the dimension J is computed and checked as

`<ltMatrices dim 4> ≡`

```

J <- floor((1 + sqrt(1 + 4 * 2 * nrow(object))) / 2 - diag)
if (nrow(object) != J * (J - 1) / 2 + diag * J)
  stop("Dimension of object does not correspond to lower
       triangular part of a square matrix")

```

◇

Fragment referenced in [6a](#).

Typically the J dimensions are associated with names, and we therefore compute identifiers for the vector elements in either column- or row-major order on request (for later printing)

<ltMatrices names 5a> ≡

```
nonames <- FALSE
if (!isTRUE(names)) {
  if (is.character(names))
    stopifnot(is.character(names) &&
              length(unique(names)) == J)
  else
    nonames <- TRUE
} else {
  names <- as.character(seq_len(J))
}

if (!nonames) {
  L1 <- matrix(names, nrow = J, ncol = J)
  L2 <- matrix(names, nrow = J, ncol = J, byrow = TRUE)
  L <- matrix(paste(L1, L2, sep = "."), nrow = J, ncol = J)
  if (byrow)
    rownames(object) <- t(L)[upper.tri(L, diag = diag)]
  else
    rownames(object) <- L[lower.tri(L, diag = diag)]
} # else {      ### add later
# warning("ltMatrices objects should be properly named")
# }
◇
```

Fragment referenced in [6a](#).

If `object` is already a classed object representing lower triangular matrices (we will use the class name `ltMatrices`), we might want to change the storage form from row- to column-major or the other way round.

<ltMatrices input 5b> ≡

```
if (is.ltMatrices(object)) {
  cls <- class(object)          ### keep inheriting classes
  ret <- .reorder(object, byrow = byrow)
  class(ret) <- class(object)
  return(ret)
}
◇
```

Fragment referenced in [6a](#).

The constructor essentially attaches attributes to a matrix `object`, possibly after some reordering / transposing

< ltMatrices 6a > ≡

```
ltMatrices <- function(object, diag = FALSE, byrow = FALSE, names = TRUE) {  
  if (!is.matrix(object))  
    object <- matrix(object, ncol = 1L)  
  
  < ltMatrices input 5b >  
  < ltMatrices dim 4 >  
  < ltMatrices names 5a >  
  
  attr(object, "J")      <- J  
  attr(object, "diag")   <- diag  
  attr(object, "byrow")  <- byrow  
  attr(object, "rcnames") <- names  
  
  class(object) <- c("ltMatrices", class(object))  
  object  
}  
◇
```

Fragment referenced in 2.

For the sake of completeness, we also add a constructor for multiple symmetric matrices

< syMatrices 6b > ≡

```
as.syMatrices <- function(x) {  
  if (is.syMatrices(x))  
    return(x)  
  x <- as.ltMatrices(x)      ### make sure "ltMatrices"  
                             ### is first class  
  class(x)[1L] <- "syMatrices"  
  return(x)  
}  
syMatrices <- function(object, diag = FALSE, byrow = FALSE, names = TRUE)  
  as.syMatrices(ltMatrices(object = object, diag = diag, byrow = byrow,  
    names = names))  
◇
```

Fragment referenced in 2.

The dimensions of such an object are always $N \times J \times J$ and are given by

< dim ltMatrices 6c > ≡

```
dim.ltMatrices <- function(x) {  
  J <- attr(x, "J")  
  return(c(attr(x, "dim")[2L], J, J)) ### ncol(unclass(x)) may trigger gc  
}  
dim.syMatrices <- dim.ltMatrices  
◇
```

Fragment referenced in 2.

The corresponding dimnames can be extracted as

< dimnames ltMatrices 7a > ≡

```
dimnames.ltMatrices <- function(x)
  return(list(attr(x, "dimnames")[[2L]], attr(x, "rcnames"), attr(x, "rcnames")))
dimnames.syMatrices <- dimnames.ltMatrices
◇
```

Fragment referenced in 2.

The names identifying rows and columns in each C_i are

< names ltMatrices 7b > ≡

```
names.ltMatrices <- function(x) {
  return(attr(x, "dimnames")[[1L]])
}
names.syMatrices <- names.ltMatrices
◇
```

Fragment referenced in 2.

Finally, let's add two functions for checking the class and a function for coersing classes inheriting from `ltMatrices` to the latter, the same for `syMatrices`. Furthermode, `as.ltMatrices` coerces objects inheriting from `syMatrices` or `ltMatrices` to class `ltMatrices` (that is, `chol` or `invchol` is removed from the class list, unlike a call to the constructor `ltMatrices`). A default method is added in Chapter 8.

< is.ltMatrices 7c > ≡

```
is.ltMatrices <- function(x) inherits(x, "ltMatrices")
is.syMatrices <- function(x) inherits(x, "syMatrices")
as.ltMatrices <- function(x) UseMethod("as.ltMatrices")
as.ltMatrices.syMatrices <- function(x) {
  cls <- class(x)
  class(x) <- cls[which(cls == "syMatrices"):length(cls)]
  class(x)[1L] <- "ltMatrices"
  return(x)
}
as.ltMatrices.ltMatrices <- function(x) {
  cls <- class(x)
  class(x) <- cls[which(cls == "ltMatrices"):length(cls)]
  return(x)
}
◇
```

Fragment referenced in 2.

Let's set-up an example for illustration. Throughout this document, we will compare numerical results using

```
> chk <- function(...) stopifnot(isTRUE(all.equal(...)))
```

We start with a a simple example demonstrating how to set-up `ltMatrices` objects

```
> library("mvtnorm")
> set.seed(290875)
```

```

> N <- 4L
> J <- 5L
> rn <- paste0("C_", 1:N)
> nm <- LETTERS[1:J]
> Jn <- J * (J - 1) / 2
> ## data
> xn <- matrix(runif(N * Jn), ncol = N)
> colnames(xn) <- rn
> xd <- matrix(runif(N * (Jn + J)), ncol = N)
> colnames(xd) <- rn
> (lxn <- ltMatrices(xn, byrow = TRUE, names = nm))

, , C_1

      A      B      C      D      E
A 1.0000000 .      .      .      .
B 0.5123660 1.0000000 .      .      .
C 0.0584725 0.9095137 1.0000000 .      .
D 0.3944872 0.6612143 0.2335259 1.0000000 .
E 0.5164752 0.2979867 0.0751775 0.8182123 1.0000000

, , C_2

      A      B      C      D      E
A 1.0000000 .      .      .      .
B 0.8590665 1.0000000 .      .      .
C 0.3744315 0.1022684 1.0000000 .      .
D 0.1165248 0.7956529 0.8930589 1.0000000 .
E 0.1948049 0.4730419 0.2377852 0.2146060 1.0000000

, , C_3

      A      B      C      D      E
A 1.0000000 .      .      .      .
B 0.4530153 1.0000000 .      .      .
C 0.9045608 0.9269936 1.0000000 .      .
D 0.4490011 0.1326375 0.4153967 1.0000000 .
E 0.9574833 0.4917481 0.7160702 0.2938002 1.0000000

, , C_4

      A      B      C      D      E
A 1.0000000 .      .      .      .
B 0.4877241 1.0000000 .      .      .
C 0.0593046 0.7625270 1.0000000 .      .
D 0.0005227 0.1995700 0.4705089 1.0000000 .
E 0.4913541 0.2849431 0.0059611 0.8901458 1.0000000

> dim(lxn)

[1] 4 5 5

> dimnames(lxn)

[[1]]
[1] "C_1" "C_2" "C_3" "C_4"

```

```

[[2]]
[1] "A" "B" "C" "D" "E"

[[3]]
[1] "A" "B" "C" "D" "E"

> lxd <- ltMatrices(xd, byrow = TRUE, diag = TRUE, names = nm)
> dim(lxd)

[1] 4 5 5

> dimnames(lxd)

[[1]]
[1] "C_1" "C_2" "C_3" "C_4"

[[2]]
[1] "A" "B" "C" "D" "E"

[[3]]
[1] "A" "B" "C" "D" "E"

> lxn <- as.syMatrices(lxn)
> lxn

, , C_1

      A      B      C      D      E
A 1.00000 0.5124 0.05847 0.3945 0.51648
B 0.51237 1.0000 0.90951 0.6612 0.29799
C 0.05847 0.9095 1.00000 0.2335 0.07518
D 0.39449 0.6612 0.23353 1.0000 0.81821
E 0.51648 0.2980 0.07518 0.8182 1.00000

, , C_2

      A      B      C      D      E
A 1.0000 0.8591 0.3744 0.1165 0.1948
B 0.8591 1.0000 0.1023 0.7957 0.4730
C 0.3744 0.1023 1.0000 0.8931 0.2378
D 0.1165 0.7957 0.8931 1.0000 0.2146
E 0.1948 0.4730 0.2378 0.2146 1.0000

, , C_3

      A      B      C      D      E
A 1.0000 0.4530 0.9046 0.4490 0.9575
B 0.4530 1.0000 0.9270 0.1326 0.4917
C 0.9046 0.9270 1.0000 0.4154 0.7161
D 0.4490 0.1326 0.4154 1.0000 0.2938
E 0.9575 0.4917 0.7161 0.2938 1.0000

, , C_4

```

	A	B	C	D	E
A	1.0000000	0.4877	0.059305	0.0005227	0.491354
B	0.4877241	1.0000	0.762527	0.1995700	0.284943
C	0.0593046	0.7625	1.000000	0.4705089	0.005961
D	0.0005227	0.1996	0.470509	1.0000000	0.890146
E	0.4913541	0.2849	0.005961	0.8901458	1.000000

2.2 Printing

For pretty printing, we coerce objects of class `ltMatrices` to `array`. The method has a logical argument called `symmetric`, forcing the lower triangular matrix to be interpreted as a symmetric matrix.

`<extract slots 10> ≡`

```
diag <- attr(x, "diag")
byrow <- attr(x, "byrow")
d <- dim(x)
J <- d[2L]
dn <- dimnames(x)
◇
```

Fragment referenced in [11](#), [12](#), [13](#), [17](#), [19](#), [21](#), [23a](#), [27](#).

<print ltMatrices 11> ≡

```
as.array.ltMatrices <- function(x, symmetric = FALSE, ...) {  
  <extract slots 10>  
  x <- unclass(x)  
  
  L <- matrix(1L, nrow = J, ncol = J)  
  diag(L) <- 2L  
  if (byrow) {  
    L[upper.tri(L, diag = diag)] <- floor(2L + 1:(J * (J - 1) / 2L + diag * J))  
    L <- t(L)  
  } else {  
    L[lower.tri(L, diag = diag)] <- floor(2L + 1:(J * (J - 1) / 2L + diag * J))  
  }  
  if (symmetric) {  
    L[upper.tri(L)] <- 0L  
    dg <- diag(L)  
    L <- L + t(L)  
    diag(L) <- dg  
  }  
  ret <- rbind(0, 1, x)[c(L), , drop = FALSE]  
  class(ret) <- "array"  
  dim(ret) <- d[3:1]  
  dimnames(ret) <- dn[3:1]  
  return(ret)  
}  
  
as.array.syMatrices <- function(x, ...)  
  return(as.array.ltMatrices(x, symmetric = TRUE))  
  
print.ltMatrices <- function(x, zero.print = ".", ...) {  
  if (is.null(attr(x, "dimnames")[[2L]]))  
    attr(x, "dimnames")[[2L]] <- as.character(seq_len(dim(x)[1L]))  
  print(as.table(as.array(x)), zero.print = zero.print, ...)  
}  
  
print.syMatrices <- function(x, ...)  
  print(as.array(x))  
  ◊
```

Fragment referenced in 2.

Symmetric matrices are represented by lower triangular matrix objects, but we change the class from `ltMatrices` to `syMatrices` (which disables all functionality except printing and coercion to arrays).

2.3 Reordering

It is sometimes convenient to have access to lower triangular matrices in either column- or row-major order and this little helper function switches between the two forms

< reorder ltMatrices 12 > ≡

```
.reorder <- function(x, byrow = FALSE) {  
  
  stopifnot(is.ltMatrices(x))  
  if (attr(x, "byrow") == byrow) return(x)  
  
  < extract slots 10 >  
  
  x <- unclass(x)  
  
  rL <- cL <- diag(0, nrow = J)  
  rL[lower.tri(rL, diag = diag)] <- cL[upper.tri(cL, diag = diag)] <- 1:nrow(x)  
  cL <- t(cL)  
  if (byrow) ### row -> col order  
    return(ltMatrices(x[cL[lower.tri(cL, diag = diag)], , drop = FALSE],  
                     diag = diag, byrow = FALSE, names = dn[[2L]]))  
  ### col -> row order  
  return(ltMatrices(x[t(rL)[upper.tri(rL, diag = diag)], , drop = FALSE],  
                   diag = diag, byrow = TRUE, names = dn[[2L]]))  
}  
◇
```

Fragment referenced in 2.

We can check if this works by switching back and forth between column-major and row-major order

```
> ## constructor + .reorder + as.array  
> a <- as.array(ltMatrices(xn, byrow = TRUE))  
> b <- as.array(ltMatrices(ltMatrices(xn, byrow = TRUE),  
+                       byrow = FALSE))  
> chk(a, b)  
> a <- as.array(ltMatrices(xn, byrow = FALSE))  
> b <- as.array(ltMatrices(ltMatrices(xn, byrow = FALSE),  
+                       byrow = TRUE))  
> chk(a, b)  
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE))  
> b <- as.array(ltMatrices(ltMatrices(xd, byrow = TRUE, diag = TRUE),  
+                       byrow = FALSE))  
> chk(a, b)  
> a <- as.array(ltMatrices(xd, byrow = FALSE, diag = TRUE))  
> b <- as.array(ltMatrices(ltMatrices(xd, byrow = FALSE, diag = TRUE),  
+                       byrow = TRUE))  
> chk(a, b)
```

2.4 Subsetting

We might want to select subsets of observations $i \in \{1, \dots, N\}$ or rows/columns $j \in \{1, \dots, J\}$ of the corresponding matrices \mathbf{C}_i .

⟨.subset ltMatrices 13⟩ ≡

```
.subset_ltMatrices <- function(x, i, j, ..., drop = FALSE) {  
  
  if (drop) warning("argument drop is ignored")  
  if (missing(i) && missing(j)) return(x)  
  
  ⟨extract slots 10⟩  
  
  x <- unclass(x)  
  
  if (!missing(j)) {  
  
    if (is.character(j)) {  
      stopifnot(all(j %in% dn[[2L]]))  
      j <- match(j, dn[[2L]])  
    }  
  
    j <- (1:J)[j] ### get rid of negative indices  
  
    if (length(j) == 1L && !diag) {  
      return(ltMatrices(matrix(1, ncol = ncol(x), nrow = 1), diag = TRUE,  
                             byrow = byrow, names = dn[[2L]][j]))  
    }  
    L <- diag(0L, nrow = J)  
    Jp <- sum(upper.tri(L, diag = diag))  
    if (byrow) {  
      L[upper.tri(L, diag = diag)] <- 1:Jp  
      L <- L + t(L)  
      diag(L) <- diag(L) / 2  
      L <- L[j, j, drop = FALSE]  
      L <- L[upper.tri(L, diag = diag)]  
    } else {  
      L[lower.tri(L, diag = diag)] <- 1:Jp  
      L <- L + t(L)  
      diag(L) <- diag(L) / 2  
      L <- L[j, j, drop = FALSE]  
      L <- L[lower.tri(L, diag = diag)]  
    }  
    if (missing(i)) {  
      return(ltMatrices(x[c(L), , drop = FALSE], diag = diag,  
                       byrow = byrow, names = dn[[2L]][j]))  
    }  
    return(ltMatrices(x[c(L), i, drop = FALSE], diag = diag,  
                     byrow = byrow, names = dn[[2L]][j]))  
  }  
  return(ltMatrices(x[, i, drop = FALSE], diag = diag,  
                   byrow = byrow, names = dn[[2L]]))  
}  
◇
```

Fragment referenced in 14.

`<subset ltMatrices 14> ≡`

```
<.subset ltMatrices 13>
### if j is not ordered, result is not a lower triangular matrix
[".ltMatrices" <- function(x, i, j, ..., drop = FALSE) {
  if (!missing(j)) {
    if (is.character(j)) {
      stopifnot(all(j %in% dimnames(x)[[2L]]))
      j <- match(j, dimnames(x)[[2L]])
    }
    if (all(j > 0)) {
      if (any(diff(j) < 0)) stop("invalid subset argument j")
    }
  }

  return(.subset_ltMatrices(x = x, i = i, j = j, ..., drop = drop))
}

[".syMatrices" <- function(x, i, j, ..., drop = FALSE) {
  x <- as.syMatrices(x)
  ret <- .subset_ltMatrices(x = x, i = i, j = j, ..., drop = drop)
  class(ret)[1L] <- "syMatrices"
  ret
}
◇
```

Fragment referenced in [2](#).

We check if this works by first subsetting the `ltMatrices` object. Second, we coerce the object to an array and do the subset for the latter object. Both results must agree.

```
> ## subset
> a <- as.array(ltMatrices(xn, byrow = FALSE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = FALSE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xn, byrow = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm))[j, j, i]
> chk(a, b)
```

We start with both indices being positive integers

```
> i <- colnames(xn)[1:2]
> j <- 2:4
> ## subset
> a <- as.array(ltMatrices(xn, byrow = FALSE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = FALSE, names = nm))[j, j, i]
```

```

> chk(a, b)
> a <- as.array(ltMatrices(xn, byrow = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm))[j, j, i]
> chk(a, b)

```

proceed with characters

```

> i <- 1:2
> j <- nm[2:4]
> ## subset
> a <- as.array(ltMatrices(xn, byrow = FALSE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = FALSE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xn, byrow = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm))[j, j, i]
> chk(a, b)

```

a different subset

```

> j <- c(1, 3, 5)
> ## subset
> a <- as.array(ltMatrices(xn, byrow = FALSE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = FALSE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xn, byrow = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,

```

```

+                               names = nm))[j, j, i]
> chk(a, b)

```

and characters again

```

> j <- nm[c(1, 3, 5)]
> ## subset
> a <- as.array(ltMatrices(xn, byrow = FALSE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = FALSE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xn, byrow = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm))[j, j, i]
> chk(a, b)

```

and finally with with negative subsets

```

> j <- -c(1, 3, 5)
> ## subset
> a <- as.array(ltMatrices(xn, byrow = FALSE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = FALSE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xn, byrow = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xn, byrow = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = FALSE,
+                       diag = TRUE, names = nm))[j, j, i]
> chk(a, b)
> a <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm)[i, j])
> b <- as.array(ltMatrices(xd, byrow = TRUE, diag = TRUE,
+                       names = nm))[j, j, i]
> chk(a, b)

```

and with non-increasing argument j (this won't work for lower triangular matrices, only for symmetric matrices)

```

> ## subset
> j <- nm[sample(1:J)]
> ltM <- ltMatrices(xn, byrow = FALSE, names = nm)
> try(ltM[i, j])
> ltM <- as.syMatrices(ltM)
> a <- as.array(ltM[i, j])
> b <- as.array(ltM)[j, j, i]
> chk(a, b)

```

Extracting the lower triangular elements from an `ltMatrices` object (or from an object of class `syMatrices`) returns a matrix with N columns, undoing the effect of `ltMatrices`. Note that ordering of the rows of this matrix depend on the `byrow` attribute of `x`, unless the `byrow` to this function is used to overwrite it explicitly

<lower triangular elements 17> \equiv

```
Lower_tri <- function(x, diag = FALSE, byrow = attr(x, "byrow")) {
  if (is.syMatrices(x))
    x <- as.ltMatrices(x)
  adiaq <- diag
  x <- ltMatrices(x, byrow = byrow)

  <extract slots 10>

  if (diag == adiaq)
    return(removeClass(x)[, , drop = FALSE]) ### remove attributes

  if (!diag && adiaq) {
    diagonals(x) <- 1
    return(removeClass(x)[, , drop = FALSE]) ### remove attributes
  }

  x <- unclass(x)
  if (J == 1) {
    idx <- 1L
  } else {
    if (byrow)
      idx <- cumsum(c(1, 2:J))
    else
      idx <- cumsum(c(1, J:2))
  }
  return(x[-idx, , drop = FALSE])
}
◇
```

Fragment referenced in 2.

```
> ## J <- 4
> M <- ltMatrices(matrix(1:10, nrow = 10, ncol = 2), diag = TRUE)
> Lower_tri(M, diag = FALSE)
```

```
  [,1] [,2]
2.1   2   2
3.1   3   3
4.1   4   4
3.2   6   6
4.2   7   7
4.3   9   9
```

```
> Lower_tri(M, diag = TRUE)
```

```
  [,1] [,2]
1.1   1   1
2.1   2   2
3.1   3   3
```

```

4.1  4  4
2.2  5  5
3.2  6  6
4.2  7  7
3.3  8  8
4.3  9  9
4.4 10 10

```

```

> M <- ltMatrices(matrix(1:6, nrow = 6, ncol = 2), diag = FALSE)
> Lower_tri(M, diag = FALSE)

```

```

      [,1] [,2]
2.1     1     1
3.1     2     2
4.1     3     3
3.2     4     4
4.2     5     5
4.3     6     6

```

```

> Lower_tri(M, diag = TRUE)

```

```

      [,1] [,2]
1.1     1     1
2.1     1     1
3.1     2     2
4.1     3     3
2.2     1     1
3.2     4     4
4.2     5     5
3.3     1     1
4.3     6     6
4.4     1     1

```

```

> ## multiple symmetric matrices
> Lower_tri(invchol2cor(M))

```

```

      [,1] [,2]
2.1 -0.7071 -0.7071
3.1  0.4364  0.4364
4.1 -0.4481 -0.4481
3.2 -0.9258 -0.9258
4.2  0.9189  0.9189
4.3 -0.9974 -0.9974

```

2.5 Diagonal Elements

The diagonal elements of each matrix \mathbf{C}_i can be extracted and are always returned as an $J \times N$ matrix.

< diagonals ltMatrices 19 > ≡

```
diagonals <- function(x, ...)
  UseMethod("diagonals")

diagonals.ltMatrices <- function(x, ...) {
  < extract slots 10 >

  x <- unclass(x)

  if (!diag) {
    ret <- matrix(1, nrow = J, ncol = ncol(x))
    colnames(ret) <- dn[[1L]]
    rownames(ret) <- dn[[2L]]
    return(ret)
  } else {
    if (J == 1L) return(x)
    if (byrow)
      idx <- cumsum(c(1, 2:J))
    else
      idx <- cumsum(c(1, J:2))
    ret <- x[idx, , drop = FALSE]
    rownames(ret) <- dn[[2L]]
    return(ret)
  }
}

diagonals.syMatrices <- diagonals.ltMatrices

diagonals.matrix <- function(x, ...) diag(x)
◇
```

Fragment referenced in 2.

```
> all(diagonals(ltMatrices(xn, byrow = TRUE)) == 1L)
[1] TRUE
```

Sometimes we need to add diagonal elements to an `ltMatrices` object which was set-up with constant $c_{jj} = 1$ diagonal elements.

< add diagonal elements 20 > ≡

```
.adddiag <- function(x) {  
  stopifnot(is.ltMatrices(x))  
  if (attr(x, "diag")) return(x)  
  byrow_orig <- attr(x, "byrow")  
  x <- ltMatrices(x, byrow = FALSE)  
  N <- dim(x)[1L]  
  J <- dim(x)[2L]  
  nm <- dimnames(x)[[2L]]  
  L <- diag(J)  
  L[lower.tri(L, diag = TRUE)] <- 1:(J * (J + 1) / 2)  
  D <- diag(J)  
  ret <- matrix(D[lower.tri(D, diag = TRUE)],  
               nrow = J * (J + 1) / 2, ncol = N)  
  colnames(ret) <- dimnames(x)[[1L]]  
  ret[L[lower.tri(L, diag = FALSE)],] <- unclass(x)  
  ret <- ltMatrices(ret, diag = TRUE, byrow = FALSE, names = nm)  
  ret <- ltMatrices(ret, byrow = byrow_orig)  
  ret  
}
```

◇
Fragment referenced in [2](#).

< assign diagonal elements 21 > ≡

```
"diagonals<-" <- function(x, value)
  UseMethod("diagonals<-")

"diagonals<-.ltMatrices" <- function(x, value) {
  < extract slots 10 >

  if (byrow)
    idx <- cumsum(c(1, 2:J))
  else
    idx <- cumsum(c(1, J:2))

  ### diagonals(x) <- NULL returns ltMatrices(..., diag = FALSE)
  if (is.null(value)) {
    if (!attr(x, "diag")) return(x)
    if (J == 1L) {
      x[] <- 1
      return(x)
    }
    return(ltMatrices(unclass(x)[-idx,,drop = FALSE], diag = FALSE,
                      byrow = byrow, names = dn[[2L]]))
  }

  x <- .adddiag(x)

  if (!is.matrix(value))
    value <- matrix(value, nrow = J, ncol = d[1L])

  stopifnot(is.matrix(value) && nrow(value) == J
            && ncol(value) == d[1L])

  if (J == 1L) {
    x[] <- value
    return(x)
  }

  x[idx, ] <- value

  return(x)
}

"diagonals<-.syMatrices" <- function(x, value) {
  x <- as.ltMatrices(x)
  diagonals(x) <- value
  class(x)[1L] <- "syMatrices"

  return(x)
}
◇
```

Fragment referenced in [2](#).

```
> lxd2 <- lxn
> diagonals(lxd2) <- 1
> chk(as.array(lxd2), as.array(lxn))
```

A unit diagonal matrix is not treated as a special case but as an `ltMatrices` object with all lower triangular elements being zero

`<diagonal matrix 22> ≡`

```
diagonals.integer <- function(x, ...)
  ltMatrices(rep(0, x * (x - 1) / 2), diag = FALSE, ...)
```

◇

Fragment referenced in [2](#).

```
> (I5 <- diagonals(5L))
```

```
, , 1
     1 2 3 4 5
1 1 . . . .
2 . 1 . . .
3 . . 1 . .
4 . . . 1 .
5 . . . . 1
```

```
> diagonals(I5) <- 1:5
> I5
```

```
, , 1
     1 2 3 4 5
1 1 . . . .
2 . 2 . . .
3 . . 3 . .
4 . . . 4 .
5 . . . . 5
```

2.6 Multiplication

Products $\mathbf{C}_i \mathbf{y}_i$ or $\mathbf{C}_i^\top \mathbf{y}_i$ with $\mathbf{y}_i \in \mathbb{R}^J$ for $i = 1, \dots, N$ can be computed with `y` being an $J \times N$ matrix of columns-wise stacked vectors ($\mathbf{y}_1 \mid \mathbf{y}_2 \mid \dots \mid \mathbf{y}_N$). If `y` is a single vector, it is recycled N times.

If the number of columns of a matrix `y` is neither one nor N , we compute $\mathbf{C}_i \mathbf{y}_j$ for all $i = 1, \dots, N$ and j . This is dangerous but needed in [Section 2.13](#) for defining `cond_mvnorm` later on.

For $\mathbf{C}_i \mathbf{y}_i$, we call C code computing the product efficiently without copying data by leveraging the lower triangular structure of $\mathbf{x} = \mathbf{C}_i$

< mult ltMatrices 23a > ≡

```
### C %*% y
Mult <- function(x, y, ...)
  UseMethod("Mult")
Mult.default <- function(x, y, transpose = FALSE, ...) {
  if (!transpose) return(x %*% y)
  return(crossprod(x, y))
}
Mult.ltMatrices <- function(x, y, transpose = FALSE, ...) {

  < extract slots 10 >

  stopifnot(is.numeric(y))
  if (!is.matrix(y)) y <- matrix(y, nrow = d[2L], ncol = d[1L])
  N <- ifelse(d[1L] == 1, ncol(y), d[1L])
  stopifnot(nrow(y) == d[2L])
  if (ncol(y) != N)
    return(sapply(1:ncol(y), function(i) Mult(x, y[,i], transpose = transpose)))

  < mult ltMatrices transpose 25 >

  x <- ltMatrices(x, byrow = TRUE)
  if (!is.double(x)) storage.mode(x) <- "double"
  if (!is.double(y)) storage.mode(y) <- "double"

  ret <- .Call(mvtnorm_R_ltMatrices_Mult, x, y, as.integer(N),
              as.integer(d[2L]), as.logical(diag))

  rownames(ret) <- dn[[2L]]
  if (length(dn[[1L]]) == N)
    colnames(ret) <- dn[[1L]]
  return(ret)
}
◇
```

Fragment referenced in 2.

The underlying C code assumes C_i (here called C) to be in row-major order.

< RC input 23b > ≡

```
/* pointer to C matrices */
double *dC = REAL(C);
/* number of matrices */
int iN = INTEGER(N)[0];
/* dimension of matrices */
int iJ = INTEGER(J)[0];
/* C contains diagonal elements */
Rboolean Rdiag = asLogical(diag);
/* p = J * (J - 1) / 2 + diag * J */
int len = iJ * (iJ - 1) / 2 + Rdiag * iJ;
◇
```

Fragment referenced in 24b, 26, 29, 30, 33a, 36, 44a.

We also allow C_i to be constant (N is then determined from $\text{ncol}(y)$). The following fragment ensures that we only loop over C_i if $\text{dim}(x)[1L] > 1$

$\langle C \text{ length } 24a \rangle \equiv$

```
int p;
if (LENGTH(C) == len)
  /* C is constant for i = 1, ..., N */
  p = 0;
else
  /* C contains C_1, ..., C_N */
  p = len;
◇
```

Fragment referenced in [24b](#), [26](#), [29](#), [33a](#), [44a](#).

The C workhorse is now

$\langle \text{mult } 24b \rangle \equiv$

```
SEXP R_ltMatrices_Mult (SEXP C, SEXP y, SEXP N, SEXP J, SEXP diag) {

  SEXP ans;
  double *dans, *dy = REAL(y);
  int i, j, k, start;

   $\langle RC \text{ input } 23b \rangle$ 
   $\langle C \text{ length } 24a \rangle$ 

  PROTECT(ans = allocMatrix(REALSXP, iJ, iN));
  dans = REAL(ans);

  for (i = 0; i < iN; i++) {
    start = 0;
    for (j = 0; j < iJ; j++) {
      dans[j] = 0.0;
      for (k = 0; k < j; k++)
        dans[j] += dC[start + k] * dy[k];
      if (Rdiag) {
        dans[j] += dC[start + j] * dy[j];
        start += j + 1;
      } else {
        dans[j] += dy[j];
        start += j;
      }
    }
    dC += p;
    dy += iJ;
    dans += iJ;
  }
  UNPROTECT(1);
  return(ans);
}
◇
```

Fragment referenced in [3](#).

Some checks for $C_i y_i$

```
> lxn <- ltMatrices(xn, byrow = TRUE)
> lxd <- ltMatrices(xd, byrow = TRUE, diag = TRUE)
```

```

> y <- matrix(runif(N * J), nrow = J)
> a <- Mult(lxn, y)
> A <- as.array(lxn)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(A[,i] %*% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> a <- Mult(lxd, y)
> A <- as.array(lxd)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(A[,i] %*% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> ### recycle C
> chk(Mult(lxn[rep(1, N),], y), Mult(lxn[,1], y), check.attributes = FALSE)
> ### recycle y
> chk(Mult(lxn, y[,1]), Mult(lxn, y[,rep(1, N)]))
> ### tcrossprod as multiplication
> i <- sample(1:N)[1]
> M <- t(as.array(lxn)[,i])
> a <- sapply(1:J, function(j) Mult(lxn[i,], M[,j,drop = FALSE]))
> rownames(a) <- colnames(a) <- dimnames(lxn)[[2L]]
> b <- as.array(Tcrossprod(lxn[i,]))[,1]
> chk(a, b, check.attributes = FALSE)

```

For $\mathbf{C}_i^\top \mathbf{y}_i$ (transpose = TRUE), we add a dedicated C function paying attention to the lower triangular structure of $\mathbf{x} = \mathbf{C}_i$. This function assumes \mathbf{x} in column-major order, so we coerce this object when necessary:

(mult ltMatrices transpose 25) ≡

```

if (transpose) {
  x <- ltMatrices(x, byrow = FALSE)
  if (!is.double(x)) storage.mode(x) <- "double"
  if (!is.double(y)) storage.mode(y) <- "double"

  ret <- .Call(mvtnorm_R_ltMatrices_Mult_transpose, x, y, as.integer(N),
              as.integer(d[2L]), as.logical(diag))

  rownames(ret) <- dn[[2L]]
  if (length(dn[[1L]]) == N)
    colnames(ret) <- dn[[1L]]
  return(ret)
}

```

Fragment referenced in [23a](#).

before moving to C for the low-level computations:

$\langle \text{mult transpose 26} \rangle \equiv$

```
SEXP R_ltMatrices_Mult_transpose (SEXP C, SEXP y, SEXP N, SEXP J, SEXP diag) {

    SEXP ans;
    double *dans, *dy = REAL(y);
    int i, j, k, start;

     $\langle RC \text{ input 23b} \rangle$ 
     $\langle C \text{ length 24a} \rangle$ 

    PROTECT(ans = allocMatrix(REALSXP, iJ, iN));
    dans = REAL(ans);

    for (i = 0; i < iN; i++) {
        start = 0;
        for (j = 0; j < iJ; j++) {
            dans[j] = 0.0;
            if (Rdiag) {
                dans[j] += dC[start] * dy[j];
                start++;
            } else {
                dans[j] += dy[j];
            }
            for (k = 0; k < (iJ - j - 1); k++)
                dans[j] += dC[start + k] * dy[j + k + 1];
            start += iJ - j - 1;
        }
        dC += p;
        dy += iJ;
        dans += iJ;
    }
    UNPROTECT(1);
    return(ans);
}
◇
```

Fragment referenced in 3.

and wrap-up with some tests for computing $\mathbf{C}_i^\top \mathbf{y}_i$

```
> a <- Mult(lxn, y, transpose = TRUE)
> A <- as.array(lxn)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(t(A[,i]) %*% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> a <- Mult(lxd, y, transpose = TRUE)
> A <- as.array(lxd)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(t(A[,i]) %*% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> ### recycle C
> chk(Mult(lxn[rep(1, N)], y, transpose = TRUE),
+   Mult(lxn[1,], y, transpose = TRUE), check.attributes = FALSE)
> ### recycle y
> chk(Mult(lxn, y[,1], transpose = TRUE),
+   Mult(lxn, y[,rep(1, N)], transpose = TRUE))
```

Now we can add a `Mult` method for multiple symmetric matrices, noting that for a symmetric matrix $\mathbf{A} = \mathbf{C} + \mathbf{C}^\top - \text{diag}(\mathbf{C})$ with lower triangular part \mathbf{C} (including the diagonal) we can compute $\mathbf{A}\mathbf{y} = \mathbf{C}\mathbf{y} + \mathbf{C}^\top\mathbf{y} - \text{diag}(\mathbf{C})\mathbf{y}$ using `Mult` applied to the lower triangular part:

`<mult syMatrices 27> ≡`

```
Mult.syMatrices <- function(x, y, ...) {
  < extract slots 10 >
  x <- as.ltMatrices(x)
  stopifnot(is.numeric(y))
  if (!is.matrix(y)) y <- matrix(y, nrow = d[2L], ncol = d[1L])
  N <- ifelse(d[1L] == 1, ncol(y), d[1L])
  stopifnot(nrow(y) == d[2L])
  stopifnot(ncol(y) == N)

  ret <- Mult(x, y) + Mult(x, y, transpose = TRUE) - y * c(diagonals(x))
  return(ret)
}
◇
```

Fragment referenced in 2.

```
> J <- 5
> N1 <- 10
> ex <- expression({
+   C <- syMatrices(matrix(runif(N2 * J * (J + c(-1, 1)[DIAG + 1L]) / 2),
+                         ncol = N2),
+                   diag = DIAG)
+   x <- matrix(runif(N1 * J), nrow = J)
+   Ca <- as.array(C)
+   p1 <- do.call("cbind", lapply(1:N1, function(i)
+     Ca[, , c(1, i)][(N2 > 1) + 1] %*% x[, i]))
+   p2 <- Mult(C, x)
+   chk(p1, p2)
+ })
> N2 <- N1
> DIAG <- TRUE
> eval(ex)
> N2 <- 1
> DIAG <- TRUE
> eval(ex)
> N2 <- 1
> DIAG <- FALSE
> eval(ex)
> N2 <- N1
> DIAG <- FALSE
> eval(ex)
```

2.7 Solving Linear Systems

Computing \mathbf{C}_i^{-1} or solving $\mathbf{C}_i\mathbf{x}_i = \mathbf{y}_i$ for \mathbf{x}_i for all $i = 1, \dots, N$ is another important task. We sometimes also need $\mathbf{C}_i^\top\mathbf{x}_i = \mathbf{y}_i$ triggered by `transpose = TRUE`.

\mathbf{C} is $\mathbf{C}_i, i = 1, \dots, N$ in column-major order (matrix of dimension $J(J-1)/2 + J \text{diag} \times N$), and \mathbf{y} is the $J \times N$ matrix $(\mathbf{y}_1 | \mathbf{y}_2 | \dots | \mathbf{y}_N)$. This function returns the $J \times N$ matrix $(\mathbf{x}_1 | \mathbf{x}_2 | \dots | \mathbf{x}_N)$ of solutions.

If \mathbf{y} is not given, \mathbf{C}_i^{-1} is returned in the same order as the original matrix \mathbf{C}_i . If all \mathbf{C}_i have unit diagonals, so will \mathbf{C}_i^{-1} .

We start with some options for the LAPACK workhorses

lapack options 28 \equiv

```

char di, lo = 'L';
if (Rdiag) {
    /* non-unit diagonal elements */
    di = 'N';
} else {
    /* unit diagonal elements; NOTE: these diagonals is ARE always present but
       ignored in the computations */
    di = 'U';
}

```

Fragment referenced in [29](#), [30](#).

and set-up a dedicated C function for computing $\mathbf{C}_i \mathbf{x}_i = \mathbf{y}_i$

$\langle solve\ 29 \rangle \equiv$

```
SEXP R_ltMatrices_solve (SEXP C, SEXP y, SEXP N, SEXP J, SEXP diag, SEXP transpose)
{
    SEXP ans;
    double *dans, *dy;
    int i, ONE = 1;

     $\langle RC\ input\ 23b \rangle$ 
    /* diagonal elements are always present */
    if (!Rdiag) len += iJ;
     $\langle C\ length\ 24a \rangle$ 
     $\langle lapack\ options\ 28 \rangle$ 

    char tr = 'N';
    /* t(C) instead of C */
    Rboolean Rtranspose = asLogical(transpose);
    if (Rtranspose) {
        /* t(C) */
        tr = 'T';
    } else {
        /* C */
        tr = 'N';
    }

    dy = REAL(y);
    PROTECT(ans = allocMatrix(REALSXP, iJ, iN));
    dans = REAL(ans);
    memcpy(dans, dy, iJ * iN * sizeof(double));

    /* loop over matrices, ie columns of C / y */
    for (i = 0; i < iN; i++) {

        /* solve linear system */
        F77_CALL(dtpsv)(&lo, &tr, &di, &iJ, dC, dans, &ONE FCONE FCONE FCONE);
        dans += iJ;
        dC += p;
    }

    UNPROTECT(1);
    return(ans);
}

```

Fragment referenced in 3.

and then for computing C_i^{-1} explicitly

< solve C 30 > ≡

```
SEXP R_ltMatrices_solve_C (SEXP C, SEXP N, SEXP J, SEXP diag, SEXP transpose)
{
    SEXP ans;
    double *dans;
    int i, info;

    < RC input 23b >
    /* diagonal elements are always present */
    if (!Rdiag) len += iJ;
    < lapack options 28 >

    PROTECT(ans = allocMatrix(REALSXP, len, iN));
    dans = REAL(ans);
    memcpy(dans, dC, iN * len * sizeof(double));

    /* loop over matrices, ie columns of C / y */
    for (i = 0; i < iN; i++) {

        /* compute inverse */
        F77_CALL(dtptri)(&lo, &di, &iJ, dans, &info FCONE FCONE);
        if (info != 0)
            error("Cannot solve ltmatrices");

        dans += len;
    }

    UNPROTECT(1);
    /* note: ans always includes diagonal elements */
    return(ans);
}
◇
```

Fragment referenced in [3](#).

with R interface

< solve ltMatrices 31 > ≡

```
solve.ltMatrices <- function(a, b, transpose = FALSE, ...) {  
  
  byrow_orig <- attr(a, "byrow")  
  
  x <- ltMatrices(a, byrow = FALSE)  
  diag <- attr(x, "diag")  
  ### dtptri and dtpsv require diagonal elements being present  
  if (!diag) diagonals(x) <- diagonals(x)  
  d <- dim(x)  
  J <- d[2L]  
  dn <- dimnames(x)  
  if (!is.double(x)) storage.mode(x) <- "double"  
  
  if (!missing(b)) {  
    if (!is.matrix(b)) b <- matrix(b, nrow = J, ncol = d[1L])  
    stopifnot(nrow(b) == J)  
    N <- ifelse(d[1L] == 1, ncol(b), d[1L])  
    stopifnot(ncol(b) == N)  
    if (!is.double(b)) storage.mode(b) <- "double"  
    ret <- .Call(mvtnorm_R_ltMatrices_solve, x, b,  
                as.integer(N), as.integer(J), as.logical(diag),  
                as.logical(transpose))  
    if (d[1L] == N) {  
      colnames(ret) <- dn[[1L]]  
    } else {  
      colnames(ret) <- colnames(b)  
    }  
    rownames(ret) <- dn[[2L]]  
    return(ret)  
  }  
  
  if (transpose) stop("cannot compute inverse of t(a)")  
  ret <- .Call(mvtnorm_R_ltMatrices_solve_C, x,  
              as.integer(d[1L]), as.integer(J), as.logical(diag),  
              as.logical(FALSE))  
  colnames(ret) <- dn[[1L]]  
  
  if (!diag)  
    ### ret always includes diagonal elements, remove here  
    ret <- ret[- cumsum(c(1, J:2)), , drop = FALSE]  
  
  ret <- ltMatrices(ret, diag = diag, byrow = FALSE, names = dn[[2L]])  
  ret <- ltMatrices(ret, byrow = byrow_orig)  
  return(ret)  
}  
◇
```

Fragment referenced in [2](#).

and some checks

```
> ## solve  
> A <- as.array(1xn)  
> a <- solve(1xn)  
> a <- as.array(a)  
> b <- array(apply(A, 3L, function(x) solve(x), simplify = TRUE),
```

```

+           dim = rev(dim(lxn)))
> chk(a, b, check.attributes = FALSE)
> A <- as.array(lxd)
> a <- as.array(solve(lxd))
> b <- array(apply(A, 3L, function(x) solve(x), simplify = TRUE),
+           dim = rev(dim(lxd)))
> chk(a, b, check.attributes = FALSE)
> chk(solve(lxn, y), Mult(solve(lxn), y))
> chk(solve(lxd, y), Mult(solve(lxd), y))
> ### recycle C
> chk(solve(lxn[1,], y), as.array(solve(lxn[1,]))[,1] %*% y)
> chk(solve(lxn[rep(1, N),], y), solve(lxn[1,], y), check.attributes = FALSE)
> ### recycle y
> chk(solve(lxn, y[,1]), solve(lxn, y[,rep(1, N)]))

    also for  $\mathbf{C}_i^\top \mathbf{x}_i = \mathbf{y}_i$ 

> chk(solve(lxn[1,], y, transpose = TRUE),
+     t(as.array(solve(lxn[1,]))[,1]) %*% y)

```

2.8 Log-determinants

For computing the log-determinant $\log(\det(\mathbf{C}_i)) = \sum_{j=1}^J \log(\text{diag}(\mathbf{C}_i)_j)$ we sum over the log-diagonal entries of a lower triangular matrix in \mathbf{C} , both when the data are stored in row- and column-major order:

logdet 33a ≡

```
SEXP R_ltMatrices_logdet (SEXP C, SEXP N, SEXP J, SEXP diag, SEXP byrow) {

  SEXP ans;
  double *dans;
  int i, j, k;

  < RC input 23b >
  Rboolean Rbyrow = asLogical(byrow);
  < C length 24a >

  PROTECT(ans = allocVector(REALSXP, iN));
  dans = REAL(ans);

  for (i = 0; i < iN; i++) {
    dans[i] = 0.0;
    if (Rdiag) {
      k = 1;
      for (j = 0; j < iJ; j++) {
        dans[i] += log(dC[k - 1]);
        k += (Rbyrow ? j + 2 : iJ - j);
      }
      dC += p;
    }
  }

  UNPROTECT(1);
  return(ans);
}
◇
```

Fragment referenced in 3.

The R interface now simply calls this low-level function

logdet ltMatrices 33b ≡

```
logdet <- function(x) {

  if (!is.ltMatrices(x))
    stop("x is not an ltMatrices object")

  byrow <- attr(x, "byrow")
  diag <- attr(x, "diag")
  d <- dim(x)
  J <- d[2L]
  dn <- dimnames(x)
  if (!is.double(x)) storage.mode(x) <- "double"

  ret <- .Call(mvtnorm_R_ltMatrices_logdet, x,
              as.integer(d[1L]), as.integer(J), as.logical(diag),
              as.logical(byrow))
  names(ret) <- dn[[1L]]
  return(ret)
}
◇
```

Fragment referenced in 2.

We test the functionality by extracting the diagonal elements from different matrices and summing over their logarithms

```
> chk(logdet(lxn), colSums(log(diagonals(lxn))))
> chk(logdet(lxd[1,]), colSums(log(diagonals(lxd[1,]))))
> chk(logdet(lxd), colSums(log(diagonals(lxd))))
> lxd2 <- ltMatrices(lxd, byrow = !attr(lxd, "byrow"))
> chk(logdet(lxd2), colSums(log(diagonals(lxd2))))
```

2.9 Crossproducts

We want to compute $\mathbf{C}_i \mathbf{C}_i^\top$ or $\text{diag}(\mathbf{C}_i \mathbf{C}_i^\top)$ (`diag_only = TRUE`) for $i = 1, \dots, N$. These are symmetric matrices, so we store them as a lower triangular matrix using a different class name `syMatrices`. We write one C function for computing $\mathbf{C}_i \mathbf{C}_i^\top$ or $\mathbf{C}_i^\top \mathbf{C}_i$ (`Rtranspose` being `TRUE`).

We differentiate between computation of the diagonal elements of the crossproduct

<first element 34a> \equiv

```
    dans[0] = 1.0;
    if (Rdiag)
        dans[0] = pow(dC[0], 2);
    if (Rtranspose) { // crossprod
        for (k = 1; k < iJ; k++)
            dans[0] += pow(dC[IDX(k + 1, 1, iJ, Rdiag)], 2);
    }
    ◇
```

Fragment referenced in 34b, 35a.

<tcrossprod diagonal only 34b> \equiv

```
    PROTECT(ans = allocMatrix(REALSXP, iJ, iN));
    dans = REAL(ans);
    for (n = 0; n < iN; n++) {
        <first element 34a>
        for (i = 1; i < iJ; i++) {
            dans[i] = 0.0;
            if (Rtranspose) { // crossprod
                for (k = i + 1; k < iJ; k++)
                    dans[i] += pow(dC[IDX(k + 1, i + 1, iJ, Rdiag)], 2);
            } else { // tcrossprod
                for (k = 0; k < i; k++)
                    dans[i] += pow(dC[IDX(i + 1, k + 1, iJ, Rdiag)], 2);
            }
            if (Rdiag) {
                dans[i] += pow(dC[IDX(i + 1, i + 1, iJ, Rdiag)], 2);
            } else {
                dans[i] += 1.0;
            }
        }
        dans += iJ;
        dC += len;
    }
    ◇
```

Fragment referenced in 36.

and computation of the full $J \times J$ crossproduct matrix

$\langle \text{tcrossprod full 35a} \rangle \equiv$

```

nrow = iJ * (iJ + 1) / 2;
PROTECT(ans = allocMatrix(REALSXP, nrow, iN));
dans = REAL(ans);
for (n = 0; n < INTEGER(N)[0]; n++) {
   $\langle \text{first element 34a} \rangle$ 
  for (i = 1; i < iJ; i++) {
    for (j = 0; j <= i; j++) {
      ix = IDX(i + 1, j + 1, iJ, 1);
      dans[ix] = 0.0;
      if (Rtranspose) { // crossprod
        for (k = i + 1; k < iJ; k++)
          dans[ix] +=
            dC[IDX(k + 1, i + 1, iJ, Rdiag)] *
            dC[IDX(k + 1, j + 1, iJ, Rdiag)];
      } else { // tcrossprod
        for (k = 0; k < j; k++)
          dans[ix] +=
            dC[IDX(i + 1, k + 1, iJ, Rdiag)] *
            dC[IDX(j + 1, k + 1, iJ, Rdiag)];
      }
      if (Rdiag) {
        if (Rtranspose) {
          dans[ix] +=
            dC[IDX(i + 1, i + 1, iJ, Rdiag)] *
            dC[IDX(i + 1, j + 1, iJ, Rdiag)];
        } else {
          dans[ix] +=
            dC[IDX(i + 1, j + 1, iJ, Rdiag)] *
            dC[IDX(j + 1, j + 1, iJ, Rdiag)];
        }
      } else {
        if (j < i)
          dans[ix] += dC[IDX(i + 1, j + 1, iJ, Rdiag)];
        else
          dans[ix] += 1.0;
      }
    }
  }
  dans += nrow;
  dC += len;
}

```

Fragment referenced in 36.

and put both cases together

$\langle \text{IDX 35b} \rangle \equiv$

```

#define IDX(i, j, n, d) ((i) >= (j) ? (n) * ((j) - 1) - ((j) - 2) * ((j) - 1)/2 + (i) - (j) - (!d) * (

```

Fragment referenced in 36, 44a.

$\langle \text{tcrossprod } 36 \rangle \equiv$

$\langle \text{IDX } 35b \rangle$

```
SEXP R_ltMatrices_tcrossprod (SEXP C, SEXP N, SEXP J, SEXP diag,  
                             SEXP diag_only, SEXP transpose) {
```

```
    SEXP ans;  
    double *dans;  
    int i, j, n, k, ix, nrow;
```

$\langle \text{RC input } 23b \rangle$

```
Rboolean Rdiag_only = asLogical(diag_only);  
Rboolean Rtranspose = asLogical(transpose);
```

```
if (Rdiag_only) {  
     $\langle \text{tcrossprod diagonal only } 34b \rangle$   
} else {  
     $\langle \text{tcrossprod full } 35a \rangle$   
}  
UNPROTECT(1);  
return(ans);
```

```
}  
◇
```

Fragment referenced in 3.

with R interface

< tcrossprod ltMatrices 37 > ≡

```
### C %*% t(C) => returns object of class syMatrices
### diag(C %*% t(C)) => returns matrix of diagonal elements
.Tcrossprod <- function(x, diag_only = FALSE, transpose = FALSE) {

  if (!is.ltMatrices(x)) {
    ret <- tcrossprod(x)
    if (diag_only) ret <- diag(ret)
    return(ret)
  }

  byrow_orig <- attr(x, "byrow")
  diag <- attr(x, "diag")
  d <- dim(x)
  N <- d[1L]
  J <- d[2L]
  dn <- dimnames(x)

  x <- ltMatrices(x, byrow = FALSE)
  if (!is.double(x)) storage.mode(x) <- "double"

  ret <- .Call(mvtnorm_R_ltMatrices_tcrossprod, x, as.integer(N), as.integer(J),
              as.logical(diag), as.logical(diag_only), as.logical(transpose))
  colnames(ret) <- dn[[1L]]
  if (diag_only) {
    rownames(ret) <- dn[[2L]]
  } else {
    ret <- ltMatrices(ret, diag = TRUE, byrow = FALSE, names = dn[[2L]])
    ret <- as.syMatrices(ltMatrices(ret, byrow = byrow_orig))
  }
  return(ret)
}
.Tcrossprod <- function(x, diag_only = FALSE)
  .Tcrossprod(x = x, diag_only = diag_only, transpose = FALSE)
◇
```

Fragment referenced in [2](#).

We could have created yet another generic `tcrossprod`, but `base::tcrossprod` is more general and, because speed is an issue, we don't want to waste time on methods dispatch.

```
> ## Tcrossprod
> a <- as.array(Tcrossprod(lxn))
> b <- array(apply(as.array(lxn), 3L, function(x) tcrossprod(x), simplify = TRUE),
+           dim = rev(dim(lxn)))
> chk(a, b, check.attributes = FALSE)
> # diagonal elements only
> d <- Tcrossprod(lxn, diag_only = TRUE)
> chk(d, apply(a, 3, diag))
> chk(d, diagonals(Tcrossprod(lxn)))
> a <- as.array(Tcrossprod(lxd))
> b <- array(apply(as.array(lxd), 3L, function(x) tcrossprod(x), simplify = TRUE),
+           dim = rev(dim(lxd)))
> chk(a, b, check.attributes = FALSE)
> # diagonal elements only
> d <- Tcrossprod(lxd, diag_only = TRUE)
```

```
> chk(d, apply(a, 3, diag))
> chk(d, diagonals(Tcrossprod(lxd)))
```

We also add `Crossprod`, which is a call to `Tcrossprod` with the `transpose` switch turned on

(*crossprod* *ltMatrices* 38) \equiv

```
Crossprod <- function(x, diag_only = FALSE)
  .Tcrossprod(x, diag_only = diag_only, transpose = TRUE)
```

◇

Fragment referenced in [2](#).

and run some checks

```
> ## Crossprod
> a <- as.array(Crossprod(lxn))
> b <- array(apply(as.array(lxn), 3L, function(x) crossprod(x), simplify = TRUE),
+           dim = rev(dim(lxn)))
> chk(a, b, check.attributes = FALSE)
> # diagonal elements only
> d <- Crossprod(lxn, diag_only = TRUE)
> chk(d, apply(a, 3, diag))
> chk(d, diagonals(Crossprod(lxn)))
> a <- as.array(Crossprod(lxd))
> b <- array(apply(as.array(lxd), 3L, function(x) crossprod(x), simplify = TRUE),
+           dim = rev(dim(lxd)))
> chk(a, b, check.attributes = FALSE)
> # diagonal elements only
> d <- Crossprod(lxd, diag_only = TRUE)
> chk(d, apply(a, 3, diag))
> chk(d, diagonals(Crossprod(lxd)))
```

We now also add methods implementing a standard user interface

< crossprod tcrossprod methods 39 > ≡

```
crossprod.ltMatrices <- function(x, y = NULL, ...) {
  if (is.null(y))
    return(Crossprod(x = x))
  return(Mult(x, y, transpose = TRUE))
}

crossprod.syMatrices <- crossprod.ltMatrices

tcrossprod.ltMatrices <- function(x, y = NULL, ...) {
  if (is.null(y))
    return(Tcrossprod(x = x))
  return(Mult(x, y, transpose = FALSE))
}

tcrossprod.syMatrices <- tcrossprod.ltMatrices

'%*%.ltMatrices' <- function(x, y)
  Mult(x, y)

'%*%.syMatrices' <- function(x, y)
  Mult(x, y)
◇
```

Fragment referenced in 2.

which allows coding closer to the standard

```
> ### tcrossprod
> a <- as.array(tcrossprod(lxn))
> b <- array(apply(as.array(lxn), 3L, function(x) tcrossprod(x), simplify = TRUE),
+           dim = rev(dim(lxn)))
> chk(a, b, check.attributes = FALSE)
> a <- as.array(tcrossprod(lxd))
> b <- array(apply(as.array(lxd), 3L, function(x) tcrossprod(x), simplify = TRUE),
+           dim = rev(dim(lxd)))
> chk(a, b, check.attributes = FALSE)
> ## crossprod
> a <- as.array(crossprod(lxn))
> b <- array(apply(as.array(lxn), 3L, function(x) crossprod(x), simplify = TRUE),
+           dim = rev(dim(lxn)))
> chk(a, b, check.attributes = FALSE)
> a <- as.array(crossprod(lxd))
> b <- array(apply(as.array(lxd), 3L, function(x) crossprod(x), simplify = TRUE),
+           dim = rev(dim(lxd)))
> chk(a, b, check.attributes = FALSE)
```

Multiplication also works in this simple way

```
> a <- lxn %*% y
> A <- as.array(lxn)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(A[,i] %*% y[,i,drop = FALSE])))
```

```

> chk(a, t(b), check.attributes = FALSE)
> a <- lxd %**% y
> A <- as.array(lxd)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(A[,i] %**% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> ### recycle C
> chk(lxn[rep(1, N),] %**% y, lxn[1,] %**% y, check.attributes = FALSE)
> ### recycle y
> chk(lxn %**% y[,1], lxn %**% y[,rep(1, N)])
> ### tcrossprod as multiplication
> i <- sample(1:N)[1]
> M <- t(as.array(lxn)[,i])
> a <- sapply(1:J, function(j) lxn[i,] %**% M[j,drop = FALSE])
> rownames(a) <- colnames(a) <- dimnames(lxn)[[2L]]
> b <- as.array(tcrossprod(lxn[i,]))[,1]
> chk(a, b, check.attributes = FALSE)
> a <- crossprod(lxn, y)
> A <- as.array(lxn)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(t(A[,i]) %**% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> a <- crossprod(lxd, y)
> A <- as.array(lxd)
> b <- do.call("rbind", lapply(1:ncol(y),
+   function(i) t(t(A[,i]) %**% y[,i,drop = FALSE])))
> chk(a, t(b), check.attributes = FALSE)
> ### recycle C
> chk(crossprod(lxn[rep(1, N),], y),
+   crossprod(lxn[1,], y), check.attributes = FALSE)
> ### recycle y
> chk(crossprod(lxn, y[,1]),
+   crossprod(lxn, y[,rep(1, N)]))

```

2.10 Cholesky Factorisation

One might want to compute the Cholesky factorisations $\Sigma_i = \mathbf{C}_i \mathbf{C}_i^\top$ for multiple symmetric matrices Σ_i , stored as a matrix in class `syMatrices`.

< chol syMatrices 41 > ≡

```
chol.syMatrices <- function(x, ...) {  
  
  byrow_orig <- attr(x, "byrow")  
  dnm <- dimnames(x)  
  stopifnot(attr(x, "diag"))  
  d <- dim(x)  
  
  ### x is of class syMatrices, coerce to ltMatrices first and re-arrange  
  ### second  
  x <- ltMatrices(unclass(x), diag = TRUE,  
                 byrow = byrow_orig, names = dnm[[2L]])  
  x <- ltMatrices(x, byrow = FALSE)  
  # class(x) <- class(x)[-1]  
  if (!is.double(x)) storage.mode(x) <- "double"  
  
  ret <- .Call(mvtnorm_R_syMatrices_chol, x,  
              as.integer(d[1L]), as.integer(d[2L]))  
  colnames(ret) <- dnm[[1L]]  
  
  ret <- ltMatrices(ret, diag = TRUE,  
                   byrow = FALSE, names = dnm[[2L]])  
  ret <- ltMatrices(ret, byrow = byrow_orig)  
  
  return(ret)  
}  
◇
```

Fragment referenced in 2.

Luckily, we already have the data in the correct packed column-major storage, so we swiftly loop over $i = 1, \dots, N$ in C and hand over to LAPACK

$\langle chol\ 42 \rangle \equiv$

```
SEXP R_syMatrices_chol (SEXP Sigma, SEXP N, SEXP J) {

    SEXP ans;
    double *dans, *dSigma;
    int iJ = INTEGER(J)[0];
    int pJ = iJ * (iJ + 1) / 2;
    int iN = INTEGER(N)[0];
    int i, j, info = 0;
    char lo = 'L';

    PROTECT(ans = allocMatrix(REALSXP, pJ, iN));
    dans = REAL(ans);
    dSigma = REAL(Sigma);

    for (i = 0; i < iN; i++) {

        /* copy data */
        for (j = 0; j < pJ; j++)
            dans[j] = dSigma[j];

        F77_CALL(dpptrf)(&lo, &iJ, dans, &info FCONE);

        if (info != 0) {
            if (info > 0)
                error("the leading minor of order %d is not positive definite",
                    info);
            error("argument %d of Lapack routine %s had invalid value",
                -info, "dpptrf");
        }

        dSigma += pJ;
        dans += pJ;
    }
    UNPROTECT(1);
    return(ans);
}
◇
```

Fragment referenced in 3.

This new chol method can be used to revert Tcrossprod for ltMatrices with and without unit diagonals:

```
> Sigma <- tcrossprod(lxd)
> chk(chol(Sigma), lxd)
> Sigma <- tcrossprod(lxn)
> ## Sigma and chol(Sigma) always have diagonal, lxn doesn't
> chk(as.array(chol(Sigma)), as.array(lxn))
```

2.11 Kronecker Products

We sometimes need to compute $\text{vec}(\mathbf{S})^\top (\mathbf{A}^\top \otimes \mathbf{C})$, where \mathbf{S} is a lower triangular or other $J \times J$ matrix and \mathbf{A} and \mathbf{C} are lower triangular $J \times J$ matrices. With the “vec trick”, we have $\text{vec}(\mathbf{S})^\top (\mathbf{A}^\top \otimes \mathbf{C}) = \text{vec}(\mathbf{C}^\top \mathbf{S} \mathbf{A}^\top)^\top$. The LAPACK function `dtrmm` computes products of lower triangular matrices with other matrices, so we simply call this function looping over $i = 1, \dots, N$.

$\langle t(C) S t(A) 43 \rangle \equiv$

```
char siR = 'R', siL = 'L', lo = 'L', tr = 'N', trT = 'T', di = 'N', trs;
double ONE = 1.0;
int iJ2 = iJ * iJ;

double tmp[iJ2];
for (j = 0; j < iJ2; j++) tmp[j] = 0.0;

ans = PROTECT(allocMatrix(REALSXP, iJ2, iN));
dans = REAL(ans);

for (i = 0; i < LENGTH(ans); i++) dans[i] = 0.0;

for (i = 0; i < iN; i++) {

    /* A := C */
    for (j = 0; j < iJ; j++) {
        for (k = 0; k <= j; k++)
            tmp[k * iJ + j] = dC[IDX(j + 1, k + 1, iJ, 1L)];
    }

    /* S was already expanded in R code; B = S */
    for (j = 0; j < iJ2; j++) dans[j] = dS[j];

    /* B := t(A) %*% B */
    trs = (RtC ? trT : tr);
    F77_CALL(dtrmm)(&siL, &lo, &trs, &di, &iJ, &iJ, &ONE, tmp, &iJ,
        dans, &iJ FCONE FCONE FCONE FCONE);

    /* A */
    for (j = 0; j < iJ; j++) {
        for (k = 0; k <= j; k++)
            tmp[k * iJ + j] = dA[IDX(j + 1, k + 1, iJ, 1L)];
    }

    /* B := B %*% t(A) */
    trs = (RtA ? trT : tr);
    F77_CALL(dtrmm)(&siR, &lo, &trs, &di, &iJ, &iJ, &ONE, tmp, &iJ,
        dans, &iJ FCONE FCONE FCONE FCONE);

    dans += iJ2;
    dC += p;
    dS += iJ2;
    dA += p;
}
◇
```

Fragment referenced in [44a](#).

< vec trick 44a > \equiv

< IDX 35b >

```
SEXP R_vectrick(SEXP C, SEXP N, SEXP J, SEXP S, SEXP A, SEXP diag, SEXP trans) {  
  
    int i, j, k;  
    SEXP ans;  
    double *dS, *dans, *dA;  
  
    /* note: diag is needed by this chunk but has no consequences */  
    < RC input 23b >  
    < C length 24a >  
    dS = REAL(S);  
    dA = REAL(A);  
  
    Rboolean RtC = LOGICAL(trans)[0];  
    Rboolean RtA = LOGICAL(trans)[1];  
  
    < t(C) S t(A) 43 >  
  
    UNPROTECT(1);  
    return(ans);  
}  
◇
```

Fragment referenced in 3.

In R, we compute $\mathbf{C}^\top \mathbf{S} \mathbf{A}^\top$ by default or $\mathbf{C} \mathbf{S} \mathbf{A}^\top$ or $\mathbf{C}^\top \mathbf{S} \mathbf{A}$ or $\mathbf{C}^\top \mathbf{S} \mathbf{A}^\top$ by using the `trans` argument in `vectrick`. Argument `C` is an `ltMatrices` object

< check C argument 44b > \equiv

```
C <- as.ltMatrices(C)  
if (!attr(C, "diag")) diagonals(C) <- 1  
C_byrow_orig <- attr(C, "byrow")  
C <- ltMatrices(C, byrow = FALSE)  
dC <- dim(C)  
nm <- attr(C, "rcnames")  
N <- dC[1L]  
J <- dC[2L]  
class(C) <- class(C)[-1L]   ### works because of as.ltMatrices(c)  
if (!is.double(C)) storage.mode(C) <- "double"  
◇
```

Fragment referenced in 46.

`S` can be an `ltMatrices` object or a $J^2 \times N$ matrix featuring columns of vectorised $J \times J$ matrices

< check S argument 45a > ≡

```
SltM <- is.ltMatrices(S)
if (SltM) {
  if (!attr(S, "diag")) diagonals(S) <- 1
  S_byrow_orig <- attr(S, "byrow")
  stopifnot(S_byrow_orig == C_byrow_orig)
  S <- ltMatrices(S, byrow = FALSE)
  dS <- dim(S)
  stopifnot(dC[2L] == dS[2L])
  if (dC[1] != 1L) {
    stopifnot(dC[1L] == dS[1L])
  } else {
    N <- dS[1L]
  }
  ## argument A in dtrmm is not in packed form, so expand in J x J
  ## matrix
  S <- matrix(as.array(S), ncol = dS[1L])
} else {
  stopifnot(is.matrix(S))
  stopifnot(nrow(S) == J^2)
  if (dC[1] != 1L) {
    stopifnot(dC[1L] == ncol(S))
  } else {
    N <- ncol(S)
  }
}
if (!is.double(S)) storage.mode(S) <- "double"
◇
```

Fragment referenced in [46](#).

A is an ltMatrices object

< check A argument 45b > ≡

```
if (missing(A)) {
  A <- C
} else {
  A <- as.ltMatrices(A)
  if (!attr(A, "diag")) diagonals(A) <- 1
  A_byrow_orig <- attr(A, "byrow")
  stopifnot(C_byrow_orig == A_byrow_orig)
  A <- ltMatrices(A, byrow = FALSE)
  dA <- dim(A)
  stopifnot(dC[2L] == dA[2L])
  class(A) <- class(A)[-1L]
  if (!is.double(A)) storage.mode(A) <- "double"
  if (dC[1L] != dA[1L]) {
    if (dC[1L] == 1L)
      C <- C[, rep(1, N), drop = FALSE]
    if (dA[1L] == 1L)
      A <- A[, rep(1, N), drop = FALSE]
    stopifnot(ncol(A) == ncol(C))
  }
}
◇
```

Fragment referenced in [46](#).

We put everything together in function `vecrick`

<kronecker vec trick 46> \equiv

```
vecrick <- function(C, S, A, transpose = c(TRUE, TRUE)) {  
  
  stopifnot(all(is.logical(transpose)))  
  stopifnot(length(transpose) == 2L)  
  
  < check C argument 44b >  
  < check S argument 45a >  
  < check A argument 45b >  
  
  ret <- .Call(mvtnorm_R_vecrick, C, as.integer(N), as.integer(J), S, A,  
             as.logical(TRUE), as.logical(transpose))  
  
  if (!SltM) return(matrix(c(ret), ncol = N))  
  
  L <- matrix(1:(J^2), nrow = J)  
  ret <- ltMatrices(ret[L[lower.tri(L, diag = TRUE)],,drop = FALSE],  
                  diag = TRUE, byrow = FALSE, names = nm)  
  ret <- ltMatrices(ret, byrow = C_byrow_orig)  
  return(ret)  
}  
◇
```

Fragment referenced in [2](#).

Here is a small example

```
> J <- 10  
> d <- TRUE  
> L <- diag(J)  
> L[lower.tri(L, diag = d)] <- prm <- runif(J * (J + c(-1, 1)[d + 1]) / 2)  
> C <- solve(L)  
> D <- -kronecker(t(C), C)  
> S <- diag(J)  
> S[lower.tri(S, diag = TRUE)] <- x <- runif(J * (J + 1) / 2)  
> SD0 <- matrix(c(S) %*% D, ncol = J)  
> SD1 <- -crossprod(C, tcrossprod(S, C))  
> a <- ltMatrices(C[lower.tri(C, diag = TRUE)], diag = TRUE, byrow = FALSE)  
> b <- ltMatrices(x, diag = TRUE, byrow = FALSE)  
> SD2 <- -vecrick(a, b, a)  
> SD2a <- -vecrick(a, b)  
> chk(SD2, SD2a)  
> chk(SD0[lower.tri(SD0, diag = d)],  
+     SD1[lower.tri(SD1, diag = d)])  
> chk(SD0[lower.tri(SD0, diag = d)],  
+     c(unclass(SD2)))  
> ### same; but SD2 is vec(SD0)  
> S <- t(matrix(as.array(b), byrow = FALSE, nrow = 1))  
> SD2 <- -vecrick(a, S, a)  
> SD2a <- -vecrick(a, S)  
> chk(SD2, SD2a)  
> chk(c(SD0), c(SD2))  
> ### N > 1
```

```

> N <- 4L
> prm <- runif(J * (J - 1) / 2)
> C <- ltMatrices(prm)
> S <- matrix(runif(J^2 * N), ncol = N)
> A <- vectrick(C, S, C)
> Cx <- as.array(C)[,,1]
> B <- apply(S, 2, function(x) t(Cx) %*% matrix(x, ncol = J) %*% t(Cx))
> chk(A, B)
> A <- vectrick(C, S, C, transpose = c(FALSE, FALSE))
> Cx <- as.array(C)[,,1]
> B <- apply(S, 2, function(x) Cx %*% matrix(x, ncol = J) %*% Cx)
> chk(A, B)

```

2.12 Convenience Functions

We add a few convenience functions for computing covariance matrices $\Sigma_i = \mathbf{C}_i \mathbf{C}_i^\top$, precision matrices $\mathbf{P}_i = \mathbf{L}_i^\top \mathbf{L}_i$, correlation matrices $\mathbf{R}_i = \tilde{\mathbf{C}}_i \tilde{\mathbf{C}}_i^\top$ (where $\tilde{\mathbf{C}}_i = \text{diag}(\mathbf{C}_i \mathbf{C}_i^\top)^{-\frac{1}{2}} \mathbf{C}_i$), or matrices of partial correlations $\mathbf{A}_i = -\tilde{\mathbf{L}}_i^\top \tilde{\mathbf{L}}_i$ with $\tilde{\mathbf{L}}_i = \mathbf{L}_i \text{diag}(\mathbf{L}_i^\top \mathbf{L}_i)^{-\frac{1}{2}}$ from \mathbf{L}_i (`invchol`) or $\mathbf{C}_i = \mathbf{L}_i^{-1}$ (`chol`) for $i = 1, \dots, N$.

Before we start, let us put a label on lower triangular matrices, such that we can differentiate between \mathbf{C} and \mathbf{L} .

`<chol classes 47> ≡`

```

is.chol <- function(x) inherits(x, "chol")
as.chol <- function(x) {
  stopifnot(is.ltMatrices(x))
  if (is.chol(x)) return(x)
  if (is.invchol(x))
    return(invchol2chol(x))
  class(x) <- c("chol", class(x))
  return(x)
}
is.invchol <- function(x) inherits(x, "invchol")
as.invchol <- function(x) {
  stopifnot(is.ltMatrices(x))
  if (is.invchol(x)) return(x)
  if (is.chol(x))
    return(chol2invchol(x))
  class(x) <- c("invchol", class(x))
  return(x)
}

```

Fragment referenced in 50.

First, we set-up functions for computing $\tilde{\mathbf{C}}_i$

$\langle D \text{ times } C \text{ 48} \rangle \equiv$

```
Dchol <- function(x, D = 1 / sqrt(Tcrossprod(x, diag_only = TRUE))) {  
  if (is.invchol(x)) stop("Dchol cannot work with invchol objects")  
  x <- .adddiag(x)  
  byrow_orig <- attr(x, "byrow")  
  x <- ltMatrices(x, byrow = TRUE)  
  N <- dim(x)[1L]  
  J <- dim(x)[2L]  
  nm <- dimnames(x)[[2L]]  
  ### for some parameter configurations logdet(ret) would  
  ### be -Inf; make sure this doesn't happen  
  if (any(D < .Machine$double.eps))  
    D[D < .Machine$double.eps] <- 2 * .Machine$double.eps  
  if (any(D > 1 / .Machine$double.eps))  
    D[D > 1 / .Machine$double.eps] <- (1 / .Machine$double.eps) / 2  
  x <- unclass(x) * D[rep(1:J, 1:J),,drop = FALSE]  
  ret <- ltMatrices(x, diag = TRUE, byrow = TRUE, names = nm)  
  ret <- as.chol(ltMatrices(ret, byrow = byrow_orig))  
  return(ret)  
}
```

◇
Fragment referenced in [50](#).

and $\tilde{\mathbf{C}}_i^{-1} = \mathbf{L}_i \text{diag}(\mathbf{L}_i^{-1} \mathbf{L}_i^{-\top})^{\frac{1}{2}}$

$\langle L \text{ times } D \text{ 49} \rangle \equiv$

```
### invcholD = solve(Dchol)
invcholD <- function(x, D = sqrt(Tcrossprod(solve(x), diag_only = TRUE))) {

  if (is.chol(x)) stop("invcholD cannot work with chol objects")

  x <- .adddiag(x)

  byrow_orig <- attr(x, "byrow")

  x <- ltMatrices(x, byrow = FALSE)

  N <- dim(x)[1L]
  J <- dim(x)[2L]
  nm <- dimnames(x)[[2L]]

  ### for some parameter configurations logdet(ret) would
  ### be -Inf; make sure this doesn't happen
  if (any(D < .Machine$double.eps))
    D[D < .Machine$double.eps] <- 2 * .Machine$double.eps
  if (any(D > 1 / .Machine$double.eps))
    D[D > 1 / .Machine$double.eps] <- (1 / .Machine$double.eps) / 2

  x <- unclass(x) * D[rep(1:J, J:1),,drop = FALSE]

  ret <- ltMatrices(x, diag = TRUE, byrow = FALSE, names = nm)
  ret <- as.invchol(ltMatrices(ret, byrow = byrow_orig))
  return(ret)
}
◇
```

Fragment referenced in [50](#).

and now the convenience functions are one-liners:

< convenience functions 50 > ≡

```
< chol classes 47 >
< D times C 48 >
< L times D 49 >

### C -> Sigma
chol2cov <- function(x)
  Tcrossprod(x)

### L -> C
invchol2chol <- function(x)
  as.chol(solve(x))

### C -> L
chol2invchol <- function(x)
  as.invchol(solve(x))

### L -> Sigma
invchol2cov <- function(x)
  chol2cov(invchol2chol(x))

### L -> Precision
invchol2pre <- function(x)
  Crossprod(x)

### C -> Precision
chol2pre <- function(x)
  Crossprod(chol2invchol(x))

### C -> R
chol2cor <- function(x) {
  ret <- Tcrossprod(Dchol(x))
  diagonals(ret) <- NULL
  return(ret)
}

### L -> R
invchol2cor <- function(x) {
  ret <- chol2cor(invchol2chol(x))
  diagonals(ret) <- NULL
  return(ret)
}

### L -> A
invchol2pc <- function(x) {
  ret <- -Crossprod(invcholD(x, D = 1 / sqrt(Crossprod(x, diag_only = TRUE))))
  diagonals(ret) <- 0
  ret
}

### C -> A
chol2pc <- function(x)
  invchol2pc(solve(x))
◇
```

Fragment referenced in 2.

Here are some tests

```
> prec2pc <- function(x) {
+   ret <- -cov2cor(x)
+   diag(ret) <- 0
+   ret
+ }
> L <- lxn
> Sigma <- apply(as.array(L), 3,
+               function(x) tcrossprod(solve(x)), simplify = FALSE)
> Prec <- lapply(Sigma, solve)
> Corr <- lapply(Sigma, cov2cor)
> CP <- lapply(Corr, solve)
> PC <- lapply(Prec, function(x) prec2pc(x))
> chk(unlist(Sigma), c(as.array(invchol2cov(L))),
+     check.attributes = FALSE)
> chk(unlist(Prec), c(as.array(invchol2pre(L))),
+     check.attributes = FALSE)
> chk(unlist(Corr), c(as.array(invchol2cor(L))),
+     check.attributes = FALSE)
> chk(unlist(CP), c(as.array(crossprod(invcholD(L)))),
+     check.attributes = FALSE)
> chk(unlist(PC), c(as.array(invchol2pc(L))),
+     check.attributes = FALSE)

> C <- lxn
> Sigma <- apply(as.array(C), 3,
+               function(x) tcrossprod(x), simplify = FALSE)
> Prec <- lapply(Sigma, solve)
> Corr <- lapply(Sigma, cov2cor)
> CP <- lapply(Corr, solve)
> PC <- lapply(Prec, function(x) prec2pc(x))
> chk(unlist(Sigma), c(as.array(chol2cov(C))),
+     check.attributes = FALSE)
> chk(unlist(Prec), c(as.array(chol2pre(C))),
+     check.attributes = FALSE)
> chk(unlist(Corr), c(as.array(chol2cor(C))),
+     check.attributes = FALSE)
> chk(unlist(CP), c(as.array(crossprod(solve(Dchol(C))))),
+     check.attributes = FALSE)
> chk(unlist(PC), c(as.array(chol2pc(C))),
+     check.attributes = FALSE)

> L <- lxd
> Sigma <- apply(as.array(L), 3,
+               function(x) tcrossprod(solve(x)), simplify = FALSE)
> Prec <- lapply(Sigma, solve)
> Corr <- lapply(Sigma, cov2cor)
> CP <- lapply(Corr, solve)
> PC <- lapply(Prec, function(x) prec2pc(x))
> chk(unlist(Sigma), c(as.array(invchol2cov(L))),
+     check.attributes = FALSE)
> chk(unlist(Prec), c(as.array(invchol2pre(L))),
+     check.attributes = FALSE)
> chk(unlist(Corr), c(as.array(invchol2cor(L))),
```

```

+   check.attributes = FALSE)
> chk(unlist(CP), c(as.array(crossprod(invcholD(L))))),
+   check.attributes = FALSE)
> chk(unlist(PC), c(as.array(invchol2pc(L))),
+   check.attributes = FALSE)

> C <- lxd
> Sigma <- apply(as.array(C), 3,
+   function(x) tcrossprod(x), simplify = FALSE)
> Prec <- lapply(Sigma, solve)
> Corr <- lapply(Sigma, cov2cor)
> CP <- lapply(Corr, solve)
> PC <- lapply(Prec, function(x) prec2pc(x))
> chk(unlist(Sigma), c(as.array(chol2cov(C))),
+   check.attributes = FALSE)
> chk(unlist(Prec), c(as.array(chol2pre(C))),
+   check.attributes = FALSE)
> chk(unlist(Corr), c(as.array(chol2cor(C))),
+   check.attributes = FALSE)
> chk(unlist(CP), c(as.array(crossprod(solve(Dchol(C)))))),
+   check.attributes = FALSE)
> chk(unlist(PC), c(as.array(chol2pc(C))),
+   check.attributes = FALSE)

```

We also add an `aperm` method for class `ltMatrices`, implementing the parameters ($\tilde{\mathbf{C}}_i$ or $\tilde{\mathbf{L}}_i$) for permuted versions of the random vectors \mathbf{Y}_i . Let π denote a permutation of $1, \dots, J$ and Π the corresponding permutation matrix. Then, we have $\Pi \mathbf{Y}_i \sim \mathbb{N}_J(\mathbf{0}_J, \Pi \mathbf{C}_i \mathbf{C}_i^\top \Pi^\top)$. Unfortunately, $\Pi \mathbf{C}_i$ is no longer lower triangular, so we have to find the Cholesky decomposition $\tilde{\mathbf{C}}_i \tilde{\mathbf{C}}_i^\top$ of $\Pi \mathbf{C}_i \mathbf{C}_i^\top \Pi^\top$. Of course, $\tilde{\mathbf{L}}_i = \tilde{\mathbf{C}}_i^{-1}$.

The function `aperm`, with argument `perm = π` , now computes the Cholesky factor $\tilde{\mathbf{C}}_i$ of the permuted covariance matrix, or the inverse thereof (in case `x` is of class `invchol`). We start with some tests

```

⟨aperm checks 52⟩ ≡

  J <- dim(a)[2L]
  if (missing(perm)) return(a)
  if (is.character(perm))
    perm <- match(perm, dimnames(a)[[2L]])
  stopifnot(all(perm %in% 1:J))

  args <- list(...)
  if (length(args) > 0L)
    warning("Additional arguments", names(args), "ignored")
  ◇

```

Fragment referenced in 53a.

and then implement the two methods

`< aperm 53a > ≡`

```
aperm.chol <- function(a, perm, ...) {  
  < aperm checks 52 >  
  return(as.chol(chol(chol2cov(a)[,perm])))  
}  
aperm.invchol <- function(a, perm, ...) {  
  < aperm checks 52 >  
  return(chol2invchol(chol(invchol2cov(a)[,perm])))  
}  
◇
```

Fragment defined by [53ab](#).
Fragment referenced in [2](#).

```
> L <- as.invchol(1xn)  
> J <- dim(L)[2L]  
> Lp <- aperm(a = L, perm = p <- sample(1:J))  
> chk(invchol2cov(L)[,p], invchol2cov(Lp))  
> C <- as.chol(1xn)  
> J <- dim(C)[2L]  
> Cp <- aperm(a = C, perm = p <- sample(1:J))  
> chk(chol2cov(C)[,p], chol2cov(Cp))
```

We finally add a method for class `ltMatrices`, for which we actually cannot provide a reasonable result, and for symmetric matrices, where we simply fall-back on subsetting

`< aperm 53b > ≡`

```
aperm.ltMatrices <- function(a, perm, ...)  
  stop("Cannot permute objects of class ltMatrices,  
  consider calling as.chol() or as.invchol() first")  
  
aperm.syMatrices <- function(a, perm, ...)  
  return(a[,perm])  
◇
```

Fragment defined by [53ab](#).
Fragment referenced in [2](#).

2.13 Marginal and Conditional Normal Distributions

Marginal and conditional distributions from distributions $\mathbf{Y}_i \sim \mathbb{N}_J(\boldsymbol{\mu}_i, \mathbf{C}_i \mathbf{C}_i^\top)$ (`chol` argument for \mathbf{C}_i for $i = 1, \dots, N$) or $\mathbf{Y}_i \sim \mathbb{N}_J(\boldsymbol{\mu}_i, \mathbf{L}_i^{-1} \mathbf{L}_i^{-\top})$ (`invchol` argument for \mathbf{L}_i for $i = 1, \dots, N$) shall be computed.

< mc input checks 54a > ≡

```

stopifnot(xor(missing(chol), missing(invchol)))
x <- if (missing(chol)) invchol else chol

stopifnot(is.ltMatrices(x))

N <- dim(x)[1L]
J <- dim(x)[2L]

if (missing(which)) return(x)

if (is.character(which)) which <- match(which, dimnames(x)[[2L]])
stopifnot(all(which %in% 1:J))
◇

```

Fragment referenced in 54b, 57.

The first j marginal distributions can be obtained from subsetting \mathbf{C} or \mathbf{L} directly. Arbitrary marginal distributions are based on the corresponding subset of the covariance matrix for which we compute a corresponding Cholesky factor (such that we can use `lpmvnorm` later on).

< marginal 54b > ≡

```

marg_mvnorm <- function(chol, invchol, which = 1L) {

  < mc input checks 54a >

  if (which[1] == 1L && (length(which) == 1L ||
                        all(diff(which) == 1L))) {
    ### which is 1:j
    tmp <- x[,which]
  } else {
    if (missing(chol)) x <- invchol2chol(x)
    ### note: aperm would work but computes
    ### Cholesky of J^2, here only length(which)^2
    ### is needed
    tmp <- base::chol(chol2cov(x)[,which])
    if (missing(chol)) tmp <- chol2invchol(tmp)
  }

  if (missing(chol))
    ret <- list(invchol = tmp)
  else
    ret <- list(chol = tmp)

  ret
}
◇

```

Fragment referenced in 2.

We compute conditional distributions from the precision matrices $\Sigma_i^{-1} = \mathbf{P}_i = \mathbf{L}_i^\top \mathbf{L}_i$ (we omit the i index from now on). For an arbitrary subset $\mathbf{j} \subset \{1, \dots, J\}$, the conditional distribution of $\mathbf{Y}_{-\mathbf{j}}$ given $\mathbf{Y}_{\mathbf{j}} = \mathbf{y}_{\mathbf{j}}$ is

$$\mathbf{Y}_{-\mathbf{j}} \mid \mathbf{Y}_{\mathbf{j}} = \mathbf{y}_{\mathbf{j}} \sim \mathbb{N}_{|\mathbf{j}|} \left(\boldsymbol{\mu}_{-\mathbf{j}} - \mathbf{P}_{-\mathbf{j},-\mathbf{j}}^{-1} \mathbf{P}_{-\mathbf{j},\mathbf{j}} (\mathbf{y}_{\mathbf{j}} - \boldsymbol{\mu}_{\mathbf{j}}), \mathbf{P}_{-\mathbf{j},-\mathbf{j}}^{-1} \right)$$

and we return a Cholesky factor $\tilde{\mathbf{C}}$ such that $\mathbf{P}_{-j,-j}^{-1} = \tilde{\mathbf{C}}\tilde{\mathbf{C}}^\top$ (if `chol` was given) or $\tilde{\mathbf{L}} = \tilde{\mathbf{C}}^{-1}$ (if `invchol` was given).

We can implement this as

(cond general 55) \equiv

```

stopifnot(!center)

if (!missing(chol)) ### chol is C = Cholesky of covariance
  P <- Crossprod(solve(chol)) ### P = t(L) %*% L with L = C^-1
else
  ### invchol is L = Cholesky of precision
  P <- Crossprod(invchol)

Pw <- P[, -which]
chol <- solve(base::chol(Pw))
Pa <- as.array(P)
Sa <- as.array(S <- Crossprod(chol))
if (dim(chol)[1L] == 1L) {
  Pa <- Pa[, ,1]
  Sa <- Sa[, ,1]
  mean <- -Sa %*% Pa[-which, which, drop = FALSE] %*% given
} else {
  if (ncol(given) == N) {
    mean <- sapply(1:N, function(i)
      -Sa[, ,i] %*% Pa[-which, which, i] %*% given[, i, drop = FALSE])
  } else { ### compare to Mult() with ncol(y) != in% (1, N)
    mean <- sapply(1:N, function(i)
      -Sa[, ,i] %*% Pa[-which, which, i] %*% given)
  }
}
}

```

Fragment referenced in 57.

If $\mathbf{j} = \{1, \dots, j < J\}$ and \mathbf{L} is given, computations simplify a lot because the conditional precision matrix is

$$\mathbf{P}_{-j,-j} = (\mathbf{L}^\top \mathbf{L})_{-j,-j} = \mathbf{L}_{-j,-j}^\top \mathbf{L}_{-j,-j}$$

and thus we return $\tilde{\mathbf{L}} = \mathbf{L}_{-j,-j}$ (if `invchol` was given) or $\tilde{\mathbf{C}} = \mathbf{L}_{-j,-j}^{-1}$ (if `chol` was given). The conditional mean is

$$\begin{aligned} \boldsymbol{\mu}_{-j} - \mathbf{P}_{-j,-j}^{-1} \mathbf{P}_{-j,j} (\mathbf{y}_j - \boldsymbol{\mu}_j) &= \boldsymbol{\mu}_{-j} - \mathbf{L}_{-j,-j}^{-1} \mathbf{L}_{-j,-j}^\top \mathbf{L}_{-j,-j} \mathbf{L}_{-j,j} (\mathbf{y}_j - \boldsymbol{\mu}_j) \\ &= \boldsymbol{\mu}_{-j} - \mathbf{L}_{-j,-j}^{-1} \mathbf{L}_{-j,j} (\mathbf{y}_j - \boldsymbol{\mu}_j). \end{aligned}$$

We sometimes, for example when scores with respect to $\mathbf{L}_{-j,-j}^{-1}$ shall be computed in `slpmvnorm`, need the negative rescaled mean

$$\mathbf{L}_{-j,j} (\mathbf{y}_j - \boldsymbol{\mu}_j) \tag{2.1}$$

and the `center = TRUE` argument triggers this values to be returned.

The implementation reads

< cond simple 56 > ≡

```
if (which[1] == 1L && (length(which) == 1L ||
    all(diff(which) == 1L))) {
  ### which is 1:j
  L <- if (missing(invchol)) solve(chol) else invchol
  tmp <- matrix(0, ncol = ncol(given), nrow = J - length(which))
  centerm <- Mult(L, rbind(given, tmp))
  ### if ncol(given) is not N = dim(L)[1L] > 1, then
  ### solve() below won't work and we loop over
  ### columns of centerm
  if (dim(L)[1L] > 1 && ncol(given) != N) {
    centerm <- lapply(1:ncol(centerm), function(j)
      matrix(centerm[,j], nrow = J, ncol = N)[-which,,drop = FALSE]
    )
  } else {
    centerm <- centerm[-which,,drop = FALSE]
  }
  L <- L[,-which]
  ct <- centerm
  if (!is.matrix(ct)) ct <- do.call("rbind", ct)
  if (is.matrix(centerm)) {
    m <- -solve(L, centerm)
  } else {
    m <- do.call("rbind", lapply(centerm, function(cm) -solve(L, cm)))
  }
  if (missing(invchol)) {
    if (center)
      return(list(center = ct, chol = solve(L)))
    return(list(mean = m, chol = solve(L)))
  }
  if (center)
    return(list(center = ct, invchol = L))
  return(list(mean = m, invchol = L))
}
◇
```

Fragment referenced in [57](#).

Note that we could have avoided the general case altogether by first computing a Cholesky decomposition of the permuted covariance matrix (such that the conditioning variables come first). The code above only decomposes the marginal (and thus lower-dimensional) covariance. However, we didn't implement the `center = TRUE` case, so we can fall back on the permuted version if this option is requested. Putting everything together gives

< conditional 57 > ≡

```
cond_mvnorm <- function(chol, invchol, which_given = 1L, given, center = FALSE) {  
  
  which <- which_given  
  < mc input checks 54a >  
  
  if (N == 1) N <- NCOL(given)  
  stopifnot(is.matrix(given) && nrow(given) == length(which))  
  
  < cond simple 56 >  
  
  ### general with center = TRUE => permute first and go simple  
  if (center) {  
    perm <- c(which, (1:J)[!(1:J) %in% which])  
    if (!missing(chol))  
      return(cond_mvnorm(chol = aperm(as.chol(chol), perm = perm),  
                          which_given = 1:length(which), given = given,  
                          center = center))  
    return(cond_mvnorm(invchol = aperm(as.invchol(invchol), perm = perm),  
                        which_given = 1:length(which), given = given,  
                        center = center))  
  }  
  
  < cond general 55 >  
  
  chol <- base::chol(S)  
  if (missing(invchol))  
    return(list(mean = mean, chol = chol))  
  
  return(list(mean = mean, invchol = solve(chol)))  
}  
◇
```

Fragment referenced in 2.

Let's check this against the commonly used formula based on the covariance matrix, first for the marginal distribution

```
> Sigma <- tcrossprod(lxd)  
> j <- 1:3  
> chk(Sigma[,j], tcrossprod(marg_mvnorm(chol = lxd, which = j)$chol))  
> j <- 2:4  
> chk(Sigma[,j], tcrossprod(marg_mvnorm(chol = lxd, which = j)$chol))  
> Sigma <- tcrossprod(solve(lxd))  
> j <- 1:3  
> chk(Sigma[,j], tcrossprod(solve(marg_mvnorm(invchol = lxd, which = j)$invchol))  
> j <- 2:4  
> chk(Sigma[,j], tcrossprod(solve(marg_mvnorm(invchol = lxd, which = j)$invchol))
```

and then for conditional distributions. The general case is

```
> Sigma <- as.array(tcrossprod(lxd))[, , 1]  
> j <- 2:4  
> y <- matrix(c(-1, 2, 1), nrow = 3)  
> cm <- Sigma[-j, j, drop = FALSE] %*% solve(Sigma[j, j]) %*% y  
> cS <- Sigma[-j, -j] - Sigma[-j, j, drop = FALSE] %*%
```

```

+       solve(Sigma[j,j]) %*% Sigma[j,-j,drop = FALSE]
> cmv <- cond_mvnorm(chol = lxd[1,], which_given = j, given = y)
> chk(cm, cmv$mean)
> chk(cS, as.array(tcrossprod(cmv$chol))[,1])
> Sigma <- as.array(tcrossprod(solve(lxd)))[,1]
> j <- 2:4
> y <- matrix(c(-1, 2, 1), nrow = 3)
> cm <- Sigma[-j, j,drop = FALSE] %*% solve(Sigma[j,j]) %*% y
> cS <- Sigma[-j, -j] - Sigma[-j,j,drop = FALSE] %*%
+       solve(Sigma[j,j]) %*% Sigma[j,-j,drop = FALSE]
> cmv <- cond_mvnorm(invchol = lxd[1,], which_given = j, given = y)
> chk(cm, cmv$mean)
> chk(cS, as.array(tcrossprod(solve(cmv$invchol)))[,1])

```

and the simple case is

```

> Sigma <- as.array(tcrossprod(lxd))[,1]
> j <- 1:3
> y <- matrix(c(-1, 2, 1), nrow = 3)
> cm <- Sigma[-j, j,drop = FALSE] %*% solve(Sigma[j,j]) %*% y
> cS <- Sigma[-j, -j] - Sigma[-j,j,drop = FALSE] %*%
+       solve(Sigma[j,j]) %*% Sigma[j,-j,drop = FALSE]
> cmv <- cond_mvnorm(chol = lxd[1,], which_given = j, given = y)
> chk(c(cm), c(cmv$mean))
> chk(cS, as.array(tcrossprod(cmv$chol))[,1])
> Sigma <- as.array(tcrossprod(solve(lxd)))[,1]
> j <- 1:3
> y <- matrix(c(-1, 2, 1), nrow = 3)
> cm <- Sigma[-j, j,drop = FALSE] %*% solve(Sigma[j,j]) %*% y
> cS <- Sigma[-j, -j] - Sigma[-j,j,drop = FALSE] %*%
+       solve(Sigma[j,j]) %*% Sigma[j,-j,drop = FALSE]
> cmv <- cond_mvnorm(invchol = lxd[1,], which_given = j, given = y)
> chk(c(cm), c(cmv$mean))
> chk(cS, as.array(tcrossprod(solve(cmv$invchol)))[,1])

```

2.14 Continuous Log-likelihoods

With $\mathbf{Z} \sim \mathbb{N}_J(0, \mathbf{I}_J)$ and $\mathbf{Y} = \mathbf{C}_i \mathbf{Z} + \boldsymbol{\mu}_i \sim \mathbb{N}_J(\boldsymbol{\mu}_i, \mathbf{C}_i \mathbf{C}_i^\top)$ we want to evaluate the log-likelihood contributions for observations $\mathbf{y}_1, \dots, \mathbf{y}_N$ in a function called `ldmvnorm`

<ldmvnorm 59a> ≡

```
ldmvnorm <- function(obs, mean = 0, chol, invchol, logLik = TRUE) {  
  
  stopifnot(xor(missing(chol), missing(invchol)))  
  if (!is.matrix(obs)) obs <- matrix(obs, ncol = 1L)  
  p <- ncol(obs)  
  
  if (!missing(chol)) {  
    <ldmvnorm chol 61a>  
  } else {  
    <ldmvnorm invchol 61b>  
  }  
  
  names(logretval) <- colnames(obs)  
  if (logLik) return(sum(logretval))  
  return(logretval)  
}  
◇
```

Fragment referenced in 66.

We first check if the observations $\mathbf{y}_1, \dots, \mathbf{y}_N$ are given in an $J \times N$ matrix `obs` with corresponding means $\boldsymbol{\mu}_1, \dots, \boldsymbol{\mu}_N$ in `means` and return the mean-centered observations.

<check obs 59b> ≡

```
.check_obs <- function(obs, mean, J, N) {  
  
  nr <- nrow(obs)  
  nc <- ncol(obs)  
  if (nc != N)  
    stop("obs and (inv)chol have non-conforming size")  
  if (nr != J)  
    stop("obs and (inv)chol have non-conforming size")  
  if (identical(unique(mean), 0)) return(obs)  
  if (length(mean) == J)  
    return(obs - c(mean))  
  if (!is.matrix(mean))  
    stop("obs and mean have non-conforming size")  
  if (nrow(mean) != nr)  
    stop("obs and mean have non-conforming size")  
  if (ncol(mean) != nc)  
    stop("obs and mean have non-conforming size")  
  return(obs - mean)  
}  
◇
```

Fragment referenced in 2.

With $\boldsymbol{\Sigma}_i = \mathbf{C}_i \mathbf{C}_i^\top$ the log-likelihood function for $\mathbf{Y}_i = \mathbf{y}_i$ is

$$\ell_i(\boldsymbol{\mu}_i, \mathbf{C}_i) = -\frac{k}{2} \log(2\pi) - \frac{1}{2} \log |\boldsymbol{\Sigma}_i| - \frac{1}{2} (\mathbf{y}_i - \boldsymbol{\mu}_i)^\top \boldsymbol{\Sigma}_i^{-1} (\mathbf{y}_i - \boldsymbol{\mu}_i)$$

Because $\log |\boldsymbol{\Sigma}_i| = \log |\mathbf{C}_i \mathbf{C}_i^\top| = 2 \log |\mathbf{C}_i| = 2 \sum_{j=1}^J \log \text{diag}(\mathbf{C}_i)_j$ we get the simpler expression

$$\ell_i(\boldsymbol{\mu}_i, \mathbf{C}_i) = -\frac{k}{2} \log(2\pi) - \sum_{j=1}^J \log \text{diag}(\mathbf{C}_i)_j - \frac{1}{2} (\mathbf{y}_i - \boldsymbol{\mu}_i)^\top \mathbf{C}_i^{-\top} \mathbf{C}_i^{-1} (\mathbf{y}_i - \boldsymbol{\mu}_i). \quad (2.2)$$

We need to compute `colSums(dnorm(z, log = TRUE))` quite often. This turns out to be time-consuming and memory intensive, so we provide a small internal helper function focusing on the necessary computations.

`< colSumsdnorm 60a > ≡`

```
SEXP R_ltMatrices_colSumsdnorm (SEXP z, SEXP N, SEXP J) {
  /* number of columns */
  int iN = INTEGER(N)[0];
  /* number of rows */
  int iJ = INTEGER(J)[0];
  SEXP ans;
  double *dans, J12pi, *dz;

  J12pi = iJ * log(2 * M_PI);
  PROTECT(ans = allocVector(REALSXP, iN));
  dans = REAL(ans);
  dz = REAL(z);

  for (int i = 0; i < iN; i++) {
    dans[i] = 0.0;
    for (int j = 0; j < iJ; j++)
      dans[i] += pow(dz[j], 2);
    dans[i] = - 0.5 * (J12pi + dans[i]);
    dz += iJ;
  }

  UNPROTECT(1);
  return(ans);
}
◇
```

Fragment referenced in 3.

`< colSumsdnorm ltMatrices 60b > ≡`

```
.colSumsdnorm <- function(z) {
  stopifnot(is.numeric(z))
  if (!is.matrix(z))
    z <- matrix(z, nrow = 1, ncol = length(z))
  ret <- .Call(mvtnorm_R_ltMatrices_colSumsdnorm, z, ncol(z), nrow(z))
  names(ret) <- colnames(z)
  return(ret)
}
◇
```

Fragment referenced in 2.

The main part is now

`<ldmnorm chol 61a> ≡`

```

if (missing(chol))
  stop("either chol or invchol must be given")
## chol is given
if (!is.ltMatrices(chol))      ### NOTE: replace with is.chol
  stop("chol is not an object of class ltMatrices")
N <- dim(chol)[1L]
N <- ifelse(N == 1, p, N)
J <- dim(chol)[2L]
## NOTE: obs is now mean-centered
obs <- .check_obs(obs = obs, mean = mean, J = J, N = N)
z <- solve(chol, obs)
logretval <- .colSumsdnorm(z)
if (attr(chol, "diag"))
  logretval <- logretval - logdet(chol)
◇

```

Fragment referenced in [59a](#).

where we can use the efficient implementations of `solve` and `logdet`.

If $\mathbf{L}_i = \mathbf{C}_i^{-1}$ is given, we obtain

$$\ell_i(\boldsymbol{\mu}_i, \mathbf{L}_i) = -\frac{k}{2} \log(2\pi) + \sum_{j=1}^J \log \text{diag}(\mathbf{L}_i)_j - \frac{1}{2} (\mathbf{y}_i - \boldsymbol{\mu}_i)^\top \mathbf{L}_i^\top \mathbf{L}_i (\mathbf{y}_i - \boldsymbol{\mu}_i).$$

`<ldmnorm invchol 61b> ≡`

```

## invchol is given
if (!is.ltMatrices(invchol))  ### NOTE: replace with is.invchol
  stop("invchol is not an object of class ltMatrices")
N <- dim(invchol)[1L]
N <- ifelse(N == 1, p, N)
J <- dim(invchol)[2L]
obs <- .check_obs(obs = obs, mean = mean, J = J, N = N)
## NOTE: obs is (J x N) and mean-centered
## dnorm takes rather long
z <- Mult(invchol, obs)
logretval <- .colSumsdnorm(z)
## note that the second summand gets recycled the correct number
## of times in case dim(invchol)[1L] == 1 but ncol(obs) > 1
if (attr(invchol, "diag"))
  logretval <- logretval + logdet(invchol)
◇

```

Fragment referenced in [59a](#).

The score function with respect to `obs` is

$$\frac{\partial \ell_i(\boldsymbol{\mu}_i, \mathbf{L}_i)}{\partial \mathbf{y}_i} = -\mathbf{L}_i^\top \mathbf{L}_i (\mathbf{y}_i - \boldsymbol{\mu}_i)$$

and with respect to `invchol` we have

$$\frac{\partial \ell_i(\boldsymbol{\mu}_i, \mathbf{L}_i)}{\partial \mathbf{L}_i} = -2\mathbf{L}_i (\mathbf{y}_i - \boldsymbol{\mu}_i) (\mathbf{y}_i - \boldsymbol{\mu}_i)^\top + \text{diag}(\mathbf{L}_i)^{-1}.$$

The score function with respect to `chol` post-processes the above score using the vec-trick (Section 2.11). For the log-likelihood (2.2), the score with respect to \mathbf{C}_i is the sum of the score functions of the two terms. We start with the simpler first term

$$\frac{\partial - \sum_{j=1}^J \log \text{diag}(\mathbf{C}_i)_j}{\partial \mathbf{C}_i} = -\text{diag}(\mathbf{C}_i)^{-1}$$

The second term gives (we omit the mean for the sake of simplicity)

$$\begin{aligned} \frac{\partial - \mathbf{y}_i^\top \mathbf{C}_i^{-\top} \mathbf{C}_i^{-1} \mathbf{y}_i}{\partial \mathbf{C}_i} &= - \left. \frac{\partial \mathbf{y}_i^\top \mathbf{A}^\top \mathbf{A} \mathbf{y}_i}{\partial \mathbf{A}} \right|_{\mathbf{A}=\mathbf{C}_i^{-1}} \left. \frac{\partial \mathbf{A}^{-1}}{\partial \mathbf{A}} \right|_{\mathbf{A}=\mathbf{C}_i} \\ &= -2\text{vec}(\mathbf{C}_i^{-1} \mathbf{y}_i \mathbf{y}_i^\top)^\top (-1) (\mathbf{C}_i^{-\top} \otimes \mathbf{C}_i^{-1}) \\ &= 2\text{vec}(\mathbf{C}_i^{-\top} \mathbf{C}_i^{-1} \mathbf{y}_i \mathbf{y}_i^\top \mathbf{C}_i^{-\top})^\top \end{aligned}$$

In `sldmnorm`, we compute the score with respect to \mathbf{L}_i and use the above relationship to compute the score with respect to \mathbf{C}_i .

< sldmvnorm 63 > ≡

```
sldmvnorm <- function(obs, mean = 0, chol, invchol, logLik = TRUE) {  
  
  stopifnot(xor(missing(chol), missing(invchol)))  
  if (!is.matrix(obs)) obs <- matrix(obs, ncol = 1L)  
  
  if (!missing(invchol)) {  
  
    N <- dim(invchol)[1L]  
    N <- ifelse(N == 1, ncol(obs), N)  
    J <- dim(invchol)[2L]  
    obs <- .check_obs(obs = obs, mean = mean, J = J, N = N)  
    ## NOTE: obs is mean-centered now  
  
    Mix <- Mult(invchol, obs)  
    sobs <- - Mult(invchol, Mix, transpose = TRUE)  
  
    Y <- matrix(obs, byrow = TRUE, nrow = J, ncol = N * J)  
    ret <- - matrix(Mix[, rep(1:N, each = J)] * Y, ncol = N)  
  
    M <- matrix(1:(J^2), nrow = J, byrow = FALSE)  
    ret <- ret[M[lower.tri(M, diag = attr(invchol, "diag"))],,drop = FALSE]  
    if (!is.null(dimnames(invchol)[[1L]]))  
      colnames(ret) <- dimnames(invchol)[[1]]  
    ret <- ltMatrices(ret,  
                      diag = attr(invchol, "diag"), byrow = FALSE,  
                      names = dimnames(invchol)[[2L]])  
    ret <- ltMatrices(ret, diag = attr(invchol, "diag"),  
                      byrow = attr(invchol, "byrow"))  
    if (attr(invchol, "diag")) {  
      ### recycle properly  
      diagonals(ret) <- diagonals(ret) + c(1 / diagonals(invchol))  
    } else {  
      diagonals(ret) <- 0  
    }  
    ret <- list(obs = sobs, invchol = ret)  
    if (logLik)  
      ret$logLik <- ldmvnorm(obs = obs,  
                            invchol = invchol, logLik = FALSE)  
    return(ret)  
  }  
  
  invchol <- solve(chol)  
  ret <- sldmvnorm(obs = obs, mean = mean, invchol = invchol)  
  ### this means: ret$chol <- - vectrick(invchol, ret$invchol, invchol)  
  ret$chol <- as.chol(- vectrick(invchol, ret$invchol))  
  ret$invchol <- NULL  
  return(ret)  
}  
◇
```

Fragment referenced in 66.

2.15 Application Example

Let's say we have $\mathbf{Y}_i \sim \mathbb{N}_J(\mathbf{0}_J, \mathbf{C}_i \mathbf{C}_i^\top)$ for $i = 1, \dots, N$ and we know the Cholesky factors $\mathbf{L}_i = \mathbf{C}_i^{-1}$ of the N precision matrices $\Sigma_i^{-1} = \mathbf{L}_i \mathbf{L}_i^\top$. We generate $\mathbf{Y}_i = \mathbf{L}_i^{-1} \mathbf{Z}_i$ from $\mathbf{Z}_i \sim \mathbb{N}_J(\mathbf{0}_J, \mathbf{I}_J)$. Evaluating the corresponding log-likelihood is now straightforward and fast, compared to repeated calls to `dmvnorm`

```
> N <- 1000L
> J <- 50L
> lt <- ltMatrices(matrix(runif(N * J * (J + 1) / 2) + 1, ncol = N),
+                   diag = TRUE, byrow = FALSE)
> Z <- matrix(rnorm(N * J), ncol = N)
> Y <- solve(lt, Z)
> ll1 <- sum(dnorm(lt %*% Y, log = TRUE)) + sum(log(diagonals(lt)))
> S <- as.array(tcrossprod(solve(lt)))
> ll2 <- sum(sapply(1:N, function(i)
+           dmvnorm(x = Y[,i], sigma = S[, ,i], log = TRUE)))
> chk(ll1, ll2)
```

The `ldmvnorm` function now also has `chol` and `invchol` arguments such that we can use

```
> ll3 <- ldmvnorm(obs = Y, invchol = lt)
> chk(ll1, ll3)
```

Note that argument `obs` in `ldmvnorm` is an $J \times N$ matrix whereas the traditional interface in `dmvnorm` expects an $N \times J$ matrix `x`. The reason is that `Mult` or `solve` work with $J \times N$ matrices and we want to avoid matrix transposes.

Sometimes it is preferable to split the joint distribution into a marginal distribution of some elements and the conditional distribution given these elements. The joint density is, of course, the product of the marginal and conditional densities and we can check if this works for our example by

```
> ## marginal of and conditional on these
> (j <- 1:5 * 10)

[1] 10 20 30 40 50

> md <- marg_mvnorm(invchol = lt, which = j)
> cd <- cond_mvnorm(invchol = lt, which_given = j, given = Y[j,])
> ll3 <- sum(dnorm(md$invchol %*% Y[j,], log = TRUE)) +
+         sum(log(diagonals(md$invchol))) +
+         sum(dnorm(cd$invchol %*% (Y[-j,] - cd$mean), log = TRUE)) +
+         sum(log(diagonals(cd$invchol)))
> chk(ll1, ll3)
```

Chapter 3

Multivariate Normal Log-likelihoods

We now discuss code for evaluating the log-likelihood

$$\sum_{i=1}^N \log(p_i(\mathbf{C}_i \mid \mathbf{a}_i, \mathbf{b}_i))$$

This is relatively simple to achieve using the existing `pmvnorm` function, so a prototype might look like

< lpmvnormR 65 > \equiv

```
lpmvnormR <- function(lower, upper, mean = 0, center = NULL, chol, logLik = TRUE, ...) {  
  < input checks 68 >  
  
  sigma <- Tcrossprod(chol)  
  S <- as.array(sigma)  
  idx <- 1  
  
  ret <- error <- numeric(N)  
  for (i in 1:N) {  
    if (dim(sigma)[[1L]] > 1) idx <- i  
    tmp <- pmvnorm(lower = lower[,i], upper = upper[,i], sigma = S[, ,idx], ...)  
    ret[i] <- tmp  
    error[i] <- attr(tmp, "error")  
  }  
  attr(ret, "error") <- error  
  
  if (logLik)  
    return(sum(log(pmax(ret, .Machine$double.eps))))  
  
  ret  
}
```

\diamond

Fragment never referenced.

However, the underlying FORTRAN code first computes the Cholesky factor based on the covariance matrix, which is clearly a waste of time. Repeated calls to FORTRAN also cost some time. The code (based on and evaluated in [Genz and Bretz, 2002](#)) implements a specific form of quasi-Monte-Carlo integration without allowing the user to change the scheme (or to fall-back to simple Monte-Carlo). We therefore implement our own simplified version, with the aim to speed-things up such that maximum-likelihood estimation becomes a bit faster.

Let's look at an example first. This code estimates p_1, \dots, p_{10} for a 5-dimensional normal

```
> J <- 5L
> N <- 10L
> x <- matrix(runif(N * J * (J + 1) / 2), ncol = N)
> lx <- ltMatrices(x, byrow = TRUE, diag = TRUE)
> a <- matrix(runif(N * J), nrow = J) - 2
> a[sample(J * N)[1:2]] <- -Inf
> b <- a + 2 + matrix(runif(N * J), nrow = J)
> b[sample(J * N)[1:2]] <- Inf
> (phat <- c(lpmvnormR(a, b, chol = lx, logLik = FALSE)))

[1] 0.2369 0.2337 0.2842 0.3915 0.4662 0.0000 0.5901 0.4619 0.4873 0.0000
```

We want to achieve the same result a bit more general and a bit faster, by making the code more modular and, most importantly, by providing score functions for all arguments \mathbf{a}_i , \mathbf{b}_i , and \mathbf{C}_i .

3.1 Algorithm

"lpmvnorm.R" 66≡

```
⟨ R Header 145 ⟩
⟨ lpmvnorm 77 ⟩
⟨ slpmvnorm 89 ⟩
⟨ ldmvnorm 59a ⟩
⟨ sldmvnorm 63 ⟩
⟨ ldpmvnorm 101 ⟩
⟨ sldpmvnorm 103 ⟩
⟨ deperma 108 ⟩
⟨ standardize 110 ⟩
⟨ destandardize 112 ⟩
⟨ lpRR 142 ⟩
⟨ slpRR 143 ⟩
◇
```

"lpmvnorm.c" 67≡

```
⟨ C Header 146 ⟩
#ifndef USE_FC_LEN_T
# define USE_FC_LEN_T
#endif
#include <Rconfig.h>
#include <R_ext/BLAS.h> /* for dtrmm */
#ifndef FCONE
# define FCONE
#endif
#include <R.h>
#include <Rmath.h>
#include <Rinternals.h>
#include <Rdefines.h>
⟨ pnorm fast 72b ⟩
⟨ pnorm slow 72c ⟩
⟨ R lpmvnorm 75 ⟩
⟨ R slpmvnorm 86 ⟩
◇
```

We implement the algorithm described by [Genz \(1992\)](#). The key point here is that the original J -dimensional problem (1.1) is transformed into an integral over $[0, 1]^{J-1}$.

For each $i = 1, \dots, N$, do

1. Input \mathbf{C}_i (chol), \mathbf{a}_i (lower), \mathbf{b}_i (upper), and control parameters α , ϵ , and M_{\max} (M).

(input checks 68) \equiv

```

if (!is.matrix(lower)) lower <- matrix(lower, ncol = 1)
if (!is.matrix(upper)) upper <- matrix(upper, ncol = 1)
stopifnot(isTRUE(all.equal(dim(lower), dim(upper))))

stopifnot(is.ltMatrices(chol))          ### NOTE: replace with is.chol
byrow_orig <- attr(chol, "byrow")
chol <- ltMatrices(chol, byrow = TRUE)
d <- dim(chol)
### allow single matrix C
N <- ifelse(d[1L] == 1, ncol(lower), d[1L])
J <- d[2L]

stopifnot(nrow(lower) == J && ncol(lower) == N)
stopifnot(nrow(upper) == J && ncol(upper) == N)
if (is.matrix(mean)) {
  if (ncol(mean) == 1L)
    mean <- mean[,rep(1, N),drop = FALSE]
  stopifnot(nrow(mean) == J && ncol(mean) == N)
}

lower <- lower - mean
upper <- upper - mean

if (!is.null(center)) {
  if (!is.matrix(center)) center <- matrix(center, ncol = 1)
  stopifnot(nrow(center) == J && ncol(center) == N)
}

```

Fragment referenced in [65](#), [77](#), [89](#).

2. Standardise integration limits $a_j^{(i)}/c_{jj}^{(i)}$, $b_j^{(i)}/c_{jj}^{(i)}$, and rows $c_{jj}^{(i)}/c_{jj}^{(i)}$ for $1 \leq j < j < J$.

< standardise 69a > ≡

```
if (attr(chol, "diag")) {
  ### diagonals returns J x N and lower/upper are J x N, so
  ### elementwise standardisation is simple
  dchol <- diagonals(chol)
  ### zero diagonals not allowed
  stopifnot(all(abs(dchol) > (.Machine$double.eps)))
  ac <- lower / c(dchol)
  bc <- upper / c(dchol)
  C <- Dchol(chol, D = 1 / dchol)
  if (J > 1) { ### else: univariate problem; C is no longer used
    uC <- Lower_tri(C)
  } else {
    uC <- unclass(C)
  }
} else {
  ac <- lower
  bc <- upper
  uC <- Lower_tri(chol)
}
◇
```

Fragment referenced in 77, 89.

3. Initialise $\text{intsum} = \text{varsum} = 0$, $M = 0$, $d_1 = \Phi(a_1^{(i)})$, $e_1 = \Phi(b_1^{(i)})$ and $f_1 = e_1 - d_1$.

Here, `center` is the negative rescaled mean (2.1) allowing to evaluate probabilities for conditional multivariate normal distributions in `lpmvnorm` and corresponding scores in `slpmvnorm`. This feature is not present in the original formulation of the algorithm.

< initialisation 69b > ≡

```
x0 = 0.0;
if (LENGTH(center))
  x0 = -dcenter[0];
d0 = pnorm_ptr(da[0], x0);
e0 = pnorm_ptr(db[0], x0);
emd0 = e0 - d0;
f0 = emd0;
intsum = (iJ > 1 ? 0.0 : f0);
◇
```

Fragment referenced in 75, 86.

4. Repeat

< init logLik loop 69c > ≡

```
d = d0;
f = f0;
emd = emd0;
start = 0;
◇
```

Fragment referenced in 75, 81b.

- (a) Generate uniform $w_1, \dots, w_{J-1} \in [0, 1]$.
 (b) For $j = 2, \dots, J$ set

$$y_{j-1} = \Phi^{-1}(d_{j-1} + w_{j-1}(e_{j-1} - d_{j-1}))$$

We either generate w_{j-1} on the fly or use pre-computed weights (\mathbf{w}).

< compute y 70a > \equiv

```

Wtmp = (W == R_NilValue ? unif_rand() : dW[j - 1]);
tmp = d + Wtmp * emd;
if (tmp < dtol) {
  y[j - 1] = q0;
} else {
  if (tmp > mdtol)
    y[j - 1] = -q0;
  else
    y[j - 1] = qnorm(tmp, 0.0, 1.0, 1L, 0L);
}

```

Fragment referenced in [71b](#), [85a](#).

$$x_{j-1} = \sum_{j=1}^{j-1} c_{jj}^{(i)} y_j$$

< compute x 70b > \equiv

```

x = 0.0;
if (LENGTH(center)) {
  for (k = 0; k < j; k++)
    x += dC[start + k] * (y[k] - dcenter[k]);
  x -= dcenter[j];
} else {
  for (k = 0; k < j; k++)
    x += dC[start + k] * y[k];
}

```

Fragment referenced in [71b](#), [85a](#).

$$d_j = \Phi\left(a_j^{(i)} - x_{j-1}\right)$$

$$e_j = \Phi\left(b_j^{(i)} - x_{j-1}\right)$$

< update d, e 70c > \equiv

```

d = pnorm_ptr(da[j], x);
e = pnorm_ptr(db[j], x);
emd = e - d;

```

Fragment referenced in [71b](#), [85a](#).

$$f_j = (e_j - d_j)f_{j-1}.$$

`< update f 71a > ≡`

```

    start += j;
    f *= emd;
    ◇

```

Fragment referenced in [71b](#), [85a](#).

We put everything together in a loop starting with the second dimension

`< inner logLik loop 71b > ≡`

```

    for (j = 1; j < iJ; j++) {
        < compute y 70a >
        < compute x 70b >
        < update d, e 70c >
        < update f 71a >
    }
    ◇

```

Fragment referenced in [75](#).

- (c) Set $\text{intsum} = \text{intsum} + f_J$, $\text{varsum} = \text{varsum} + f_J^2$, $M = M + 1$, and $\text{error} = \sqrt{(\text{varsum}/M - (\text{intsum}/M)^2)/M}$.

`< increment 71c > ≡`

```

    intsum += f;
    ◇

```

Fragment referenced in [75](#).

We refrain from early stopping and error estimation.

Until $\text{error} < \epsilon$ or $M = M_{\max}$

5. Output $\hat{p}_i = \text{intsum}/M$.

We return $\log \hat{p}_i$ for each i , or we immediately sum-up over i .

`< output 71d > ≡`

```

    dans[0] += (intsum < dtol ? 10 : log(intsum)) - 1M;
    if (!RlogLik)
        dans += 1L;
    ◇

```

Fragment referenced in [75](#).

and move on to the next observation (note that p might be 0 in case $\mathbf{C}_i \equiv \mathbf{C}$).

< move on 72a > ≡

```
da += iJ;
db += iJ;
dC += p;
if (LENGTH(center)) dcenter += iJ;
◇
```

Fragment referenced in 75, 86.

It turned out that calls to `pnorm` are expensive, so a slightly faster alternative (suggested by [Matić *et al.*, 2018](#)) might provide an alternative which can be requested from using (`fast = TRUE` in the calls to `lpmvnorm` and `slpmvnorm`):

< pnorm fast 72b > ≡

```
/* see https://doi.org/10.2139/ssrn.2842681 */
const double g2 = -0.0150234471495426236132;
const double g4 = 0.000666098511701018747289;
const double g6 = 5.07937324518981103694e-06;
const double g8 = -2.92345273673194627762e-06;
const double g10 = 1.34797733516989204361e-07;
const double m2dpi = -2.0 / M_PI; //3.141592653589793115998;

double C_pnorm_fast (double x, double m) {

    double tmp, ret;
    double x2, x4, x6, x8, x10;

    if (R_FINITE(x)) {
        x = x - m;
        x2 = x * x;
        x4 = x2 * x2;
        x6 = x4 * x2;
        x8 = x6 * x2;
        x10 = x8 * x2;
        tmp = 1 + g2 * x2 + g4 * x4 + g6 * x6 + g8 * x8 + g10 * x10;
        tmp = m2dpi * x2 * tmp;
        ret = .5 + ((x > 0) - (x < 0)) * sqrt(1 - exp(tmp)) / 2.0;
    } else {
        ret = (x > 0 ? 1.0 : 0.0);
    }
    return(ret);
}
◇
```

Fragment referenced in 67.

< pnorm slow 72c > ≡

```
double C_pnorm_slow (double x, double m) {
    return(pnorm(x, m, 1.0, 1L, 0L));
}
◇
```

Fragment referenced in 67.

The `fast` argument can be used to switch on the faster but less accurate version of `pnorm`

< pnorm 73a > ≡

```
Rboolean Rfast = asLogical(fast);
double (*pnorm_ptr)(double, double) = C_pnorm_slow;
if (Rfast)
  pnorm_ptr = C_pnorm_fast;
◇
```

Fragment referenced in [75](#), [86](#).

We allow a new set of weights for each observation or one set for all observations. In the former case, the number of columns is $M \times N$ and in the latter just M .

< W length 73b > ≡

```
int pW = 0;
if (W != R_NilValue) {
  if (LENGTH(W) == (iJ - 1) * iM) {
    pW = 0;
  } else {
    if (LENGTH(W) != (iJ - 1) * iN * iM)
      error("Length of W incorrect");
    pW = 1;
  }
  dW = REAL(W);
}
◇
```

Fragment referenced in [75](#), [86](#).

< dimensions 73c > ≡

```
int iM = INTEGER(M)[0];
int iN = INTEGER(N)[0];
int iJ = INTEGER(J)[0];

da = REAL(a);
db = REAL(b);
dC = REAL(C);
dW = REAL(C); // make -Wmaybe-uninitialized happy

if (LENGTH(C) == iJ * (iJ - 1) / 2)
  p = 0;
else
  p = LENGTH(C) / iN;
◇
```

Fragment referenced in [75](#), [86](#).

< setup return object 74a > ≡

```
len = (RlogLik ? 1 : iN);
PROTECT(ans = allocVector(REALSXP, len));
dans = REAL(ans);
for (int i = 0; i < len; i++)
    dans[i] = 0.0;
◇
```

Fragment referenced in [75](#).

The case $J = 1$ does not loop over M

< univariate problem 74b > ≡

```
if (iJ == 1) {
    iM = 0;
    lM = 0.0;
} else {
    lM = log((double) iM);
}
◇
```

Fragment referenced in [75](#).

< init center 74c > ≡

```
dcenter = REAL(center);
if (LENGTH(center)) {
    if (LENGTH(center) != iN * iJ)
        error("incorrect dimensions of center");
}
◇
```

Fragment referenced in [75](#), [86](#).

We put the code together in a dedicated C function

< R slpmvnorm variables 74d > ≡

```
SEXP ans;
double *da, *db, *dC, *dW, *dans, dtol = REAL(tol)[0];
double *dcenter;
double mdtol = 1.0 - dtol;
double d0, e0, emd0, f0, q0;
◇
```

Fragment referenced in [75](#), [86](#).

⟨ *R lpmvnorm 75* ⟩ ≡

```
SEXP R_lpmvnorm(SEXP a, SEXP b, SEXP C, SEXP center, SEXP N, SEXP J,  
                SEXP W, SEXP M, SEXP tol, SEXP logLik, SEXP fast) {
```

```
  ⟨ R slpmvnorm variables 74d ⟩
```

```
  double l0, lM, x0, intsum;  
  int p, len;
```

```
  Rboolean RlogLik = asLogical(logLik);
```

```
  ⟨ pnorm 73a ⟩
```

```
  ⟨ dimensions 73c ⟩
```

```
  ⟨ W length 73b ⟩
```

```
  ⟨ init center 74c ⟩
```

```
  int start, j, k;
```

```
  double tmp, Wtmp, e, d, f, emd, x, y[(iJ > 1 ? iJ - 1 : 1)];
```

```
  ⟨ setup return object 74a ⟩
```

```
  q0 = qnorm(dtol, 0.0, 1.0, 1L, 0L);
```

```
  l0 = log(dtol);
```

```
  ⟨ univariate problem 74b ⟩
```

```
  if (W == R_NilValue)
```

```
    GetRNGstate();
```

```
  for (int i = 0; i < iN; i++) {
```

```
    x0 = 0;
```

```
    ⟨ initialisation 69b ⟩
```

```
    if (W != R_NilValue && pW == 0)
```

```
      dW = REAL(W);
```

```
    for (int m = 0; m < iM; m++) {
```

```
      ⟨ init logLik loop 69c ⟩
```

```
      ⟨ inner logLik loop 71b ⟩
```

```
      ⟨ increment 71c ⟩
```

```
      if (W != R_NilValue)
```

```
        dW += iJ - 1;
```

```
    }
```

```
    ⟨ output 71d ⟩
```

```
    ⟨ move on 72a ⟩
```

```
  }
```

```
  if (W == R_NilValue)
```

```
    PutRNGstate();
```

```
  UNPROTECT(1);
```

```
  return(ans);
```

```
  }
```

```
  ◇
```

Fragment referenced in 67.

The R user interface consists of some checks and a call to C. Note that we need to specify both w and M in case we want a new set of weights for each observation.

⟨ init random seed, reset on exit 76a ⟩ ≡

```
### from stats::simulate.lm
if (!exists(".Random.seed", envir = .GlobalEnv, inherits = FALSE))
  runif(1)
if (is.null(seed))
  RNGstate <- get(".Random.seed", envir = .GlobalEnv)
else {
  R.seed <- get(".Random.seed", envir = .GlobalEnv)
  set.seed(seed)
  RNGstate <- structure(seed, kind = as.list(RNGkind()))
  on.exit(assign(".Random.seed", R.seed, envir = .GlobalEnv))
}
◇
```

Fragment referenced in 77, 89.

⟨ check and / or set integration weights 76b ⟩ ≡

```
if (!is.null(w) && J > 1) {
  stopifnot(is.matrix(w))
  stopifnot(nrow(w) == J - 1)
  if (is.null(M))
    M <- ncol(w)
  stopifnot(ncol(w) %in% c(M, M * N))
  if (!is.double(w)) storage.mode(w) <- "double"
} else {
  if (J > 1) {
    if (is.null(M)) stop("either w or M must be specified")
  } else {
    M <- 1L
  }
}
◇
```

Fragment referenced in 77, 89.

Sometimes we want to evaluate the log-likelihood based on $\mathbf{L} = \mathbf{C}^{-1}$, the inverse Cholesky factor of the covariance matrix. In this case, we explicitly invert \mathbf{L} to give \mathbf{C} (both matrices are lower triangular, so this is fast).

⟨ Cholesky of precision 76c ⟩ ≡

```
stopifnot(xor(missing(chol), missing(invchol)))
if (missing(chol)) chol <- solve(invchol)
◇
```

Fragment referenced in 77, 89.

`<lpmvnorm 77> ≡`

```
lpmvnorm <- function(lower, upper, mean = 0, center = NULL, chol, invchol,
                    logLik = TRUE, M = NULL, w = NULL, seed = NULL,
                    tol = .Machine$double.eps, fast = FALSE) {

  <init random seed, reset on exit 76a>
  <Cholesky of precision 76c>
  <input checks 68>
  <standardise 69a>
  <check and / or set integration weights 76b>

  ret <- .Call(mvtnorm_R_lpmvnorm, ac, bc, uC, as.double(center),
              as.integer(N), as.integer(J), w, as.integer(M), as.double(tol),
              as.logical(logLik), as.logical(fast));

  return(ret)
}
◇
```

Fragment referenced in 66.

Coming back to our simple example, we get (with 25000 simple Monte-Carlo iterations)

```
> phat
```

```
[1] 0.2369 0.2337 0.2842 0.3915 0.4662 0.0000 0.5901 0.4619 0.4873 0.0000
```

```
> exp(lpmvnorm(a, b, chol = lx, M = 25000, logLik = FALSE, fast = TRUE))
```

```
[1] 2.367e-01 2.341e-01 2.835e-01 3.939e-01 4.658e-01 8.882e-21 5.911e-01
[8] 4.598e-01 4.879e-01 8.882e-21
```

```
> exp(lpmvnorm(a, b, chol = lx, M = 25000, logLik = FALSE, fast = FALSE))
```

```
[1] 2.377e-01 2.372e-01 2.832e-01 3.875e-01 4.660e-01 8.882e-21 5.895e-01
[8] 4.624e-01 4.871e-01 8.882e-21
```

Next we generate some data and compare our implementation to `pmvnorm` using quasi-Monte-Carlo integration. The `pmvnorm` function uses randomised Korobov rules. The experiment here applies generalised Halton sequences. Plain Monte-Carlo (`w = NULL`) will also work but produces more variable results. Results will depend a lot on appropriate choices and it is the user's responsibility to make sure things work as intended. If you are unsure, you should use `pmvnorm` which provides a well-tested configuration.

```
> M <- 10000L
> if (require("qrng", quietly = TRUE)) {
+   ### quasi-Monte-Carlo
+   W <- t(ghalton(M, d = J - 1))
+ } else {
+   ### Monte-Carlo
+   ### byrow = TRUE because adding / removing dimensions
+   ### keeps the MC points for the remaining dimensions constant
+   W <- matrix(runif(M * (J - 1)), nrow = J - 1, byrow = TRUE)
+ }
> ### Genz & Bretz, 2002, without early stopping (really?)
> pGB <- lpmvnormR(a, b, chol = lx, logLik = FALSE,
+                 algorithm = GenzBretz(maxpts = M, abseps = 0, releps = 0))
```

```

> ### Genz 1992 with quasi-Monte-Carlo, fast pnorm
> pGqf <- exp(lpmvnorm(a, b, chol = lx, w = W, M = M, logLik = FALSE,
+                 fast = TRUE))
> ### Genz 1992, original Monte-Carlo, fast pnorm
> pGf <- exp(lpmvnorm(a, b, chol = lx, w = NULL, M = M, logLik = FALSE,
+                 fast = TRUE))
> ### Genz 1992 with quasi-Monte-Carlo, R::pnorm
> pGqs <- exp(lpmvnorm(a, b, chol = lx, w = W, M = M, logLik = FALSE,
+                 fast = FALSE))
> ### Genz 1992, original Monte-Carlo, R::pnorm
> pGs <- exp(lpmvnorm(a, b, chol = lx, w = NULL, M = M, logLik = FALSE,
+                 fast = FALSE))
> cbind(pGB, pGqf, pGf, pGqs, pGs)

```

```

      pGB      pGqf      pGf      pGqs      pGs
[1,] 0.2369 2.369e-01 2.345e-01 2.369e-01 2.360e-01
[2,] 0.2342 2.340e-01 2.319e-01 2.340e-01 2.347e-01
[3,] 0.2841 2.841e-01 2.851e-01 2.841e-01 2.870e-01
[4,] 0.3918 3.921e-01 3.932e-01 3.921e-01 3.904e-01
[5,] 0.4671 4.668e-01 4.679e-01 4.668e-01 4.691e-01
[6,] 0.0000 2.220e-20 2.220e-20 2.220e-20 2.220e-20
[7,] 0.5902 5.902e-01 5.908e-01 5.902e-01 5.929e-01
[8,] 0.4613 4.619e-01 4.612e-01 4.619e-01 4.630e-01
[9,] 0.4872 4.870e-01 4.863e-01 4.870e-01 4.821e-01
[10,] 0.0000 2.220e-20 2.220e-20 2.220e-20 2.220e-20

```

The three versions agree nicely. We now check if the code also works for univariate problems

```

> ### test univariate problem
> ### call pmvnorm
> pGB <- lpmvnormR(a[1,,drop = FALSE], b[1,,drop = FALSE], chol = lx[,1],
+                 logLik = FALSE,
+                 algorithm = GenzBretz(maxpts = M, abseps = 0, releps = 0))
> ### call lpmvnorm
> pGq <- exp(lpmvnorm(a[1,,drop = FALSE], b[1,,drop = FALSE], chol = lx[,1],
+                 logLik = FALSE))
> ### ground truth
> ptr <- pnorm(b[1,] / c(unclass(lx[,1]))) - pnorm(a[1,] / c(unclass(lx[,1])))
> cbind(c(ptr), pGB, pGq)

```

```

      pGB      pGq
[1,] 1.0000 1.0000 1.0000
[2,] 0.6109 0.6109 0.6109
[3,] 0.9076 0.9076 0.9076
[4,] 0.8980 0.8980 0.8980
[5,] 0.9589 0.9589 0.9589
[6,] 0.7863 0.7863 0.7863
[7,] 0.9983 0.9983 0.9983
[8,] 0.8745 0.8745 0.8745
[9,] 0.9386 0.9386 0.9386
[10,] 0.9120 0.9120 0.9120

```

Because the default `fast = FALSE` was used here, all results are identical.

3.2 Score Function

In addition to the log-likelihood, we would also like to have access to the scores with respect to C_i . Because every element of C_i only enters once, the chain rule rules, so to speak.

We need the derivatives of d , e , y , and f with respect to the c parameters

$\langle chol\ scores\ 79a \rangle \equiv$

```
double dp_c[Jp], ep_c[Jp], fp_c[Jp], yp_c[(iJ > 1 ? iJ - 1 : 1) * Jp];  
◇
```

Fragment referenced in 80a.

and the derivates with respect to the mean

$\langle mean\ scores\ 79b \rangle \equiv$

```
double dp_m[Jp], ep_m[Jp], fp_m[Jp], yp_m[(iJ > 1 ? iJ - 1 : 1) * Jp];  
◇
```

Fragment referenced in 80a.

and the derivates with respect to lower (a)

$\langle lower\ scores\ 79c \rangle \equiv$

```
double dp_l[Jp], ep_l[Jp], fp_l[Jp], yp_l[(iJ > 1 ? iJ - 1 : 1) * Jp];  
◇
```

Fragment referenced in 80a.

and the derivates with respect to upper (b)

$\langle upper\ scores\ 79d \rangle \equiv$

```
double dp_u[Jp], ep_u[Jp], fp_u[Jp], yp_u[(iJ > 1 ? iJ - 1 : 1) * Jp];  
◇
```

Fragment referenced in 80a.

and we start allocating the necessary memory. The output object contains the likelihood contributions (first row), the scores with respect to the mean (next J rows), with respect to the lower integration limits (next J rows), with respect to the upper integration limits (next J rows) and finally with respect to the off-diagonal elements of the Cholesky factor (last $J(J - 1)/2$ rows).

score output object 80a \equiv

```

int Jp = iJ * (iJ + 1) / 2;
< chol scores 79a >
< mean scores 79b >
< lower scores 79c >
< upper scores 79d >
double dtmp, etmp, Wtmp, ytmp, xx;

PROTECT(ans = allocMatrix(REALSXP, Jp + 1 + 3 * iJ, iN));
dans = REAL(ans);
for (j = 0; j < LENGTH(ans); j++) dans[j] = 0.0;
◇

```

Fragment referenced in 86.

For each $i = 1, \dots, N$, do

1. Input \mathbf{C}_i (chol), \mathbf{a}_i (lower), \mathbf{b}_i (upper), and control parameters α , ϵ , and M_{\max} (\mathbf{M}).
2. Standardise integration limits $a_j^{(i)}/c_{jj}^{(i)}$, $b_j^{(i)}/c_{jj}^{(i)}$, and rows $c_{jj}^{(i)}/c_{jj}^{(i)}$ for $1 \leq j < j < J$.

Note: We later need derivatives wrt $c_{jj}^{(i)}$, so we compute derivatives wrt $a_j^{(i)}$ and $b_j^{(i)}$ and post-differentiate later.

3. Initialise intsum = varsum = 0, $M = 0$, $d_1 = \Phi(a_1^{(i)})$, $e_1 = \Phi(b_1^{(i)})$ and $f_1 = e_1 - d_1$.

We start initialised the score wrt to $c_{11}^{(i)}$ (the parameter is non-existent here due to standardisation)

score c11 80b \equiv

```

if (LENGTH(center)) {
  dp_c[0] = (R_FINITE(da[0]) ? dnorm(da[0], x0, 1.0, 0L) * (da[0] - x0 - dcenter[0]) : 0);
  ep_c[0] = (R_FINITE(db[0]) ? dnorm(db[0], x0, 1.0, 0L) * (db[0] - x0 - dcenter[0]) : 0);
} else {
  dp_c[0] = (R_FINITE(da[0]) ? dnorm(da[0], x0, 1.0, 0L) * (da[0] - x0) : 0);
  ep_c[0] = (R_FINITE(db[0]) ? dnorm(db[0], x0, 1.0, 0L) * (db[0] - x0) : 0);
}
fp_c[0] = ep_c[0] - dp_c[0];
◇

```

Fragment referenced in 81b, 86.

score a, b 81a \equiv

```

dp_m[0] = (R_FINITE(da[0]) ? dnorm(da[0], x0, 1.0, 0L) : 0);
ep_m[0] = (R_FINITE(db[0]) ? dnorm(db[0], x0, 1.0, 0L) : 0);
dp_l[0] = dp_m[0];
ep_u[0] = ep_m[0];
dp_u[0] = 0;
ep_l[0] = 0;
fp_m[0] = ep_m[0] - dp_m[0];
fp_l[0] = -dp_m[0];
fp_u[0] = ep_m[0];
◇

```

Fragment referenced in 81b, 86.

4. Repeat

< init score loop 81b > \equiv

```

    < init logLik loop 69c >
    < score c11 80b >
    < score a, b 81a >
     $\diamond$ 

```

Fragment referenced in 86.

- (a) Generate uniform $w_1, \dots, w_{J-1} \in [0, 1]$.
- (b) For $j = 2, \dots, J$ set

$$y_{j-1} = \Phi^{-1}(d_{j-1} + w_{j-1}(e_{j-1} - d_{j-1}))$$

We again either generate w_{j-1} on the fly or use pre-computed weights (w). We first compute the scores with respect to the already existing parameters.

< update yp for chol 81c > \equiv

```

    ytmp = exp(- dnorm(y[j - 1], 0.0, 1.0, 1L)); // = 1 / dnorm(y[j - 1], 0.0, 1.0, 0L)

    for (k = 0; k < Jp; k++) yp_c[k * (iJ - 1) + (j - 1)] = 0.0;

    for (idx = 0; idx < (j + 1) * j / 2; idx++) {
        yp_c[idx * (iJ - 1) + (j - 1)] = ytmp;
        yp_c[idx * (iJ - 1) + (j - 1)] *= (dp_c[idx] + Wtmp * (ep_c[idx] - dp_c[idx]));
    }
     $\diamond$ 

```

Fragment referenced in 85a.

< update yp for means, lower and upper 82 > \equiv

```

    for (k = 0; k < iJ; k++)
        yp_m[k * (iJ - 1) + (j - 1)] = 0.0;

    for (idx = 0; idx < j; idx++) {
        yp_m[idx * (iJ - 1) + (j - 1)] = ytmp;
        yp_m[idx * (iJ - 1) + (j - 1)] *= (dp_m[idx] + Wtmp * (ep_m[idx] - dp_m[idx]));
    }

    for (k = 0; k < iJ; k++)
        yp_l[k * (iJ - 1) + (j - 1)] = 0.0;

    for (idx = 0; idx < j; idx++) {
        yp_l[idx * (iJ - 1) + (j - 1)] = ytmp;
        yp_l[idx * (iJ - 1) + (j - 1)] *= (dp_l[idx] + Wtmp * (dp_u[idx] - dp_l[idx]));
    }

    for (k = 0; k < iJ; k++)
        yp_u[k * (iJ - 1) + (j - 1)] = 0.0;

    for (idx = 0; idx < j; idx++) {
        yp_u[idx * (iJ - 1) + (j - 1)] = ytmp;
        yp_u[idx * (iJ - 1) + (j - 1)] *= (ep_l[idx] + Wtmp * (ep_u[idx] - ep_l[idx]));
    }
     $\diamond$ 

```

Fragment referenced in 85a.

$$x_{j-1} = \sum_{j=1}^{j-1} c_{jj}^{(i)} y_j$$

$$d_j = \Phi\left(a_j^{(i)} - x_{j-1}\right)$$

$$e_j = \Phi\left(b_j^{(i)} - x_{j-1}\right)$$

$$f_j = (e_j - d_j)f_{j-1}.$$

The scores with respect to $c_{jj}^{(i)}, j = 1, \dots, j-1$ are

(score wrt new chol off-diagonals 83a) \equiv

```

dtmp = dnorm(da[j], x, 1.0, 0L);
etmp = dnorm(db[j], x, 1.0, 0L);

for (k = 0; k < j; k++) {
  idx = start + j + k;
  if (LENGTH(center)) {
    dp_c[idx] = dtmp * (-1.0) * (y[k] - dcenter[k]);
    ep_c[idx] = etmp * (-1.0) * (y[k] - dcenter[k]);
  } else {
    dp_c[idx] = dtmp * (-1.0) * y[k];
    ep_c[idx] = etmp * (-1.0) * y[k];
  }
  fp_c[idx] = (ep_c[idx] - dp_c[idx]) * f;
}

```

Fragment referenced in [85a](#).

and the score with respect to (the here non-existing) $c_{jj}^{(i)}$ is

(score wrt new chol diagonal 83b) \equiv

```

idx = (j + 1) * (j + 2) / 2 - 1;
if (LENGTH(center)) {
  dp_c[idx] = (R_FINITE(da[j]) ? dtmp * (da[j] - x - dcenter[j]) : 0);
  ep_c[idx] = (R_FINITE(db[j]) ? etmp * (db[j] - x - dcenter[j]) : 0);
} else {
  dp_c[idx] = (R_FINITE(da[j]) ? dtmp * (da[j] - x) : 0);
  ep_c[idx] = (R_FINITE(db[j]) ? etmp * (db[j] - x) : 0);
}
fp_c[idx] = (ep_c[idx] - dp_c[idx]) * f;

```

Fragment referenced in [85a](#).

< new score means, lower and upper 83c > ≡

```
dp_m[j] = (R_FINITE(da[j]) ? dtmp : 0);
ep_m[j] = (R_FINITE(db[j]) ? etmp : 0);
dp_l[j] = dp_m[j];
ep_u[j] = ep_m[j];
dp_u[j] = 0;
ep_l[j] = 0;
fp_l[j] = - dp_m[j] * f;
fp_u[j] = ep_m[j] * f;
fp_m[j] = fp_u[j] + fp_l[j];
◇
```

Fragment referenced in [85a](#).

We next update scores for parameters introduced for smaller j

< update score for chol 84a > ≡

```
for (idx = 0; idx < j * (j + 1) / 2; idx++) {
  xx = 0.0;
  for (k = 0; k < j; k++)
    xx += dC[start + k] * yp_c[idx * (iJ - 1) + k];

  dp_c[idx] = dtmp * (-1.0) * xx;
  ep_c[idx] = etmp * (-1.0) * xx;
  fp_c[idx] = (ep_c[idx] - dp_c[idx]) * f + emd * fp_c[idx];
}
◇
```

Fragment referenced in [85a](#).

< update score means, lower and upper 84b > ≡

```
for (idx = 0; idx < j; idx++) {
  xx = 0.0;
  for (k = 0; k < j; k++)
    xx += dC[start + k] * yp_m[idx * (iJ - 1) + k];

  dp_m[idx] = dtmp * (-1.0) * xx;
  ep_m[idx] = etmp * (-1.0) * xx;
  fp_m[idx] = (ep_m[idx] - dp_m[idx]) * f + emd * fp_m[idx];
}

for (idx = 0; idx < j; idx++) {
  xx = 0.0;
  for (k = 0; k < j; k++)
    xx += dC[start + k] * yp_l[idx * (iJ - 1) + k];

  dp_l[idx] = dtmp * (-1.0) * xx;
  dp_u[idx] = etmp * (-1.0) * xx;
  fp_l[idx] = (dp_u[idx] - dp_l[idx]) * f + emd * fp_l[idx];
}

for (idx = 0; idx < j; idx++) {
  xx = 0.0;
  for (k = 0; k < j; k++)
    xx += dC[start + k] * yp_u[idx * (iJ - 1) + k];

  ep_l[idx] = dtmp * (-1.0) * xx;
  ep_u[idx] = etmp * (-1.0) * xx;
  fp_u[idx] = (ep_u[idx] - ep_l[idx]) * f + emd * fp_u[idx];
}
◇
```

Fragment referenced in 85a.

We put everything together in a loop starting with the second dimension

< inner score loop 85a > ≡

```
for (j = 1; j < iJ; j++) {

  < compute y 70a >
  < compute x 70b >
  < update d, e 70c >
  < update yp for chol 81c >
  < update yp for means, lower and upper 82 >
  < score wrt new chol off-diagonals 83a >
  < score wrt new chol diagonal 83b >
  < new score means, lower and upper 83c >
  < update score for chol 84a >
  < update score means, lower and upper 84b >
  < update f 71a >

}
◇
```

Fragment referenced in 86.

(c) Set $\text{intsum} = \text{intsum} + f_J$, $\text{varsum} = \text{varsum} + f_J^2$, $M = M + 1$, and $\text{error} = \sqrt{(\text{varsum}/M - (\text{intsum}/M)^2)/M}$.

We refrain from early stopping and error estimation.

Until $\text{error} < \epsilon$ or $M = M_{\max}$

5. Output $\hat{p}_i = \text{intsum}/M$.

We return $\log \hat{p}_i$ for each i , or we immediately sum-up over i .

< score output 85b > \equiv

```

dans[0] += f;
for (j = 0; j < Jp; j++)
  dans[j + 1] += fp_c[j];
for (j = 0; j < iJ; j++) {
  idx = Jp + j + 1;
  dans[idx] += fp_m[j];
  dans[idx + iJ] += fp_l[j];
  dans[idx + 2 * iJ] += fp_u[j];
}
◇

```

Fragment referenced in [86](#).

< init dans 85c > \equiv

```

if (iM == 0) {
  dans[0] = intsum;
  dans[1] = fp_c[0];
  dans[2] = fp_m[0];
  dans[3] = fp_l[0];
  dans[4] = fp_u[0];
}
◇

```

Fragment referenced in [86](#).

We put everything together in C

< R slpmvnorm 86 > ≡

```
SEXP R_slpmvnorm(SEXP a, SEXP b, SEXP C, SEXP center, SEXP N, SEXP J, SEXP W,  
                SEXP M, SEXP tol, SEXP fast) {
```

```
    < R slpmvnorm variables 74d >  
    double intsum;  
    int p, idx;  
    < dimensions 73c >  
    < pnorm 73a >  
    < W length 73b >  
    < init center 74c >  
    int start, j, k;  
    double tmp, e, d, f, emd, x, x0, y[(iJ > 1 ? iJ - 1 : 1)];  
  
    < score output object 80a >  
  
    q0 = qnorm(dtol, 0.0, 1.0, 1L, 0L);  
  
    /* univariate problem */  
    if (iJ == 1) iM = 0;  
  
    if (W == R_NilValue)  
        GetRNGstate();  
  
    for (int i = 0; i < iN; i++) {  
  
        < initialisation 69b >  
        < score c11 80b >  
        < score a, b 81a >  
        < init dans 85c >  
  
        if (W != R_NilValue && pW == 0)  
            dW = REAL(W);  
  
        for (int m = 0; m < iM; m++) {  
            < init score loop 81b >  
            < inner score loop 85a >  
            < score output 85b >  
            if (W != R_NilValue)  
                dW += iJ - 1;  
        }  
  
        < move on 72a >  
        dans += Jp + 1 + 3 * iJ;  
    }  
  
    if (W == R_NilValue)  
        PutRNGstate();  
  
    UNPROTECT(1);  
    return(ans);  
}  
◇
```

Fragment referenced in [67](#).

The R code is now essentially identical to `lpmvnorm`, however, we need to undo the effect of

standardisation once the scores have been computed

⟨ post differentiate mean score 87a ⟩ ≡

```

Jp <- J * (J + 1) / 2;
smean <- -ret[Jp + 1:J, , drop = FALSE]
if (attr(chol, "diag"))
  smean <- smean / c(dchol)

```

Fragment referenced in 89.

⟨ post differentiate lower score 87b ⟩ ≡

```

slower <- ret[Jp + J + 1:J, , drop = FALSE]
if (attr(chol, "diag"))
  slower <- slower / c(dchol)

```

Fragment referenced in 89.

⟨ post differentiate upper score 87c ⟩ ≡

```

supper <- ret[Jp + 2 * J + 1:J, , drop = FALSE]
if (attr(chol, "diag"))
  supper <- supper / c(dchol)

```

Fragment referenced in 89.

⟨ post differentiate chol score 87d ⟩ ≡

```

if (J == 1) {
  idx <- 1L
} else {
  idx <- cumsum(c(1, 2:J))
}
if (attr(chol, "diag")) {
  ret <- ret / c(dchol[rep(1:J, 1:J),]) ### because 1 / dchol already there
  ret[idx,] <- -ret[idx,]
}

```

Fragment referenced in 89.

We sometimes parameterise models in terms of $\mathbf{L} = \mathbf{C}^{-1}$, the Cholesky factor of the precision matrix. The log-likelihood operates on \mathbf{C} , so we need to post-differentiate the score function. We have

$$\mathbf{A} = \frac{\partial \mathbf{L}^{-1}}{\partial \mathbf{L}} = -\mathbf{L}^{-\top} \otimes \mathbf{L}^{-1}$$

and computing \mathbf{sA} for a score vector \mathbf{s} with respect to \mathbf{L} can be implemented by the “vec-trick” (Section 2.11)

$$\mathbf{sA} = \mathbf{L}^{-\top} \mathbf{S} \mathbf{L}^{-\top}$$

where $\mathbf{s} = \text{vec}(\mathbf{S})$.

< post differentiate invchol score 88a > ≡

```
if (!missing(invchol)) {
  ret <- ltMatrices(ret, diag = TRUE, byrow = TRUE,
                   names = dimnames(chol)[[2L]])
  ### this means vectrick(chol, ret, chol)
  ret <- - unclass(vectrick(chol, ret))
}
◇
```

Fragment referenced in [89](#).

If the diagonal elements are constants, we set them to zero. The function always returns an object of class `ltMatrices` with explicit diagonal elements (use `Lower_tri(, diag = FALSE)` to extract the lower triangular elements such that the scores match the input)

< post process score 88b > ≡

```
if (!attr(chol, "diag"))
  ### remove scores for constant diagonal elements
  ret[idx,] <- 0
ret <- ltMatrices(ret, diag = TRUE, byrow = TRUE,
                 names = dimnames(chol)[[2L]])
◇
```

Fragment referenced in [89](#).

We can now finally put everything together in a single score function.

`< slpmvnorm 89 > ≡`

```
slpmvnorm <- function(lower, upper, mean = 0, center = NULL,
                     chol, invchol, logLik = TRUE, M = NULL,
                     w = NULL, seed = NULL, tol = .Machine$double.eps,
                     fast = FALSE) {

  < init random seed, reset on exit 76a >
  < Cholesky of precision 76c >
  < input checks 68 >
  < standardise 69a >
  < check and / or set integration weights 76b >

  ret <- .Call(mvtnorm_R_slpmvnorm, ac, bc, uC, as.double(center), as.integer(N),
              as.integer(J), w, as.integer(M), as.double(tol), as.logical(fast));

  ll <- log(pmax(ret[1L,], tol)) - log(M)
  intsum <- ret[1L,]
  m <- matrix(intsum, nrow = nrow(ret) - 1, ncol = ncol(ret), byrow = TRUE)
  ret <- ret[-1L,,drop = FALSE] / m
  ### NOTE: division by zero may have happened for observations with
  ### probability < tol; the log-lik is constant in these cases
  ### and thus the derivative = 0
  ret[m < tol] <- 0

  < post differentiate mean score 87a >
  < post differentiate lower score 87b >
  < post differentiate upper score 87c >

  ret <- ret[1:Jp, , drop = FALSE]

  < post differentiate chol score 87d >
  < post differentiate invchol score 88a >
  < post process score 88b >

  ret <- ltMatrices(ret, byrow = byrow_orig)

  rownames(smean) <- rownames(slower) <-
    rownames(supper) <- dimnames(chol)[[2L]]

  if (logLik) {
    ret <- list(logLik = ll,
               mean = smean,
               lower = slower,
               upper = supper,
               chol = ret)
    if (!missing(invchol)) names(ret)[names(ret) == "chol"] <- "invchol"
    return(ret)
  }

  return(ret)
}
◇
```

Fragment referenced in [66](#).

Let's look at an example, where we use `numDeriv::grad` to check the results (this functionality from package `numDeriv` was absolutely instrumental for this project)

```

> J <- 5L
> N <- 4L
> S <- crossprod(matrix(runif(J^2), nrow = J))
> prm <- t(chol(S))[lower.tri(S, diag = TRUE)]
> ### define C
> mC <- ltMatrices(matrix(prm, ncol = 1), diag = TRUE)
> a <- matrix(runif(N * J), nrow = J) - 2
> b <- a + 4
> a[2,] <- -Inf
> b[3,] <- Inf
> M <- 10000L
> W <- matrix(runif(M * (J - 1)), ncol = M)
> lli <- c(lpmvnorm(a, b, chol = mC, w = W, M = M, logLik = FALSE))
> fC <- function(prm) {
+   C <- ltMatrices(matrix(prm, ncol = 1), diag = TRUE)
+   lpmvnorm(a, b, chol = C, w = W, M = M)
+ }
> sC <- slpmvnorm(a, b, chol = mC, w = W, M = M)
> chk(lli, sC$logLik)
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(fC, unclass(mC)), rowSums(unclass(sC$chol)),
+       check.attributes = FALSE)

```

We can do the same when **L** (and not **C**) is given

```

> mL <- solve(mC)
> lliL <- c(lpmvnorm(a, b, invchol = mL, w = W, M = M, logLik = FALSE))
> chk(lli, lliL)
> fL <- function(prm) {
+   L <- ltMatrices(matrix(prm, ncol = 1), diag = TRUE)
+   lpmvnorm(a, b, invchol = L, w = W, M = M)
+ }
> sL <- slpmvnorm(a, b, invchol = mL, w = W, M = M)
> chk(lliL, sL$logLik)
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(fL, unclass(mL)), rowSums(unclass(sL$invchol)),
+       check.attributes = FALSE)

```

The score function also works for univariate problems

```

> ptr <- pnorm(b[1,] / c(unclass(mC[,1]))) - pnorm(a[1,] / c(unclass(mC[,1])))
> log(ptr)

[1] -0.01166 -0.08617 -0.01240 -0.03105

> lpmvnorm(a[1,,drop = FALSE], b[1,,drop = FALSE], chol = mC[,1], logLik = FALSE)

[1] -0.01166 -0.08617 -0.01240 -0.03105

> lapply(slpvmnorm(a[1,,drop = FALSE], b[1,,drop = FALSE], chol = mC[,1],
+                 logLik = TRUE), unclass)

$logLik
[1] -0.01166 -0.08617 -0.01240 -0.03105

$mean

```

```

      [,1] [,2] [,3] [,4]
1 0.02222 0.214 0.02642 0.08861

$lower
      [,1] [,2] [,3] [,4]
1 -0.03222 -0.2145 -0.03536 -0.09096

$upper
      [,1] [,2] [,3] [,4]
1 0.009995 0.0004369 0.008944 0.002351

$chol
      [,1] [,2] [,3] [,4]
1.1 -0.1041 -0.2994 -0.1076 -0.1787
attr("J")
[1] 1
attr("diag")
[1] TRUE
attr("byrow")
[1] FALSE
attr("rcnames")
[1] "1"

> sd1 <- c(unclass(mC[,1]))
> (dnorm(b[1,] / sd1) * b[1,] - dnorm(a[1,] / sd1) * a[1,]) * (-1) / sd1^2 / ptr

[1] -0.1041 -0.2994 -0.1076 -0.1787

```

Chapter 4

Maximum-likelihood Example

We now discuss how this infrastructure can be used to estimate the Cholesky factor of a multivariate normal in the presence of interval-censored observations.

We first generate a covariance matrix $\Sigma = \mathbf{C}\mathbf{C}^\top$ and extract the Cholesky factor \mathbf{C}

```
> J <- 4
> R <- diag(J)
> R[1,2] <- R[2,1] <- .25
> R[1,3] <- R[3,1] <- .5
> R[2,4] <- R[4,2] <- .75
> Sigma <- diag(sqrt(1:J / 2)) %*% R %*% diag(sqrt(1:J / 2))
> C <- t(chol(Sigma))
```

We now represent this matrix as `ltMatrices` object

```
> prm <- C[lower.tri(C, diag = TRUE)]
> lt <- ltMatrices(matrix(prm, ncol = 1L),
+                   diag = TRUE,    ### has diagonal elements
+                   byrow = FALSE)  ### prm is column-major
> BYROW <- FALSE    ### later checks
> lt <- ltMatrices(lt,
+                   byrow = BYROW)  ### convert to row-major
> chk(C, as.array(lt)[, , 1], check.attributes = FALSE)
> chk(Sigma, as.array(tcrossprod(lt))[, , 1], check.attributes = FALSE)
```

We generate some data from $\mathbb{N}_J(\mathbf{0}_J, \Sigma)$ by first sampling from $\mathbf{Z} \sim \mathbb{N}_J(\mathbf{0}_J, \mathbf{I}_J)$ and then computing $\mathbf{Y} = \mathbf{C}\mathbf{Z} + \boldsymbol{\mu} \sim \mathbb{N}_J(\boldsymbol{\mu}, \mathbf{C}\mathbf{C}^\top)$

```
> N <- 100L
> Z <- matrix(rnorm(N * J), nrow = J)
> Y <- lt %*% Z + (mn <- 1:J)
```

Before we add some interval-censoring to the data, let's estimate the Cholesky factor \mathbf{C} (here called `lt`) from the raw continuous data. The true mean $\boldsymbol{\mu}$ and the true covariance matrix Σ can be estimated from the uncensored data via maximum likelihood as

```
> rowMeans(Y)

      1      2      3      4
0.9685 2.1269 2.9634 3.9826

> (Shat <- var(t(Y)) * (N - 1) / N)
```

```

      1      2      3      4
1 0.46656 0.18104 0.34222 0.01609
2 0.18104 0.94385 0.08992 0.84310
3 0.34222 0.08992 1.36055 0.08104
4 0.01609 0.84310 0.08104 1.63302

```

We first check if we can obtain the same results by numerical optimisation using `dmvnorm` and the scores `sldmvnorm`. The log-likelihood and the score function (for the centered means) in terms of \mathbf{C} are

```

> Yc <- Y - rowMeans(Y)
> ll <- function(parm) {
+   C <- ltMatrices(parm, diag = TRUE, byrow = BYROW)
+   -ldmvnorm(obs = Yc, chol = C)
+ }
> sc <- function(parm) {
+   C <- ltMatrices(parm, diag = TRUE, byrow = BYROW)
+   -rowSums(unclass(sldmvnorm(obs = Yc, chol = C)$chol))
+ }

```

The diagonal elements of \mathbf{C} are positive, so we need box constraints

```

> llim <- rep(-Inf, J * (J + 1) / 2)
> llim[which(rownames(unclass(lt)) %in% paste(1:J, 1:J, sep = "."))] <- 1e-4

```

The ML-estimate of $\mathbf{C}\mathbf{C}^\top$ is now used to obtain an estimate of \mathbf{C} and we check the score function for some random starting values

```

> if (BYROW) {
+   cML <- chol(Shat)[upper.tri(Shat, diag = TRUE)]
+ } else {
+   cML <- t(chol(Shat))[lower.tri(Shat, diag = TRUE)]
+ }
> ll(cML)

[1] 517.9

> start <- runif(length(cML))
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll, start), sc(start), check.attributes = FALSE)

```

Finally, we hand over to `optim` and compare the results of the analytically and numerically obtained ML estimates

```

> op <- optim(start, fn = ll, gr = sc, method = "L-BFGS-B",
+             lower = llim, control = list(trace = FALSE))
> ## ML numerically
> ltMatrices(op$par, diag = TRUE, byrow = BYROW)

, , 1

```

```

      1      2      3      4
1 0.68306      .      .      .
2 0.26505 0.93465      .      .
3 0.50102 -0.04587 1.05234      .
4 0.02356 0.89535 0.10482 0.90544

```

```

> ll(op$par)

[1] 517.9

> ## ML analytically
> t(chol(Shat))

      1      2      3      4
1 0.68305 0.00000 0.0000 0.0000
2 0.26505 0.93467 0.0000 0.0000
3 0.50102 -0.04587 1.0523 0.0000
4 0.02356 0.89535 0.1048 0.9054

> ll(cML)

[1] 517.9

> ## true C matrix
> lt

, , 1

      1      2      3      4
1 0.7071      .      .      .
2 0.2500 0.9682      .      .
3 0.6124 -0.1581 1.0488      .
4      . 1.0954 0.1651 0.8790

```

Under interval-censoring, the mean and \mathbf{C} are no longer orthogonal and there is no analytic solution to the ML estimation problem. So, we add some interval-censoring represented by `lwr` and `upr` and try to estimate the model parameters via `lpmvnorm` and corresponding scores `slpmvnorm`.

```

> prb <- 1:9 / 10
> sds <- sqrt(diag(Sigma))
> ct <- sapply(1:J, function(j) qnorm(prb, mean = mn[j], sd = sds[j]))
> lwr <- upr <- Y
> for (j in 1:J) {
+   f <- cut(Y[j,], breaks = c(-Inf, ct[,j], Inf))
+   lwr[j,] <- c(-Inf, ct[,j])[f]
+   upr[j,] <- c(ct[,j], Inf)[f]
+ }

```

Let's do some sanity and performance checks first. For different values of M , we evaluate the log-likelihood using `pmvnorm` (called in `lpmvnormR`) and the simplified implementation (fast and slow). The comparison is a bit unfair, because we do not add the time needed to setup Halton sequences, but we would do this only once and use the stored values for repeated evaluations of a log-likelihood (because the optimiser expects a deterministic function to be optimised)

```

> M <- floor(exp(0:25/10) * 1000)
> lGB <- sapply(M, function(m) {
+   st <- system.time(ret <-
+     lpmvnormR(lwr, upr, mean = mn, chol = lt, algorithm =
+       GenzBretz(maxpts = m, abseps = 0, releps = 0)))
+   return(c(st["user.self"], ll = ret))
+ })
> lH <- sapply(M, function(m) {

```

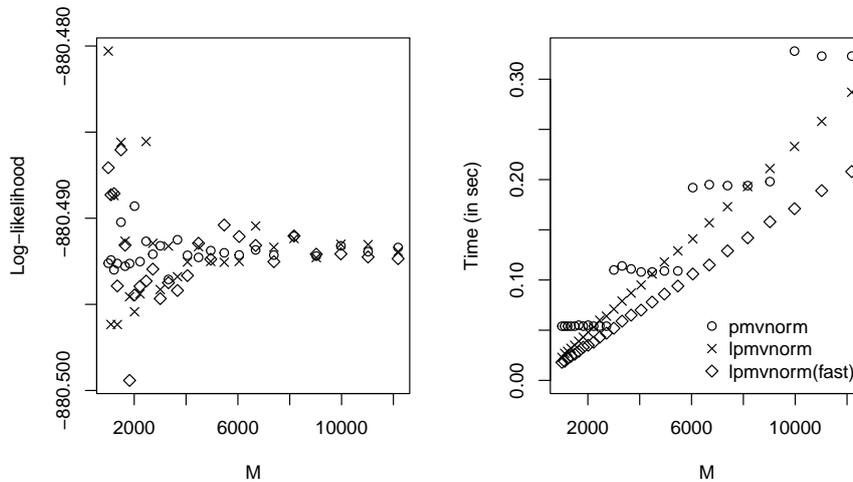


Figure 4.1: Evaluated log-likelihoods (left) and timings (right).

```

+   W <- NULL
+   if (require("qrng", quietly = TRUE))
+     W <- t(ghalton(m, d = J - 1))
+   st <- system.time(ret <- lpmvnorm(lwr, upr, mean = mn,
+                                   chol = lt, w = W, M = m))
+   return(c(st["user.self"], ll = ret))
+ })
> lHf <- sapply(M, function(m) {
+   W <- NULL
+   if (require("qrng", quietly = TRUE))
+     W <- t(ghalton(m, d = J - 1))
+   st <- system.time(ret <- lpmvnorm(lwr, upr, mean = mn, chol = lt,
+                                     w = W, M = m, fast = TRUE))
+   return(c(st["user.self"], ll = ret))
+ })

```

The evaluated log-likelihoods and corresponding timings are given in Figure 4.1. It seems that for $M \geq 3000$, results are reasonably stable.

We now define the log-likelihood function. It is important to use weights via the `w` argument (or to set the `seed`) such that only the candidate parameters `parm` change with repeated calls to `ll`. We use an extremely low number of integration points `M`, let's see if this still works out.

```

> M <- 500
> if (require("qrng", quietly = TRUE)) {
+   ### quasi-Monte-Carlo
+   W <- t(ghalton(M, d = J - 1))
+ } else {
+   ### Monte-Carlo
+   W <- matrix(runif(M * (J - 1)), nrow = J - 1, byrow = TRUE)
+ }
> ll <- function(parm, J) {
+   m <- parm[1:J]           ### mean parameters

```

```

+   parm <- parm[-(1:J)]      ### chol parameters
+   C <- matrix(c(parm), ncol = 1L)
+   C <- ltMatrices(C, diag = TRUE, byrow = BYROW)
+   -lpmvnorm(lower = lwr, upper = upr, mean = m, chol = C,
+             w = W, M = M, logLik = TRUE)
+ }

```

We can check the correctness of our log-likelihood function

```

> prm <- c(mn, unclass(lt))
> ll(prm, J = J)

[1] 880.5

> ### ATLAS gives -880.4908, M1mac gives -880.4911
> round(lpmvnormR(lwr, upr, mean = mn, chol = lt,
+               algorithm = GenzBretz(maxpts = M, abseps = 0, releps = 0)), 3)

[1] -880.5

> (llprm <- lpmvnorm(lwr, upr, mean = mn, chol = lt, w = W, M = M))

[1] -880.5

> chk(llprm, sum(lpmvnorm(lwr, upr, mean = mn, chol = lt, w = W,
+                       M = M, logLik = FALSE)))

```

Before we hand over to the optimiser, we define the score function with respect to μ and \mathbf{C}

```

> sc <- function(parm, J) {
+   m <- parm[1:J]          ### mean parameters
+   parm <- parm[-(1:J)]   ### chol parameters
+   C <- matrix(c(parm), ncol = 1L)
+   C <- ltMatrices(C, diag = TRUE, byrow = BYROW)
+   ret <- slpmvnorm(lower = lwr, upper = upr, mean = m, chol = C,
+                   w = W, M = M, logLik = TRUE)
+   return(-c(rowSums(ret$mean), rowSums(unclass(ret$chol))))
+ }

```

and check the correctness numerically

```

> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll, prm, J = J), sc(prm, J = J), check.attributes = FALSE)

```

Finally, we can hand-over to `optim`. Because we need $\text{diag}(\mathbf{C}) > 0$, we use box constraints and `method = "L-BFGS-B"`. We start with the estimates obtained from the original continuous data.

```

> llim <- rep(-Inf, J + J * (J + 1) / 2)
> llim[J + which(rownames(unclass(lt)) %in% paste(1:J, 1:J, sep = "."))] <- 1e-4
> if (BYROW) {
+   start <- c(rowMeans(Y), chol(Shat)[upper.tri(Shat, diag = TRUE)])
+ } else {
+   start <- c(rowMeans(Y), t(chol(Shat))[lower.tri(Shat, diag = TRUE)])
+ }
> ll(start, J = J)

[1] 875.4

```

```
> op <- optim(start, fn = ll, gr = sc, J = J, method = "L-BFGS-B",
+           lower = llim, control = list(trace = FALSE))
> op$value ## compare with
```

```
[1] 874.2
```

```
> ll(prm, J = J)
```

```
[1] 880.5
```

We can now compare the true and estimated Cholesky factor \mathbf{C} of our covariance matrix $\Sigma = \mathbf{C}\mathbf{C}^\top$

```
> (C <- ltMatrices(matrix(op$par[-(1:J)], ncol = 1),
+                 diag = TRUE, byrow = BYROW))
```

```
, , 1
```

	1	2	3	4
1	0.67050	.	.	.
2	0.26764	1.02232	.	.
3	0.54268	-0.05007	1.11348	.
4	0.05223	0.98430	0.08473	0.96137

```
> lt
```

```
, , 1
```

	1	2	3	4
1	0.7071	.	.	.
2	0.2500	0.9682	.	.
3	0.6124	-0.1581	1.0488	.
4	.	1.0954	0.1651	0.8790

and the estimated means

```
> op$par[1:J]
```

	1	2	3	4
	0.967	2.128	2.945	3.989

```
> mn
```

```
[1] 1 2 3 4
```

We can also compare the results on the scale of the covariance matrix

```
> ### ATLAS print issues
> round(tcrossprod(lt), 4) ### true Sigma
```

```
, , 1
```

	1	2	3	4
1	0.5000	0.1768	0.433	0.000
2	0.1768	1.0000	0.000	1.061
3	0.4330	0.0000	1.500	0.000
4	0.0000	1.0607	0.000	2.000

```

> tcrossprod(C)          ### interval-censored obs
, , 1
      1      2      3      4
1 0.44956 0.17945 0.36386 0.03502
2 0.17945 1.11677 0.09406 1.02025
3 0.36386 0.09406 1.53684 0.07341
4 0.03502 1.02025 0.07341 1.90298

> Shat                  ### "exact" obs
      1      2      3      4
1 0.46656 0.18104 0.34222 0.01609
2 0.18104 0.94385 0.08992 0.84310
3 0.34222 0.08992 1.36055 0.08104
4 0.01609 0.84310 0.08104 1.63302

```

This looks reasonably close.

Warning: Do NOT assume the choices made here (especially M and W) to be universally applicable. Make sure to investigate the accuracy depending on these parameters of the log-likelihood and score function in your application.

One could ask what this whole exercise was about statistically. We estimated a multivariate normal distribution from interval-censored data, so what? Maybe we were primarily interested in fitting a linear regression

$$\mathbb{E}(Y_1 | Y_j = y_j, j = 2, \dots, J) = \alpha + \sum_{j=2}^J \beta_j y_j.$$

Interval-censoring in the response could have been handled by some Tobit model, but what about interval-censoring in the explanatory variables? Based on the multivariate distribution just estimated, we can obtain the regression coefficients β_j as

```

> c(cond_mvnorm(chol = C, which_given = 2:J, given = diag(J - 1))$mean)
[1] 0.2602 0.2270 -0.1299

```

Alternatively, we can compute these regressions from a permuted Cholesky factor (this goes into the “simple” conditional distribution in Section 2.13)

```

> c(cond_mvnorm(chol = aperm(as.chol(C), perm = c(2:J, 1)),
+               which_given = 1:(J - 1), given = diag(J - 1))$mean)
[1] 0.2602 0.2270 -0.1299

```

or, as a third option, from the last row of the precision matrix of the permuted Cholesky factor

```

> x <- as.array(chol2pre(aperm(as.chol(C), perm = c(2:J, 1))))[J, 1]
> c(-x[-J] / x[J])
      2      3      4
0.2602 0.2270 -0.1299

```

In higher dimensions, the first option is to be preferred, because it only involves computing the Cholesky decomposition of a $(J - 1) \times (J - 1)$ matrix, whereas the latter two options are based on a decomposition of the full $J \times J$ covariance matrix.

We can compare these estimated regression coefficients with those obtained from a linear model fitted to the exact observations

```
> dY <- as.data.frame(t(Y))
> colnames(dY) <- paste0("Y", 1:J)
> coef(m1 <- lm(Y1 ~ ., data = dY))[-1L]
```

```
      Y2      Y3      Y4
0.3169 0.2405 -0.1657
```

The estimates are quite close, but what about standard errors? Interval-censoring means loss of information, so we should see larger standard errors for the interval-censored data.

Let's obtain the Hessian for all parameters first

```
> H <- optim(op$par, fn = ll, gr = sc, J = J, method = "L-BFGS-B",
+          lower = llim, hessian = TRUE,
+          control = list(trace = FALSE))$hessian
```

and next we sample from the distribution of the maximum-likelihood estimators

```
> L <- try(t(chol(H)))
> ### some check on r-oldrel-macos-arm64
> if (inherits(L, "try-error"))
+   L <- t(chol(H + 1e-4 * diag(nrow(H))))
> L <- ltMatrices(L[lower.tri(L, diag = TRUE)], diag = TRUE)
> Nsim <- 50000
> Z <- matrix(rnorm(Nsim * nrow(H)), ncol = Nsim)
> rC <- solve(L, Z)[-1:J,] + op$par[-1:J] ### remove mean parameters
```

The standard error in this sample should be close to the ones obtained from the inverse Fisher information

```
> c(sqrt(rowMeans((rC - rowMeans(rC))^2)))
      5      6      7      8      9     10     11     12     13     14
0.05130 0.07990 0.12446 0.16090 0.07609 0.11567 0.14020 0.09622 0.10415 0.08279
```

```
> c(sqrt(diagonals(crossprod(solve(L)))))
 [1] 0.06826 0.10816 0.12670 0.14074 0.05498 0.10839 0.12442 0.14312 0.08813
[10] 0.11638 0.13340 0.09587 0.10451 0.08154
```

We now coerce the matrix rC to an object of class ltMatrices

```
> rC <- ltMatrices(rC, diag = TRUE)
```

The object rC contains all sampled Cholesky factors of the covariance matrix. From each of these matrices, we compute the regression coefficient, giving us a sample we can use to compute standard errors from

```
> rbeta <- cond_mvnorm(chol = rC, which_given = 2:J, given = diag(J - 1))$mean
> sqrt(rowMeans((rbeta - rowMeans(rbeta))^2))
 [1] 0.08793 0.04869 0.07752
```

which are, as expected, slightly different from the ones obtained from the more informative exact observations

```
> sqrt(diag(vcov(m1)))[-1L]
      Y2      Y3      Y4
0.08230 0.05039 0.06246
```

Chapter 5

Continuous-discrete Likelihoods

We sometimes are faced with outcomes measured at different levels of precision. Some variables might have been observed very exactly, and therefore we might want to use the log-Lebesgue density for defining the log-likelihood. Other variables might be available as relatively wide intervals only, and thus the log-likelihood is a log-probability. We can use the infrastructure developed so far to compute a joint likelihood. Let's assume we are interested in the joint distribution of $(\mathbf{Y}_i, \mathbf{X}_i)$ and we observed $\mathbf{Y}_i = \mathbf{y}_i$ (that is, exact observations of \mathbf{Y}) and $\mathbf{a}_i < \mathbf{X}_i \leq \mathbf{b}_i$ (that is, interval-censored observations for \mathbf{X}_i). We define the log-likelihood based on the joint normal distribution $(\mathbf{Y}_i, \mathbf{X}_i) \sim \mathbb{N}_J((\boldsymbol{\mu}_i, \boldsymbol{\eta}_i)^\top, \mathbf{C}_i \mathbf{C}_i^\top)$ as

$$\ell_i(\boldsymbol{\mu}_i, \boldsymbol{\eta}_i, \mathbf{C}_i) = \ell_i(\boldsymbol{\mu}_i, \mathbf{C}_{\mathbf{Y},i}) + \log(\mathbb{P}(\mathbf{a}_i < \mathbf{X}_i \leq \mathbf{b}_i \mid \mathbf{C}_i, \boldsymbol{\mu}_i, \boldsymbol{\eta}_i, \mathbf{Y}_i = \mathbf{y}_i)).$$

where $\mathbf{C}_{\mathbf{Y},i}$ is the upper part of \mathbf{C}_i corresponding to the marginal distribution of \mathbf{Y}_i . The conditional probability of \mathbf{X} given \mathbf{Y} depends on all parameters, as explained in Section 2.13. The trick here is to decompose the joint likelihood into a product of the marginal Lebesgue density of \mathbf{Y}_i and the conditional probability of \mathbf{X}_i given $\mathbf{Y}_i = \mathbf{y}_i$.

We first check the data

```
< dp input checks 100 > ≡
```

```
stopifnot(xor(missing(chol), missing(invchol)))
cJ <- nrow(obs)
dJ <- nrow(lower)
N <- ncol(obs)
stopifnot(N == ncol(lower))
stopifnot(N == ncol(upper))
if (all(mean == 0)) {
  cmean <- 0
  dmean <- 0
} else {
  if (!is.matrix(mean) || NCOL(mean) == 1L)
    mean <- matrix(mean, nrow = cJ + dJ, ncol = N)
  stopifnot(nrow(mean) == cJ + dJ)
  stopifnot(ncol(mean) == N)
  cmean <- mean[1:cJ,, drop = FALSE]
  dmean <- mean[-(1:cJ),, drop = FALSE]
}
◇
```

Fragment referenced in 101, 103.

We can use `marg_mvnorm` and `cond_mvnorm` to compute the marginal and the conditional normal distributions and the joint log-likelihood is simply the sum of the two corresponding log-likelihoods.

<ldpmvnorm 101> ≡

```
ldpmvnorm <- function(obs, lower, upper, mean = 0, chol, invchol,
  logLik = TRUE, ...) {

  if (missing(obs) || is.null(obs))
    return(lpmvnorm(lower = lower, upper = upper, mean = mean,
      chol = chol, invchol = invchol, logLik = logLik, ...))
  if (missing(lower) && missing(upper) || is.null(lower) && is.null(upper))
    return(ldmvnorm(obs = obs, mean = mean,
      chol = chol, invchol = invchol, logLik = logLik))

  <dp input checks 100>

  if (!missing(invchol)) {
    J <- dim(invchol)[2L]
    stopifnot(cJ + dJ == J)

    md <- marg_mvnorm(invchol = invchol, which = 1:cJ)
    ret <- ldmvnorm(obs = obs, mean = cmean, invchol = md$invchol,
      logLik = logLik)

    cd <- cond_mvnorm(invchol = invchol, which_given = 1:cJ,
      given = obs - cmean, center = TRUE)
    ret <- ret + lpmvnorm(lower = lower, upper = upper, mean = dmean,
      invchol = cd$invchol, center = cd$center,
      logLik = logLik, ...)

    return(ret)
  }

  J <- dim(chol)[2L]
  stopifnot(cJ + dJ == J)

  md <- marg_mvnorm(chol = chol, which = 1:cJ)
  ret <- ldmvnorm(obs = obs, mean = cmean, chol = md$chol, logLik = logLik)

  cd <- cond_mvnorm(chol = chol, which_given = 1:cJ,
    given = obs - cmean, center = TRUE)
  ret <- ret + lpmvnorm(lower = lower, upper = upper, mean = dmean,
    chol = cd$chol, center = cd$center,
    logLik = logLik, ...)

  return(ret)
}
◇
```

Fragment referenced in 66.

The score function requires a little extra work. We start with the case when `invchol` is given

< sldpmvnorm invchol 102 > ≡

```
byrow_orig <- attr(invchol, "byrow")
invchol <- ltMatrices(invchol, byrow = TRUE)

J <- dim(invchol)[2L]
stopifnot(cJ + dJ == J)

md <- marg_mvnorm(invchol = invchol, which = 1:cJ)
cs <- sldmvnorm(obs = obs, mean = cmean, invchol = md$invchol, logLik = logLik)

obs_cmean <- obs - cmean
cd <- cond_mvnorm(invchol = invchol, which_given = 1:cJ,
                 given = obs_cmean, center = TRUE)
ds <- slpmvnorm(lower = lower, upper = upper, mean = dmean,
               center = cd$center, invchol = cd$invchol,
               logLik = logLik, ...)

tmp0 <- solve(cd$invchol, ds$mean, transpose = TRUE)
tmp <- - tmp0[rep(1:dJ, each = cJ),,drop = FALSE] *
      obs_cmean[rep(1:cJ, dJ),,drop = FALSE]

Jp <- nrow(unclass(invchol))
diag <- attr(invchol, "diag")
M <- as.array(ltMatrices(1:Jp, diag = diag, byrow = TRUE))[, ,1]
ret <- matrix(0, nrow = Jp, ncol = ncol(obs))
M1 <- M[1:cJ, 1:cJ]
idx <- t(M1)[upper.tri(M1, diag = diag)]
ret[idx,] <- Lower_tri(cs$invchol, diag = diag)

idx <- c(t(M[-(1:cJ), 1:cJ]))
ret[idx,] <- tmp

M3 <- M[-(1:cJ), -(1:cJ)]
idx <- t(M3)[upper.tri(M3, diag = diag)]
ret[idx,] <- Lower_tri(ds$invchol, diag = diag)

ret <- ltMatrices(ret, diag = diag, byrow = TRUE)
if (!diag) diagonals(ret) <- 0
ret <- ltMatrices(ret, byrow = byrow_orig)

### post differentiate mean
aL <- as.array(invchol)[-(1:cJ), 1:cJ,,drop = FALSE]
lst <- tmp0[rep(1:dJ, cJ),,drop = FALSE]
if (dim(aL)[3] == 1)
  aL <- aL[, ,rep(1, ncol(lst)), drop = FALSE]
dim <- dim(aL)
dobs <- -margin.table(aL * array(lst, dim = dim), 2:3)

ret <- c(list(invchol = ret, obs = cs$obs + dobs),
        ds[c("lower", "upper")])
if (logLik) ret$logLik <- cs$logLik + ds$logLik
ret$mean <- rbind(-ret$obs, ds$mean)
return(ret)
◇
```

Fragment referenced in [103](#).

For `chol`, we compute the above code for its inverse and post-differentiate using the vec-trick

`< sldpmvnorm 103 > ≡`

```
sldpmvnorm <- function(obs, lower, upper, mean = 0, chol, invchol,
                      logLik = TRUE, ...) {

  if (missing(obs) || is.null(obs))
    return(sldpmvnorm(lower = lower, upper = upper, mean = mean,
                     chol = chol, invchol = invchol, logLik = logLik, ...))
  if (missing(lower) && missing(upper) || is.null(lower) && is.null(upper))
    return(sldpmvnorm(obs = obs, mean = mean,
                     chol = chol, invchol = invchol, logLik = logLik))

  < dp input checks 100 >

  if (!missing(invchol)) {
    < sldpmvnorm invchol 102 >
  }

  invchol <- solve(chol)
  ret <- sldpmvnorm(obs = obs, lower = lower, upper = upper,
                  mean = mean, invchol = invchol, logLik = logLik, ...)
  ### this means: ret$chol <- - vectrick(invchol, ret$invchol, invchol)
  ret$chol <- as.chol(- vectrick(invchol, ret$invchol))
  ret$invchol <- NULL
  return(ret)
}
◇
```

Fragment referenced in 66.

Let's assume we observed the first two dimensions exactly in our small example, and the remaining two dimensions are only known in intervals. The log-likelihood and score function for μ and C are

```
> ic <- 1:2          ### position of continuous variables
> ll_cd <- function(parm, J) {
+   m <- parm[1:J]      ### mean parameters
+   parm <- parm[-(1:J)]  ### chol parameters
+   C <- matrix(c(parm), ncol = 1L)
+   C <- ltMatrices(C, diag = TRUE, byrow = BYROW)
+   -ldpmvnorm(obs = Y[ic,], lower = lwr[-ic,],
+             upper = upr[-ic,], mean = m, chol = C,
+             w = W[-ic,,drop = FALSE], M = M)
+ }
> sc_cd <- function(parm, J) {
+   m <- parm[1:J]      ### mean parameters
+   parm <- parm[-(1:J)]  ### chol parameters
+   C <- matrix(c(parm), ncol = 1L)
+   C <- ltMatrices(C, diag = TRUE, byrow = BYROW)
+   ret <- sldpmvnorm(obs = Y[ic,], lower = lwr[-ic,],
+                   upper = upr[-ic,], mean = m, chol = C,
+                   w = W[-ic,,drop = FALSE], M = M)
+   return(-c(rowSums(ret$mean),
+               rowSums(Lower_tri(ret$chol, diag = TRUE))))
+ }
```

and the score function seems to be correct

```
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll_cd, start, J = J), sc_cd(start, J = J),
+       check.attributes = FALSE, tolerance = 1e-6)
```

We can now jointly estimate all model parameters via

```
> op <- optim(start, fn = ll_cd, gr = sc_cd, J = J,
+             method = "L-BFGS-B", lower = llim,
+             control = list(trace = FALSE))
> ## estimated C
> ltMatrices(matrix(op$par[-(1:J)], ncol = 1),
+             diag = TRUE, byrow = BYROW)
```

```
, , 1
      1      2      3      4
1 0.68303      .      .      .
2 0.26504 0.93467      .      .
3 0.53509 -0.05736 1.11261      .
4 0.06749 0.95887 0.07775 0.96692
```

```
> ## compare with true C
> lt
```

```
, , 1
      1      2      3      4
1 0.7071      .      .      .
2 0.2500 0.9682      .      .
3 0.6124 -0.1581 1.0488      .
4      . 1.0954 0.1651 0.8790
```

```
> ## estimated means
> op$par[1:J]
```

```
      1      2      3      4
0.9685 2.1269 2.9441 3.9898
```

```
> ## compare with true means
> mn
```

```
[1] 1 2 3 4
```

The one restriction in both `ldpmvnorm` and `sldpmvnorm` is that the continuous variables \mathbf{Y} are ranked before the discrete variables \mathbf{X} in the observation $(\mathbf{Y}_i, \mathbf{X}_i)$, and thus also in $(\boldsymbol{\mu}, \boldsymbol{\eta})$ and \mathbf{C} (the subscript i is dropped from the parameters in the following paragraph to keep the notational complexity in check).

While the means can be simply permuted, this is not the case for the Cholesky factor \mathbf{C} (see function `aperm` in Section 2.12). Of course, we can simply permute $\hat{\mathbf{C}}_i$, but we loose standard errors in this process. Alternatively, we can permute the order of variables in \mathbf{C} to our liking in the log-likelihood function (while keeping the original order of the observations and for the mean parameters)

```

> ### discrete variables first
> perm <- c((1:J)[-ic], ic)
> ll_ap <- function(parm, J) {
+   m <- parm[1:J]          ### mean parameters; NOT permuted
+   parm <- parm[-(1:J)]    ### chol parameters
+   C <- matrix(c(parm), ncol = 1L)
+   C <- ltMatrices(C, diag = TRUE, byrow = BYROW)
+   Ct <- aperm(as.chol(C), perm = perm)
+   -ldpvmnorm(obs = Y[ic,], lower = lwr[-ic,],
+             upper = upr[-ic,], mean = m, chol = Ct,
+             w = W[-ic,,drop = FALSE], M = M)
+ }

```

Unfortunately, this distorts the score function and we need to “de-permute” the scores. We start with $\Sigma = \mathbf{C}\mathbf{C}^\top$, the Cholesky decomposition of a quadratic positive definite $J \times J$ covariance matrix. There are $J \times (J + 1)/2$ parameters in the lower triangular part (including the diagonal) of \mathbf{C} . Changing the order of the variables by a permutation π with permutation matrix Π gives a covariance $\Pi\mathbf{C}\mathbf{C}^\top\Pi^\top$. This is no longer a Cholesky decomposition, because $\Pi\mathbf{C}$ is not lower triangular. The new decomposition is

$$\Pi\mathbf{C}\mathbf{C}^\top\Pi^\top = \tilde{\mathbf{C}}\tilde{\mathbf{C}}^\top$$

($\tilde{\mathbf{C}}$ is what `aperm` computes). As \mathbf{C} , the Cholesky factor $\tilde{\mathbf{C}}$ is lower triangular with $J \times (J + 1)/2$ parameters. We could write this operation as a function

$$f_3 : \mathbb{R}^{J \times (J+1)/2} \rightarrow \mathbb{R}^{J \times (J+1)/2}$$

$$f_3(\mathbf{C}) = \tilde{\mathbf{C}},$$

where in fact $f_3 = \text{aperm}$, and we are interested in its gradient. Deriving the gradient of a Cholesky decomposition might seem hopeless (it certainly did, at least to me, for a very long time), but there is a trick. Let us define two other functions:

$$f_1 : \mathbb{R}^{J \times (J+1)/2} \rightarrow \mathbb{R}^{J \times J}$$

$$f_1(\mathbf{C}) = \Pi\mathbf{C}\mathbf{C}^\top\Pi^\top$$

$$f_2 : \mathbb{R}^{J \times (J+1)/2} \rightarrow \mathbb{R}^{J \times J}$$

$$f_2(\tilde{\mathbf{C}}) = \tilde{\mathbf{C}}\tilde{\mathbf{C}}^\top.$$

Exploiting the chain rule for the composition $f_1 = f_2 \circ f_3$, we can write the gradient of f_1 as the product of the gradients of f_2 and f_3 :

$$\frac{\partial f_1(\mathbf{C})}{\partial \mathbf{C}} = \frac{\partial f_2(\tilde{\mathbf{C}})}{\partial \tilde{\mathbf{C}}} \frac{\partial f_3(\mathbf{C})}{\partial \mathbf{C}}. \quad (5.1)$$

The last factor is what we want to compute. It turns out that it is simpler to compute the first two gradients first and, in a second step, to derive the last factor. In more detail

$$\begin{aligned} \frac{\partial f_1(\mathbf{C})}{\partial \mathbf{C}} &= \frac{\partial \Pi\mathbf{C}\mathbf{C}^\top\Pi^\top}{\partial \mathbf{C}} \\ &= \frac{\partial \Pi\mathbf{C}\mathbf{C}^\top\Pi^\top}{\partial \Pi\mathbf{C}} \frac{\partial \Pi\mathbf{C}}{\mathbf{C}} \\ &= \left((\Pi\mathbf{C} \otimes \mathbf{I}_J) + (\mathbf{I}_J \otimes \Pi\mathbf{C}) \frac{\partial \mathbf{A}^\top}{\partial \mathbf{A}} \right) (\mathbf{I}_J \otimes \Pi). \end{aligned}$$

(\mathbf{A} is a quadratic matrix and the gradient of its transpose is a permutation matrix). This analytic

expression only contains known elements and can be computed. The same applies to

$$\begin{aligned}\frac{\partial f_2(\tilde{\mathbf{C}})}{\partial \tilde{\mathbf{C}}} &= \frac{\partial \tilde{\mathbf{C}} \tilde{\mathbf{C}}^\top \Pi}{\partial \tilde{\mathbf{C}}} \\ &= (\tilde{\mathbf{C}} \otimes \mathbf{I}_J) + (\mathbf{I}_J \otimes \tilde{\mathbf{C}}) \frac{\partial \mathbf{A}^\top}{\partial \mathbf{A}}\end{aligned}$$

Both expressions treat \mathbf{C} or $\tilde{\mathbf{C}}$ as full matrices, we are only interested in the score contributions by the $J \times (J + 1)/2$ lower triangular elements. Using sloppy notation, we collect the relevant columns in matrices $\mathbf{B}_1 = \frac{\partial f_1(\mathbf{C})}{\partial \mathbf{C}} \in \mathbb{R}^{J^2 \times J \times (J+1)/2}$ and $\mathbf{B}_2 = \frac{\partial f_2(\tilde{\mathbf{C}})}{\partial \tilde{\mathbf{C}}} \in \mathbb{R}^{J^2 \times J \times (J+1)/2}$. For the last, unknown, factor, we write $\mathbf{B}_3 = \frac{\partial f_3(\tilde{\mathbf{C}})}{\partial \tilde{\mathbf{C}}} \in \mathbb{R}^{J \times (J+1)/2 \times J \times (J+1)/2}$ and, with formula (5.1), $\mathbf{B}_1 = \mathbf{B}_2 \mathbf{B}_3$. We can then solve for \mathbf{B}_3 in the system $\mathbf{B}_1^\top \mathbf{B}_1 = \mathbf{B}_1^\top \mathbf{B}_2 \mathbf{B}_3$.

With `chol = C`, `permuted_chol = C_tilde`, `perm = pi` and score `score_schol` of the log-likelihood $\ell(\tilde{\mathbf{C}})$ with respect to the parameters in $\tilde{\mathbf{C}}$, we can now implement this de-permutation of the scores. Starting with some basic sanity checks, we require lower triangular matrix objects as inputs, with diagonal elements, and check if the dimensions match

< deperma input checks chol 106a > \equiv

```
stopifnot(is.ltMatrices(chol)) ### NOTE: replace with is.chol
byrow_orig <- attr(chol, "byrow")
chol <- ltMatrices(chol, byrow = FALSE)
stopifnot(is.ltMatrices(permuted_chol)) ### NOTE: replace with is.chol
permuted_chol <- ltMatrices(permuted_chol, byrow = FALSE)
stopifnot(max(abs(dim(chol) - dim(permuted_chol))) == 0)
J <- dim(chol)[2L]
stopifnot(attr(chol, "diag"))
INVCHOL <- !missing(invchol)
◇
```

Fragment referenced in 108.

Regarding `perm`, we check if this is an actual permutation

< deperma input checks perm 106b > \equiv

```
if (missing(perm)) return(score_schol)
stopifnot(isTRUE(all.equal(sort(perm), 1:J)))
if (max(abs(perm - 1:J)) == 0) return(score_schol)
◇
```

Fragment referenced in 108.

The scores with respect to $\tilde{\mathbf{C}}$ have been computed elsewhere, we just check the dimensions. In case we were given the scores with respect to \mathbf{L} , we first compute the scores with respect to \mathbf{C} (as we were lazy and only derived the results for \mathbf{C}). As in `standardize`, the argument `score_schol` gives the score with respect to \mathbf{C} and it is the user's responsibility to provide this quantity (even when `invchol` is given).

< deperma input checks schol 107a > ≡

```
if (is.ltMatrices(score_schol)) {
  byrow_orig_s <- attr(score_schol, "byrow")
  score_schol <- ltMatrices(score_schol, byrow = FALSE)
  ### don't do this here!
  ### if (INVCHOL) score_schol <- -vectrick(permuted_invchol, score_schol)
  score_schol <- unclass(score_schol) ### this preserves byrow
}
stopifnot(is.matrix(score_schol))
N <- ncol(score_schol)
stopifnot(J * (J + 1) / 2 == nrow(score_schol))
◇
```

Fragment referenced in [108](#).

We'll have to loop over $i = 1, \dots, N$ eventually and therefore coerce all objects to objects of class `array`, there is no need to worry about row or column storage order. We set-up indices matrices and the permutation matrix Π

< deperma indices 107b > ≡

```
idx <- matrix(1:J^2, nrow = J, ncol = J)      ### assuming byrow = TRUE
tidx <- c(t(idx))
ltT <- idx[lower.tri(idx, diag = TRUE)]
P <- matrix(0, nrow = J, ncol = J)
P[cbind(1:J, perm)] <- 1
ID <- diag(J)
IDP <- (ID %x% P)
◇
```

Fragment referenced in [108](#).

and are now ready for the main course. We are gentle and also allow `invchol = L` as input, and we clean-up by post-differentiation at the very end in this case.

`< deperma 108 > ≡`

```

deperma <- function(chol = solve(invchol),
                   permuted_chol = solve(permuted_invchol),
                   invchol, permuted_invchol, perm, score_schol) {

  < deperma input checks chol 106a >
  < deperma input checks perm 106b >
  < deperma input checks schol 107a >

  < deperma indices 107b >

  Nc <- dim(chol)[1L]
  mC <- as.array(chol)[perm,,drop = FALSE]
  Ct <- as.array(permuted_chol)
  ret <- lapply(1:Nc, function(i) {
    B1 <- (mC[,i] %x% ID) + (ID %x% mC[,i])[,tidx]
    # ~~~~~ <- d t(A) / d A
    B1 <- B1 %*% IDP
    B1 <- B1[,ltT] ### relevant columns of B1
    B2 <- (Ct[,i] %x% ID) + (ID %x% Ct[,i])[,tidx]
    B2 <- B2[,ltT] ### relevant columns of B2
    B3 <- try(solve(crossprod(B2), crossprod(B2, B1)))
    if (inherits(B3, "try-error"))
      stop("failure computing permutation score")
    if (Nc == 1L)
      return(crossprod(score_schol, B3))
    return(crossprod(score_schol[,i,drop = FALSE], B3))
  })
  ret <- do.call("rbind", ret)
  ret <- ltMatrices(t(ret), diag = TRUE, byrow = FALSE)
  if (INVCHOL)
    ret <- -vectrick(chol, ret)
  ret <- ltMatrices(ret, byrow = byrow_orig_s)
  return(ret)
}
◇

```

Fragment referenced in 66.

We can now use this function to estimate the Cholesky factor for (\mathbf{X}, \mathbf{Y}) when the data comes as (\mathbf{Y}, \mathbf{X}) (which is needed because continuous variables come first in our implementation of log-likelihood and score function).

```

> sc_ap <- function(parm, J) {
+   m <- parm[1:J]          ### mean parameters; NOT permuted
+   parm <- parm[-(1:J)]   ### chol parameters
+   C <- matrix(c(parm), ncol = 1L)
+   C <- ltMatrices(C, diag = TRUE, byrow = BYROW)
+   ### permutation
+   Ct <- aperm(as.chol(C), perm = perm)
+   ret <- sldpvmnorm(obs = Y[ic,], lower = lwr[-ic,],
+                   upper = upr[-ic,], mean = m, chol = Ct,
+                   w = W[-ic,,drop = FALSE], M = M)
+   ### undo permutation for chol
+   retC <- deperma(chol = C, permuted_chol = Ct,
+                  perm = perm, score_schol = ret$chol)
+

```

```
+   return(-c(rowSums(ret$mean),
+             rowSums(Lower_tri(retC, diag = TRUE))))
+ }
```

and the score function seems to be correct

```
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll_ap, start, J = J), sc_ap(start, J = J),
+     check.attributes = FALSE, tolerance = 1e-6)
```

We can now jointly estimate all model parameters via

```
> op <- optim(start, fn = ll_ap, gr = sc_ap, J = J,
+            method = "L-BFGS-B", lower = llim,
+            control = list(trace = FALSE))
> ## estimated C for (X, Y)
> ltMatrices(matrix(op$par[-(1:J)], ncol = 1),
+            diag = TRUE, byrow = BYROW)
```

```
, , 1
```

	1	2	3	4
1	1.23596	.	.	.
2	0.05465	1.36452	.	.
3	0.29576	0.02194	0.61530	.
4	0.07133	0.66705	0.23615	0.66185

```
> ## compare with true _permuted_ C for (X, Y)
> round(as.array(aperm(as.chol(lt), perm = perm)), 4)
```

```
, , 1
```

	3	4	1	2
3	1.2247	0.000	0.0000	0.0000
4	0.0000	1.414	0.0000	0.0000
1	0.3536	0.000	0.6124	0.0000
2	0.0000	0.750	0.2887	0.5951

Chapter 6

Unstructured Gaussian Copula Estimation

With $\mathbf{Z} \sim \mathbb{N}_J(0, \mathbf{I}_J)$ and $\mathbf{Y} = \tilde{\mathbf{C}}\mathbf{Z} \sim \mathbb{N}_J(0, \tilde{\mathbf{C}}\tilde{\mathbf{C}}^\top)$ we want to estimate the off-diagonal elements of the lower triangular unit-diagonal matrix \mathbf{C} . We have $\tilde{\mathbf{C}}(\mathbf{C}) := \text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2}\mathbf{C}$ such that $\boldsymbol{\Sigma} = \tilde{\mathbf{C}}\tilde{\mathbf{C}}^\top$ is a correlation matrix ($\text{diag}(\boldsymbol{\Sigma}) = \mathbf{I}_J$). Note that directly estimating $\tilde{\mathbf{C}}$ requires $J(J+1)/2$ parameters under constraints $\text{diag}(\boldsymbol{\Sigma}) = 1$ whereas only $J(J-1)/2$ parameters are necessary when estimating the lower triangular part of \mathbf{C} . The standardisation by $\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2}$ ensures that $\text{diag}(\boldsymbol{\Sigma}) \equiv 1$, that is, unconstrained optimisation can be applied.

`<standardize 110> ≡`

```
standardize <- function(chol, invchol) {
  stopifnot(xor(missing(chol), missing(invchol)))
  if (!missing(invchol)) {
    stopifnot(!attr(invchol, "diag"))
    return(invcholD(invchol))
  }
  stopifnot(!attr(chol, "diag"))
  return(Dchol(chol))
}
◇
```

Fragment referenced in 66.

```
> C <- ltMatrices(runif(10))
> chk(as.array(chol2cov(standardize(chol = C))),
+     as.array(chol2cor(standardize(chol = C))))
> L <- solve(C)
> chk(as.array(invchol2cov(standardize(invchol = L))),
+     as.array(invchol2cor(standardize(invchol = L))))
```

The log-likelihood function is $\ell_i(\mathbf{C}_i)$ (we omit i in the following) and we assume the score

$$\frac{\partial \ell(\mathbf{C})}{\partial \mathbf{C}}$$

is already available. We want to compute the score

$$\frac{\partial \ell(\tilde{\mathbf{C}})}{\partial \mathbf{C}}$$

which gives

$$\frac{\partial \ell(\tilde{\mathbf{C}})}{\partial \mathbf{C}} = \underbrace{\frac{\partial \ell(\tilde{\mathbf{C}})}{\partial \tilde{\mathbf{C}}}}_{=: \mathbf{T}} \times \frac{\partial \tilde{\mathbf{C}}(\mathbf{C})}{\partial \mathbf{C}}$$

We further have

$$\frac{\partial \tilde{\mathbf{C}}(\mathbf{C})}{\partial \mathbf{C}} = (\mathbf{C}^\top \otimes \mathbf{I}_J) \frac{\partial \text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2}}{\partial \mathbf{C}} + (\mathbf{I}_J \otimes \text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2})$$

and thus

$$\frac{\partial \ell(\tilde{\mathbf{C}})}{\partial \mathbf{C}} = \text{vec}(\mathbf{I}_J \mathbf{T} \mathbf{C}^\top)^\top \frac{\partial \text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2}}{\partial \mathbf{C}} + \text{vec}(\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2} \mathbf{T} \mathbf{I}_J)^\top$$

and with

$$\begin{aligned} \frac{\partial \text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2}}{\partial \mathbf{C}} &= \left. \frac{\partial \text{diag}(\mathbf{A})^{-1/2}}{\partial \mathbf{A}} \right|_{\mathbf{A}=\mathbf{C}\mathbf{C}^\top} \frac{\partial \mathbf{C}\mathbf{C}^\top}{\partial \mathbf{C}} \\ &= -\frac{1}{2} \text{diag}(\text{vec}(\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-3/2})) \left[(\mathbf{C} \otimes \mathbf{I}_J) \frac{\partial \mathbf{C}}{\partial \mathbf{C}} + (\mathbf{I}_J \otimes \mathbf{C}) \frac{\partial \mathbf{C}^\top}{\partial \mathbf{C}} \right] \end{aligned}$$

we can write

$$\begin{aligned} \text{vec}(\mathbf{I}_J \mathbf{T} \mathbf{C}^\top)^\top \left(-\frac{1}{2} \right) \text{diag}(\text{vec}(\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-3/2})) &= -\frac{1}{2} \times \text{vec}(\mathbf{I}_J \mathbf{T} \mathbf{C}^\top)^\top \times \text{vec}(\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-3/2})^\top \\ &=: \mathbf{b}^\top \end{aligned}$$

thus

$$\begin{aligned} \frac{\partial \ell(\tilde{\mathbf{C}})}{\partial \mathbf{C}} &= \mathbf{b}^\top \left[(\mathbf{C} \otimes \mathbf{I}_J) \frac{\partial \mathbf{C}}{\partial \mathbf{C}} + (\mathbf{I}_J \otimes \mathbf{C}) \frac{\partial \mathbf{C}^\top}{\partial \mathbf{C}} \right] + \text{vec}(\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2} \mathbf{T} \mathbf{I}_J)^\top \\ &= \text{vec}(\mathbf{I}_J \mathbf{B} \mathbf{C})^\top + \text{vec}(\mathbf{C}^\top \mathbf{B} \mathbf{I}_J)^\top \frac{\partial \mathbf{C}^\top}{\partial \mathbf{C}} + \text{vec}(\text{diag}(\mathbf{C}\mathbf{C}^\top)^{-1/2} \mathbf{T} \mathbf{I}_J)^\top \end{aligned}$$

when $\mathbf{b} = \text{vec}(\mathbf{B})$. These scores are implemented in `destandardize` with `chol = C` and `score_schol = T`. If the model was parameterised in $\mathbf{L} = \mathbf{C}^{-1}$, we have `invchol = L`, however, we would still need to compute \mathbf{T} (`score_schol`, the score with respect to \mathbf{C} , and it is the user's responsibility to provide this quantity).

< destandardize 112 > ≡

```
destandardize <- function(chol = solve(invchol), invchol, score_schol)
{
  stopifnot(is.ltMatrices(chol))      ### NOTE: replace with is.chol
  J <- dim(chol)[2L]
  stopifnot(!attr(chol, "diag"))
  byrow_orig <- attr(chol, "byrow")
  chol <- ltMatrices(chol, byrow = FALSE)

  ### TODO: check byrow in score_schol?

  if (is.ltMatrices(score_schol))
    score_schol <- matrix(as.array(score_schol),
                          nrow = dim(score_schol)[2L]^2)
  stopifnot(is.matrix(score_schol))
  N <- ncol(score_schol)
  stopifnot(J^2 == nrow(score_schol))

  CCT <- Tcrossprod(chol, diag_only = TRUE)
  DC <- Dchol(chol, D = Dinv <- 1 / sqrt(CCT))
  SDC <- solve(DC)

  IDX <- t(M <- matrix(1:J^2, nrow = J, ncol = J))
  i <- cumsum(c(1, rep(J + 1, J - 1)))
  ID <- diagonals(as.integer(J), byrow = FALSE)
  if (dim(ID)[1L] != dim(chol)[1L])
    ID <- ID[rep(1, dim(chol)[1L]),]

  B <- vectrick(ID, score_schol, chol)
  B[i,] <- B[i,] * (-.5) * c(CCT)^(-3/2)
  B[-i,] <- 0

  Dtmp <- Dchol(ID, D = Dinv)

  ret <- vectrick(ID, B, chol, transpose = c(TRUE, FALSE)) +
    vectrick(chol, B, ID)[IDX,] +
    vectrick(Dtmp, score_schol, ID)

  if (!missing(invchol)) {
    ### this means: ret <- - vectrick(chol, ret, chol)
    ret <- - vectrick(chol, ret)
  }
  ret <- ret[M[lower.tri(M)],,drop = FALSE]
  if (!is.null(dimnames(chol)[[1L]]))
    colnames(ret) <- dimnames(chol)[[1L]]
  ret <- ltMatrices(ret,
                    diag = FALSE, byrow = FALSE,
                    names = dimnames(chol)[[2L]])
  ret <- ltMatrices(ret, byrow = byrow_orig)
  diagonals(ret) <- 0
  return(ret)
}
◇
```

Fragment referenced in [66](#).

We can now set-up the log-likelihood and score functions for a Gaussian copula model. We

start with the classical approach of generating the marginal observations \mathbf{Y} from the ECDF with denominator $N + 1$ and subsequent use of the Lebesgue density as likelihood. Because no stats text on multivariate problems is complete without a reference to Edgar Anderson's iris data, let's set up a model for these four classical variables

```
> data("iris", package = "datasets")
> J <- 4
> Z <- t(qnorm(do.call("cbind", lapply(iris[1:J], rank, ties.method = "max")) /
+         (nrow(iris) + 1)))
> (CR <- cor(t(Z)))

      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length      1.00000    -0.09064      0.8690     0.7695
Sepal.Width       -0.09064     1.00000     -0.2446    -0.1908
Petal.Length      0.86896    -0.24456     1.0000     0.8577
Petal.Width       0.76951    -0.19082     0.8577     1.0000

> ll <- function(parm) {
+   C <- ltMatrices(parm)
+   Cs <- standardize(chol = C)
+   -ldmvnorm(obs = Z, chol = Cs)
+ }
> sc <- function(parm) {
+   C <- ltMatrices(parm)
+   Cs <- standardize(chol = C)
+   -rowSums(Lower_tri(destandardize(chol = C,
+     score_schol = sldmvnorm(obs = Z, chol = Cs)$chol)))
+ }
> start <- t(chol(CR))
> start <- start[lower.tri(start)]
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll, start), sc(start), check.attributes = FALSE)
> op <- optim(start, fn = ll, gr = sc, method = "BFGS",
+   control = list(trace = FALSE), hessian = TRUE)
> op$value

[1] 605.5

> S_ML <- chol2cov(standardize(chol = ltMatrices(op$par)))
```

This approach is of course a bit strange, because we estimate the marginal distributions by nonparametric maximum likelihood whereas the joint distribution is estimated by plain maximum likelihood. For the latter, we can define the likelihood by boxes given by intervals obtained from the marginals ECDFs and estimate the Copula parameters by maximisation of this nonparametric likelihood.

```
> lwr <- do.call("cbind", lapply(iris[1:J], rank, ties.method = "min")) - 1L
> upr <- do.call("cbind", lapply(iris[1:J], rank, ties.method = "max"))
> lwr <- t(qnorm(lwr / nrow(iris)))
> upr <- t(qnorm(upr / nrow(iris)))
> M <- 500
> if (require("qrng", quietly = TRUE)) {
+   ### quasi-Monte-Carlo
+   W <- t(ghalton(M, d = J - 1))
+ } else {
```

```

+   ### Monte-Carlo
+   W <- matrix(runif(M * (J - 1)), nrow = J - 1, byrow = TRUE)
+ }
> ll <- function(parm) {
+   C <- ltMatrices(parm)
+   Cs <- standardize(chol = C)
+   -lpmvnorm(lower = lwr, upper = upr, chol = Cs, M = M, w = W)
+ }
> sc <- function(parm) {
+   C <- ltMatrices(parm)
+   Cs <- standardize(chol = C)
+   -rowSums(Lower_tri(destandardize(chol = C,
+     score_schol = slpmvnorm(lower = lwr, upper = upr, chol = Cs,
+       M = M, w = W)$chol)))
+ }
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll, start), sc(start), check.attributes = FALSE)
> op2 <- optim(start, fn = ll, gr = sc, method = "BFGS",
+   control = list(trace = FALSE), hessian = TRUE)
> S_NPML <- chol2cov(standardize(chol = ltMatrices(op2$par)))

```

For $N = 150$, the difference is (as expected) marginal:

```

> S_ML
, , 1
      1      2      3      4
1  1.0000 -0.1100  0.8797  0.7906
2 -0.1100  1.0000 -0.2597 -0.1957
3  0.8797 -0.2597  1.0000  0.8740
4  0.7906 -0.1957  0.8740  1.0000

> S_NPML
, , 1
      1      2      3      4
1  1.00000 -0.09786  0.8735  0.7833
2 -0.09786  1.00000 -0.2726 -0.2482
3  0.87346 -0.27261  1.0000  0.8849
4  0.78328 -0.24823  0.8849  1.0000

```

with relatively close standard errors

```

> sd_ML <- ltMatrices(sqrt(diag(solve(op$hessian))))
> diagonals(sd_ML) <- 0
> sd_NPML <- try(ltMatrices(sqrt(diag(solve(op2$hessian))))
> if (!inherits(sd_NPML, "try-error")) {
+   diagonals(sd_NPML) <- 0
+   print(sd_ML)
+   print(sd_NPML)
+ }
, , 1

```

	1	2	3	4
1
2	0.08219	.	.	.
3	0.13663	0.08769	.	.
4	0.12316	0.10956	0.10266	.

, , 1

	1	2	3	4
1
2	0.07731	.	.	.
3	0.14000	0.08695	.	.
4	0.13691	0.11038	0.11610	.

However, we are still in the wrong, because we pretended we know the marginal distribution functions. A holistic approach would estimate the marginal empirical distributions and the copula parameters simultaneously.

Chapter 7

Joint Mean and Covariance Estimation

From a computational points of view, the parameterisation of the multivariate normal distribution in terms of the inverse Cholesky factor \mathbf{L} and the scaled mean $\boldsymbol{\nu} = \mathbf{L}\boldsymbol{\mu}$ instead of the mean $\boldsymbol{\mu}$ is attractive, because the log-density is then jointly concave in $\boldsymbol{\nu}$ and \mathbf{L} and thus a convex optimisation problem would emerge (Barratt and Boyd, 2023). This also carries over to the interval-censored likelihood because probabilities of log-concave densities are again log-concave (Prékopa, 1973).

The package implements the log-likelihood contributions as $\ell_i(\boldsymbol{\mu}_i, \mathbf{L}_i)$ and derives the scores with respect to $\boldsymbol{\mu}_i$ and \mathbf{L}_i . With $\boldsymbol{\mu}_i = \mathbf{L}_i^{-1}\boldsymbol{\nu}$ it is simple to rewrite the log-likelihood contribution as a concave function of both parameters as

$$\ell_i^{\text{concave}}(\boldsymbol{\nu}_i, \mathbf{L}_i) = \ell_i(\mathbf{L}_i^{-1}\boldsymbol{\nu}_i, \mathbf{L}_i).$$

The implementation of the corresponding convex negative log-likelihood is also easy:

```
> J <- 5
> N <- 100
> ### mean
> m <- rnorm(J)
> L <- ltMatrices(prm <- runif(J * (J + 1) / 2), diag = TRUE)
> Z <- matrix(rnorm(N * J), nrow = J)
> Y <- solve(L, Z) + m
> ### scaled mean
> d <- L %*% m
> nll <- function(parm) {
+   d <- parm[seq_len(J)]
+   L <- ltMatrices(parm[-seq_len(J)], diag = TRUE)
+   -ldmvnorm(obs = Y, mean = solve(L, d), invchol = L)
+ }
> start <- c(d, prm)
> nll(start)

[1] 1135

> ### identical
> -ldmvnorm(obs = Y, mean = m, invchol = L)

[1] 1135
```

However, computing the scores is a bit more work. Let's first consider the scores with respect to $\boldsymbol{\nu}_i$. We have

$$\begin{aligned}\frac{\partial \ell_i^{\text{concave}}(\boldsymbol{\nu}_i, \mathbf{L}_i)}{\partial \boldsymbol{\nu}_i} &= \frac{\partial \ell_i(\mathbf{L}_i^{-1} \boldsymbol{\nu}_i, \mathbf{L}_i)}{\partial \boldsymbol{\nu}_i} \\ &= \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \boldsymbol{\mu}} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \frac{\partial \mathbf{L}_i^{-1} \boldsymbol{\nu}_i}{\partial \boldsymbol{\nu}_i} \\ &= \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \boldsymbol{\mu}} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \mathbf{L}_i^{-1}.\end{aligned}$$

The first term is the score with respect to $\boldsymbol{\mu}_i$ and thus the score with respect to $\boldsymbol{\nu}_i$ is straightforward to compute. The score with respect to \mathbf{L}_i is more complex. With the multivariate chain rule we have

$$\begin{aligned}\frac{\partial \ell_i^{\text{concave}}(\boldsymbol{\nu}_i, \mathbf{L}_i)}{\partial \mathbf{L}_i} &= \frac{\partial \ell_i(\mathbf{L}_i^{-1} \boldsymbol{\nu}_i, \mathbf{L}_i)}{\partial \mathbf{L}_i} \\ &= \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \boldsymbol{\mu}} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \frac{\partial \mathbf{L}_i^{-1} \boldsymbol{\nu}_i}{\partial \mathbf{L}_i} + \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \mathbf{L}_i} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \\ &= \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \boldsymbol{\mu}} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \frac{\partial \mathbf{L}_i^{-1} \boldsymbol{\nu}_i}{\partial \mathbf{L}_i^{-1}} \frac{\partial \mathbf{L}_i^{-1}}{\partial \mathbf{L}_i} + \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \mathbf{L}_i} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \\ &= \underbrace{\left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \boldsymbol{\mu}} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i}}_{=:\boldsymbol{\mu}'_i} (\boldsymbol{\nu}_i^\top \otimes \mathbf{I}_J)(-1)(\mathbf{L}_i^{-\top} \otimes \mathbf{L}_i^{-1}) + \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \mathbf{L}_i} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i} \\ &= \text{vec}(\boldsymbol{\mu}'_i \boldsymbol{\nu}_i^\top)(-1)(\mathbf{L}_i^{-\top} \otimes \mathbf{L}_i^{-1}) + \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{L}_i)}{\partial \mathbf{L}_i} \right|_{\boldsymbol{\mu}=\mathbf{L}_i^{-1} \boldsymbol{\nu}_i}\end{aligned}$$

where we apply the vec-trick twice in the last line.

The negative gradient is now

```
> nsc <- function(parm) {
+   d <- parm[seq_len(J)]
+   L <- ltMatrices(parm[-seq_len(J)], diag = TRUE)
+   ret <- sldmvnorm(obs = Y, mean = solve(L, d), invchol = L)
+   C <- solve(L)
+
+   J <- dim(L)[2L]
+   M <- matrix(seq_len(J^2), nrow = J, byrow = FALSE)
+   idx <- M[lower.tri(M, diag = TRUE)]
+
+   X <- -ret$obs
+   Y <- matrix(d, nrow = nrow(X), ncol = ncol(X))
+   A <- X[rep(1:nrow(X), times = nrow(X)),,drop = FALSE] *
+     Y[rep(1:nrow(Y), each = nrow(X)),,drop = FALSE]
+
+   scL <- - vectrick(C, A)
+   scL <- scL[idx,,drop = FALSE]
+   scL <- rowSums(unclass(scL) + unclass(ret$invchol))
+   - c(rowSums(solve(L, -ret$obs, transpose = TRUE)),
+     scL)
+ }
> chk(unname(nsc(start)), grad(nll, start))
```

Similarly, when the model is parameterized by $\boldsymbol{\nu}_i$ and $\mathbf{C}_i = \mathbf{L}_i^{-1}$, we can compute the log-likelihood as

```
> C <- ltMatrices(prm <- runif(J * (J + 1) / 2), diag = TRUE)
> Z <- matrix(rnorm(N * J), nrow = J)
> Y <- C %*% Z + m
> ### scaled mean
> d <- solve(C, m)
> nll <- function(parm) {
+   d <- parm[seq_len(J)]
+   C <- ltMatrices(parm[-seq_len(J)], diag = TRUE)
+   -ldmvnorm(obs = Y, mean = C %*% d, chol = C)
+ }
> start <- c(d, prm)
> nll(start)

[1] 144.1

> ### identical
> -ldmvnorm(obs = Y, mean = m, chol = C)

[1] 144.1
```

By the same arguments as above, the score with respect to $\boldsymbol{\nu}_i$ is

$$\begin{aligned} \frac{\partial \ell_i^{\text{concave}}(\boldsymbol{\nu}_i, \mathbf{C}_i)}{\partial \boldsymbol{\nu}_i} &= \frac{\partial \ell_i(\mathbf{C}_i \boldsymbol{\nu}_i, \mathbf{C}_i)}{\partial \boldsymbol{\nu}_i} \\ &= \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{C}_i)}{\partial \boldsymbol{\mu}} \right|_{\boldsymbol{\mu} = \mathbf{C}_i \boldsymbol{\nu}_i} \mathbf{C}_i. \end{aligned}$$

and the score with respect to \mathbf{C}_i is

$$\begin{aligned} \frac{\partial \ell_i^{\text{concave}}(\boldsymbol{\nu}_i, \mathbf{C}_i)}{\partial \mathbf{C}_i} &= \frac{\partial \ell_i(\mathbf{C}_i \boldsymbol{\nu}_i, \mathbf{C}_i)}{\partial \mathbf{C}_i} \\ &= \boldsymbol{\mu}'_i \boldsymbol{\nu}_i^\top + \left. \frac{\partial \ell_i(\boldsymbol{\mu}, \mathbf{C}_i)}{\partial \mathbf{C}_i} \right|_{\boldsymbol{\mu} = \mathbf{C}_i \boldsymbol{\nu}_i} \end{aligned}$$

```
> nsc <- function(parm) {
+   d <- parm[seq_len(J)]
+   C <- ltMatrices(parm[-seq_len(J)], diag = TRUE)
+   ret <- sldmvnorm(obs = Y, mean = C %*% d, chol = C)
+
+   J <- dim(C)[2L]
+   M <- matrix(seq_len(J^2), nrow = J, byrow = FALSE)
+   idx <- M[lower.tri(M, diag = TRUE)]
+
+   X <- -ret$obs
+   Y <- matrix(d, nrow = nrow(X), ncol = ncol(X))
+   A <- X[rep(1:nrow(X), times = nrow(X)),,drop = FALSE] *
+     Y[rep(1:nrow(Y), each = nrow(X)),,drop = FALSE]
+
+   scC <- A[idx,,drop = FALSE]
+   scC <- rowSums(unclass(scC) + unclass(ret$chol))
+   - c(rowSums(crossprod(C, -ret$obs)),
+     scC)
+ }
> chk(unname(nsc(start)), grad(nll, start))
```

Chapter 8

User Interface

```
"interface.R" 119a≡
```

```
  < mnorm 121a >  
  < mnorm methods 121c >  
  < mnorm simulate 122 >  
  < mnorm margDist 123 >  
  < mnorm condDist 124 >  
  < mnorm logLik 127c >  
  < mnorm LLgrad 133 >  
  ◊
```

The tools provided in the previous chapters are rather low-level, so we will invest some time into setting-up a more high-level interface for representing normal models, either as $\mathbb{N}_J(\boldsymbol{\mu}, \mathbf{C}\mathbf{C}^\top)$, or $\mathbb{N}_J(\boldsymbol{\mu}, \mathbf{L}^{-1}\mathbf{L}^{-\top})$, or $\mathbb{N}_J(\mathbf{L}^{-1}\boldsymbol{\nu}, \mathbf{L}^{-1}\mathbf{L}^{-\top})$, or $\mathbb{N}_J(\mathbf{C}\boldsymbol{\nu}, \mathbf{C}\mathbf{C}^\top)$, for simulating from such models, and for evaluating the log-likelihood and corresponding score functions. The latter functionality shall also work when only incomplete (variables are missing) or censored (observations are only known as intervals) data is available.

We start with the conversion of a lower triangular matrix \mathbf{x} to an `ltMatrices` object

```
< as.ltMatrices 119b > ≡
```

```
as.ltMatrices.default <- function(x) {  
  stopifnot(is.numeric(x))  
  if (!is.matrix(x)) x <- matrix(x)  
  DIAG <- max(abs(diag(x) - 1)) > .Machine$double.eps  
  DIAG <- DIAG & (nrow(x) > 1)  
  lt <- x[lower.tri(x, diag = DIAG)]  
  up <- x[upper.tri(x, diag = FALSE)]  
  stopifnot(max(abs(up)) < .Machine$double.eps)  
  nm <- rownames(x)  
  if (!is.null(nm))  
    return(ltMatrices(lt, diag = DIAG, names = nm))  
  return(ltMatrices(lt, diag = DIAG))  
}  
◊
```

Fragment referenced in 2.

and proceed defining a constructor for object representing, potentially multiple, multivariate normal distributions. If the Cholesky factor \mathbf{C} (or multiple Cholesky factors $\mathbf{C}_1, \dots, \mathbf{C}_N$) are

given as `chol` argument, we label them as being such objects using `as.chol`. If only a matrix is given, we convert it (if possible) to a single Cholesky factor \mathbf{C} . The same is done when \mathbf{L} is given as `invchol` argument. Of course, only one of these arguments must be specified.

`<mvnorm chol invchol 120a> ≡`

```

if (missing(chol) && missing(invchol))
  chol <- as.chol(ltMatrices(1, diag = TRUE))
stopifnot(xor(missing(chol), missing(invchol)))

if (!missing(chol)) {
  if (!is.ltMatrices(chol))
    chol <- as.ltMatrices(chol)
  scale <- as.chol(chol)
}

if (!missing(invchol)) {
  if (!is.ltMatrices(invchol))
    invchol <- as.ltMatrices(invchol)
  scale <- as.invchol(invchol)
}
ret <- list(scale = scale)
◇

```

Fragment referenced in [121a](#).

The mean, or multiple means, is stored as a $J \times 1$ or $J \times N$ matrix, and we check if dimensions and, possibly, names are in line with what was specified as `chol` or `invchol`

`<mvnorm mean 120b> ≡`

```

if (!missing(mean)) {
  stopifnot(is.numeric(mean))
  stopifnot(NROW(mean) == dim(scale)[2L])
  if (!is.matrix(mean)) {
    mean <- matrix(mean, nrow = NROW(mean))
    rownames(mean) <- names(mean)
  }
  nm <- dimnames(scale)[[2L]]
  if (is.null(rownames(mean)))
    rownames(mean) <- nm
  if (!isTRUE(all.equal(rownames(mean), nm)))
    stop("rownames of mean do not match")
  nm <- dimnames(scale)[[1L]]
  if (!is.null(nm) && dim(scale)[[2L]] == ncol(mean)) {
    if (is.null(colnames(mean)))
      colnames(mean) <- nm
    if (!isTRUE(all.equal(colnames(mean), nm)))
      stop("colnames of mean do not match")
  }
  ret$mean <- mean
}
◇

```

Fragment referenced in [121a](#).

Finally, we put everything together and return an object of class `mvnorm`, featuring `mean` and `scale`. The class of the latter slot carries the information how this object is to be interpreted (as

Cholesky factor or inverse thereof). We also allow the specification of the scaled mean $\boldsymbol{\nu} = \mathbf{L}\boldsymbol{\mu}$ as argument `invcholmean`:

`<mvnorm 121a> ≡`

```
### allow more than one distribution
mvnorm <- function(mean, invcholmean, chol, invchol) {

  <mvnorm chol invchol 120a>
  if (!missing(invcholmean)) {
    stopifnot(missing(mean))
    ret$invcholmean <- invcholmean
    if (!missing(invchol)) mean <- solve(invchol, invcholmean)
    if (!missing(chol)) mean <- Mult(chol, invcholmean)
  }
  <mvnorm mean 120b>
  class(ret) <- "mvnorm"
  return(ret)
}
◇
```

Fragment referenced in [119a](#).

After updating the object, it might be necessary to update $\boldsymbol{\nu}$ as well:

`<mean2invcholmean 121b> ≡`

```
if (!is.null(ret$mean)) {
  if (is.chol(ret$scale)) {
    ret$invcholmean <- solve(ret$scale, ret$mean)
  } else {
    ret$invcholmean <- Mult(ret$scale, ret$mean)
  }
}
◇
```

Fragment referenced in [121c](#), [123](#), [124](#).

We add a `names` and `aperm` method. The latter returns a multivariate normal distribution with permuted order of the variables

`<mvnorm methods 121c> ≡`

```
names.mvnorm <- function(x)
  dimnames(x$scale)[[2L]]

aperm.mvnorm <- function(a, perm, ...) {

  ret <- list(scale = aperm(a$scale, perm = perm, ...))
  if (!is.null(a$mean)) {
    ret$mean <- a$mean[perm,,drop = FALSE]
    <mean2invcholmean 121b>
  }
  class(ret) <- "mvnorm"
  ret
}
◇
```

Fragment referenced in [119a](#).

We are now ready to draw samples from such an object. If multiple normal distributions are contained in `object`, we return one sample each, otherwise, `nsim` samples are returned. Because most tools in this package expect data as $J \times N$ matrices, we return the data in this format. If a classical `data.frame` is preferred, `as.data.frame = TRUE` we provide one

`<mvnorm simulate 122> ≡`

```
simulate.mvnorm <- function(object, nsim = dim(object$scale)[1L], seed = NULL,
                             standardize = FALSE, as.data.frame = FALSE, ...) {

  J <- dim(object$scale)[2L]
  N <- dim(object$scale)[1L]
  if (N > 1)
    stopifnot(nsim == N)
  if (standardize) {
    if (is.chol(object$scale)) {
      object$scale <- standardize(chol = object$scale)
    } else {
      object$scale <- standardize(invchol = object$scale)
    }
  }
  Z <- matrix(rnorm(nsim * J), nrow = J)
  if (is.chol(object$scale)) {
    Y <- Mult(object$scale, Z)
  } else {
    Y <- solve(object$scale, Z)
  }
  ret <- Y
  if (!is.null(object$mean))
    ret <- ret + c(object$mean)
  rownames(ret) <- dimnames(object$scale)[[2L]]
  if (!as.data.frame)
    return(ret)
  return(as.data.frame(t(ret)))
}
◇
```

Fragment referenced in [119a](#).

It is maybe time for a first example, and we return to the iris dataset, ignoring the iris' species for the time being. We set-up a model in terms of the sample maximum-likelihood estimates

```
> data("iris", package = "datasets")
> vars <- names(iris)[-5L]
> N <- nrow(iris)
> m <- colMeans(iris[,vars])
> V <- var(iris[,vars]) * (N - 1) / N
> iris_mvn <- mvnorm(mean = m, chol = t(chol(V)))
> iris_var <- simulate(iris_mvn, nsim = nrow(iris))
```

Marginal and conditional distributions might be of interest, the `margDist` and `condDist` methods are simple wrappers to `marg_mvnorm` and `cond_mvnorm`

< mvnorm margDist 123 > ≡

```
margDist <- function(object, which, ...)
  UseMethod("margDist")

margDist.mvnorm <- function(object, which, ...) {
  if (is.chol(object$scale)) {
    ret <- list(scale = as.chol(marg_mvnorm(chol = object$scale,
                                           which = which)$chol))
  } else {
    ret <- list(scale = as.invchol(marg_mvnorm(invchol = object$scale,
                                              which = which)$invchol))
  }
  if (!is.null(object$mean)) {
    ret$mean <- object$mean[which,,drop = FALSE]
    < mean2invcholmean 121b >
  }
  class(ret) <- "mvnorm"
  return(ret)
}
◇
```

Fragment referenced in [119a](#).

<mvnorm condDist 124> ≡

```
condDist <- function(object, which_given, given, ...)
  UseMethod("condDist")

condDist.mvnorm <- function(object, which_given = 1L, given, ...) {
  if (is.chol(object$scale)) {
    ret <- cond_mvnorm(chol = object$scale, which_given = which_given,
                      given = given, ...)
    ret$scale <- as.chol(ret$chol)
    ret$chol <- NULL
  } else {
    ret <- cond_mvnorm(invchol = object$scale, which_given = which_given,
                      given = given, ...)
    ret$scale <- as.chol(ret$invchol)
    ret$invchol <- NULL
  }
  if (!is.null(object$mean)) {
    if (is.character(which_given))
      which_given <- match(which_given, dimnames(object$scale)[[2L]])
    if (ncol(object$mean) > 1L && ncol(ret$mean) > 1)
      stop("dimensions do not match")
    if (ncol(object$mean) == 1L && ncol(ret$mean) > 1L) {
      ret$mean <- object$mean[-which_given,,drop = TRUE] + ret$mean
    } else {
      ret$mean <- object$mean[-which_given,,drop = FALSE] + c(ret$mean)
    }
    <mean2invcholmean 121b>
  }
  class(ret) <- "mvnorm"
  return(ret)
}
◇
```

Fragment referenced in [119a](#).

We could now compute the marginal distribution of two Petal variables or the bivariate regressions of the two Petal variables given the observed Sepal variables. Note that the last object contains $N = 150$ different distributions

```
> j <- 3:4
> margDist(iris_mvn, which = vars[j])

$scale
, , 1

      Petal.Length Petal.Width
Petal.Length  1.7594          .
Petal.Width   0.7315    0.2051

$mean
      [,1]
Petal.Length 3.758
Petal.Width  1.199
```

```

$invcholmean      [,1]
Petal.Length      2.136
Petal.Width       -1.770

attr("class")
[1] "mvnorm"

> gm <- t(iris[,vars[-(j)]])
> iris_cmvn <- condDist(iris_mvn, which_given = vars[j], given = gm)

```

We now work towards implementating the corresponding log-likelihood function. This is a trivial task as long as all variables for all observations have been observed exactly (that is, we can interpret the data as being continuous). Here, we also want to allow imprecise, that is, interval-censored, measurements. The one constraint in `ldpmvnorm` is that the continuous variables come first, followed by the censored ones. This of course might not be in line with the variable ordering we have in mind for our model. Our log-likelihood function shall be able to evaluate the log-likelihood for arbitrary permutations of the variables and, optionally, also based on marginal distributions in case observations are missing.

The following `logLik` method for objects of class `mvnorm` is essentially a wrapper for `ldpmvnorm`, handling permutations, marginalisation, and standardisation. We begin with some sanity checks

`< argchecks 126 > ≡`

```
args <- c(object, list(...))
### mean is always there
args$invcholmean <- NULL
nargs <- missing(obs) + missing(lower) + missing(upper)
stopifnot(nargs < 3L)

nmobs <- NULL
if (!missing(obs)) {
  if (!is.null(obs)) {
    stopifnot(is.matrix(obs))
    nmobs <- rownames(obs)
  }
}
nmlower <- nmupper <- nmlu <- NULL
if (!missing(lower)) {
  if (!is.null(lower)) {
    stopifnot(is.matrix(lower))
    nmlu <- nmlower <- rownames(lower)
  }
}
if (!missing(upper)) {
  if (!is.null(lower)) {
    stopifnot(is.matrix(upper))
    nmupper <- rownames(upper)
    if (!missing(lower)) {
      stopifnot(isTRUE(all.equal(nmlower, nmupper)))
    } else {
      nmlu <- nmupper
    }
  }
}

nm <- c(nmobs, nmlu)
no <- names(object)
stopifnot(nm %in% no)
perm <- NULL
if (!isTRUE(all.equal(nm, no)))
  perm <- c(nm, no[!no %in% nm])

if (!missing(obs)) args$obs <- obs
if (!missing(lower)) args$lower <- lower
if (!missing(upper)) args$upper <- upper
◇
```

Fragment referenced in [127c](#), [133](#).

and proceed with the workhorse when **C** was given

<logLik chol 127a> ≡

```
names(args)[names(args) == "scale"] <- "chol"

if (standardize)
  args$chol <- standardize(chol = args$chol)
if (!is.null(perm)) {
  args$chol <- aperm(as.chol(args$chol), perm = perm)
  if (length(nm) < length(no))
    args$chol <- marg_mvnorm(chol = args$chol, which = nm)$chol
  args$mean <- args$mean[nm,,drop = FALSE]
}
return(do.call("ldpmvnorm", args))
◇
```

Fragment referenced in [127c](#).

For inverse Cholesky factors **L**, the code is very similar, just the argument names change

<logLik invchol 127b> ≡

```
names(args)[names(args) == "scale"] <- "invchol"
if (standardize)
  args$invchol <- standardize(invchol = args$invchol)
if (!is.null(perm)) {
  args$invchol <- aperm(as.invchol(args$invchol), perm = perm)
  if (length(nm) < length(no))
    args$invchol <- marg_mvnorm(invchol = args$invchol,
                               which = nm)$invchol
  args$mean <- args$mean[nm,,drop = FALSE]
}
return(do.call("ldpmvnorm", args))
◇
```

Fragment referenced in [127c](#).

Putting everything together in a corresponding logLik method

<mvnorm logLik 127c> ≡

```
logLik.mvnorm <- function(object, obs, lower, upper, standardize = FALSE,
  ...) {
  <argchecks 126>
  if (is.chol(object$scale)) {
    <logLik chol 127a>
  }
  <logLik invchol 127b>
}
◇
```

Fragment referenced in [119a](#).

allows us to evaluate the log-likelihood of the conditional models for iris

```
> logLik(object = iris_cmvn, obs = t(iris[,vars[-j]]))
[1] -4782
```

This implementation of the log-likelihood silently handles the case when variables have been specified in a different order than hard-wired into the model

```
> logLik(object = iris_cmvn, obs = t(iris[,rev(vars[-j])]))
```

```
[1] -4782
```

The hardest task is the implementation of a score function which features the same options as the log-likelihood function and provides the gradients with respect not only to the parameters (μ or ν and \mathbf{C} or \mathbf{L}), but also with respect to the data objects `obs`, `lower`, and `upper`.

In essence, we have to repair the damage imposed by a series of transformations in `logLik.mvnorm`, that is, by standardisation, permutation, and marginalisation. We start with the case when \mathbf{C} was given. First, we repeat all the steps performed in `logLik`, but call the score function `sldpmvnorm` instead of the log-likelihood function `ldpmvnorm`

```
<LLgrad chol 128a> ≡
```

```
names(args)[names(args) == "scale"] <- "chol"
sc <- args$chol
if (standardize)
  args$chol <- sc <- standardize(chol = args$chol)
if (!is.null(perm)) {
  if (!attr(args$chol, "diag")) {
    diagonals(args$chol) <- 1
    sc <- args$chol
  }
  args$chol <- pc <- aperm(as.chol(args$chol), perm = perm)
  if (length(nm) < length(no))
    args$chol <- marg_mvnorm(chol = args$chol, which = nm)$chol
  args$mean <- args$mean[nm,,drop = FALSE]
}
ret <- do.call("sldpmvnorm", args)
<LLgrad mean 128b>
<LLgrad marginalisation 129a>
<LLgrad deperma 129b>
<LLgrad destandarized 129c>
<LLgrad diagonals 130a>
<LLgrad return 130b>
◇
```

Fragment referenced in [133](#).

The next task is to post-differentiate all scores such that the gradients with respect to the original arguments of `logLik` are obtained. We start with the gradient with respect to μ , in case it was not given

```
<LLgrad mean 128b> ≡
```

```
### sldmvnorm returns mean score as -obs
if (is.null(ret$mean)) ret$mean <- - ret$obs
◇
```

Fragment referenced in [128a](#).

In case we marginalised over some variables, we have to set the omitted parameters to zero

<LLgrad marginalisation 129a> ≡

```
om <- length(no) - length(nm)
if (om > 0) {
  am <- matrix(0, nrow = om, ncol = ncol(ret$mean))
  rownames(am) <- no[!no %in% nm]
  ret$mean <- rbind(ret$mean, am)
  Jo <- dim(object$scale)[[2L]]
  pJ <- dim(args$invchol)[[2L]]
  am <- matrix(0, nrow = Jo * (Jo + 1) / 2 - pJ * (pJ + 1) / 2,
              ncol = dim(ret$invchol)[1L])
  byrow_orig <- attr(ret$chol, "byrow")
  ret$chol <- ltMatrices(ret$chol, byrow = TRUE)
  ### rbind only works for byrow = TRUE
  ret$chol <- ltMatrices(rbind(unclass(ret$chol), am),
                        byrow = TRUE,
                        diag = TRUE,
                        names = perm)
  ret$chol <- ltMatrices(ret$chol, byrow = byrow_orig)
}
◇
```

Fragment referenced in [128a](#).

If the order of the variables was permuted, we compute the scores for the original ordering of the variables, as explained in [Chapter 5](#)

<LLgrad deperma 129b> ≡

```
if (!is.null(perm))
  ret$chol <- deperma(chol = sc, permuted_chol = pc,
                    perm = match(perm, no),
                    score_schol = ret$chol)
◇
```

Fragment referenced in [128a](#).

The effect of standardization can be removed as discussed in [Chapter 6](#)

<LLgrad destandardized 129c> ≡

```
if (standardize)
  ret$chol <- destandardize(chol = object$scale,
                          score_schol = ret$chol)
◇
```

Fragment referenced in [128a](#).

and it remains to remove fix diagonal elements

<ILgrad diagonals 130a> ≡

```
if (!attr(sc, "diag"))
  ret$chol <- ltMatrices(Lower_tri(ret$chol, diag = FALSE),
                        diag = FALSE,
                        byrow = attr(ret$chol, "byrow"),
                        names = dimnames(ret$chol)[[2L]])
  ◇
```

Fragment referenced in [128a](#).

and to return the results, with mean scores in the correct ordering. Eventually, we have to perform the post-processing steps explained in Chapter 7 when the scaled mean $\nu = \mathbf{L}\mu$ was specified

<ILgrad return 130b> ≡

```
ret$scale <- ret$chol
ret$chol <- NULL
ret$mean <- ret$mean[no,,drop = FALSE]
if (!is.null(object$invcholmean)) {
  J <- dim(sc)[2L]
  M <- matrix(seq_len(J^2), nrow = J, byrow = FALSE)
  idx <- M[lower.tri(M, diag = TRUE)]

  X <- ret$mean
  Y <- matrix(object$invcholmean, nrow = nrow(X), ncol = ncol(X))
  A <- X[rep(1:nrow(X), times = nrow(X)),,drop = FALSE] *
      Y[rep(1:nrow(Y), each = nrow(X)),,drop = FALSE]

  scC <- ltMatrices(A[idx,,drop = FALSE], diag = TRUE, byrow = FALSE)
  if (!attr(sc, "diag"))
    scC <- ltMatrices(Lower_tri(scC, diag = FALSE), diag = FALSE, byrow = FALSE)
  scC <- ltMatrices(scC, byrow = attr(sc, "byrow"))
  ret$scale <- ltMatrices(unclass(scC) + unclass(ret$scale),
                        diag = attr(sc, "diag"),
                        byrow = attr(sc, "byrow"),
                        names = dimnames(sc)[[2L]])
  ret$invcholmean <- Mult(sc, X, transpose = TRUE)
  ret$mean <- NULL
}
return(ret)
  ◇
```

Fragment referenced in [128a](#).

The steps are essentially the same when \mathbf{L} was given, but we have to post-differentiate $\mathbf{C} = \mathbf{L}^{-1}$ with respect to \mathbf{L} . We start with marginalisation

<Lgrad invchol marginalisation 131a> ≡

```
om <- length(no) - length(nm)
if (om > 0) {
  am <- matrix(0, nrow = om, ncol = ncol(ret$mean))
  rownames(am) <- no[!no %in% nm]
  ret$mean <- rbind(ret$mean, am)
  Jo <- dim(object$scale)[[2L]]
  pJ <- dim(args$invchol)[[2L]]
  am <- matrix(0, nrow = Jo * (Jo + 1) / 2 - pJ * (pJ + 1) / 2,
              ncol = dim(ret$invchol)[1L])
  byrow_orig <- attr(ret$invchol, "byrow")
  ret$invchol <- ltMatrices(ret$invchol, byrow = TRUE)
  ### rbind only works for byrow = TRUE
  ret$invchol <- ltMatrices(rbind(unclass(ret$invchol), am),
                          byrow = TRUE,
                          diag = TRUE,
                          names = perm)
  ret$invchol <- ltMatrices(ret$invchol, byrow = byrow_orig)
}
◇
```

Fragment referenced in 132.

turn to post-processing

<Lgrad invchol post 131b> ≡

```
if (!is.null(object$invcholmean)) {
  J <- dim(si)[2L]
  M <- matrix(seq_len(J^2), nrow = J, byrow = FALSE)
  idx <- M[lower.tri(M, diag = TRUE)]

  X <- ret$mean
  Y <- matrix(object$invcholmean, nrow = nrow(X), ncol = ncol(X))
  A <- X[rep(1:nrow(X), times = nrow(X)),,drop = FALSE] *
      Y[rep(1:nrow(Y), each = nrow(X)),,drop = FALSE]

  scL <- - vectrick(solve(si), A)
  scL <- ltMatrices(scL[idx,,drop = FALSE], diag = TRUE, byrow = FALSE)
  if (!attr(si, "diag"))
    scL <- ltMatrices(Lower_tri(scL, diag = FALSE), diag = FALSE, byrow = FALSE)
  scL <- ltMatrices(scL, byrow = attr(si, "byrow"))
  ret$scale <- ltMatrices(unclass(scL) + unclass(ret$scale),
                        diag = attr(si, "diag"),
                        byrow = attr(si, "byrow"),
                        names = dimnames(si)[[2L]])
  ret$invcholmean <- solve(si, X, transpose = TRUE)
  ret$mean <- FALSE
}
◇
```

Fragment referenced in 132.

and finally put everything together

<LLgrad invchol 132> ≡

```
names(args)[names(args) == "scale"] <- "invchol"
si <- args$invchol
if (standardize)
  args$invchol <- si <- standardize(invchol = args$invchol)
if (!is.null(perm)) {
  if (!attr(args$invchol, "diag")) {
    diagonals(args$invchol) <- 1
    si <- args$invchol
  }
  args$invchol <- pi <- aperm(as.invchol(args$invchol), perm = perm)
  if (length(nm) < length(no))
    args$invchol <- marg_mvnorm(invchol = args$invchol,
                               which = nm)$invchol
  args$mean <- args$mean[nm,,drop = FALSE]
}
ret <- do.call("sldpmvnorm", args)
### sldmvnorm returns mean score as -obs
if (is.null(ret$mean)) ret$mean <- - ret$obs
```

<LLgrad invchol marginalisation 131a>

```
if (!is.null(perm))
  ret$invchol <- deperma(invchol = si, permuted_invchol = pi,
                       perm = match(perm, no),
                       score_schol = -vectrick(pi, ret$invchol))
if (standardize)
  ret$invchol <- destandardize(invchol = object$scale,
                              score_schol = -vectrick(si, ret$invchol))
if (!attr(si, "diag"))
  ret$invchol <- ltMatrices(Lower_tri(ret$invchol, diag = FALSE),
                          diag = FALSE,
                          byrow = attr(ret$invchol, "byrow"),
                          names = dimnames(ret$invchol)[[2L]])

ret$scale <- ret$invchol
ret$invchol <- NULL
ret$mean <- ret$mean[no,,drop = FALSE]
<LLgrad invchol post 131b>
return(ret)
◇
```

Fragment referenced in [133](#).

We can now provide the log-likelihood gradients

`< mvnorm lLgrad 133 > ≡`

```
lLgrad <- function(object, ...)
  UseMethod("lLgrad")

lLgrad.mvnorm <- function(object, obs, lower, upper, standardize = FALSE,
  ...) {
  < argchecks 126 >
  if (is.chol(object$scale)) {
    < lLgrad chol 128a >
  }
  < lLgrad invchol 132 >
}
◇
```

Fragment referenced in 119a.

Let's use this infrastructure to set-up maximum-likelihood estimation procedures. We start implementing the log-likelihood and score functions for the iris dataset

```
> J <- length(vars)
> obs <- t(iris[, vars])
> ll <- function(parm) {
+   C <- ltMatrices(parm[-(1:J)], diag = TRUE, names = vars)
+   x <- mvnorm(mean = parm[1:J], chol = C)
+   -logLik(object = x, obs = obs)
+ }
> sc <- function(parm) {
+   C <- ltMatrices(parm[-(1:J)], diag = TRUE, names = vars)
+   x <- mvnorm(mean = parm[1:J], chol = C)
+   ret <- lLgrad(object = x, obs = obs)
+   -c(rowSums(ret$mean), rowSums(Lower_tri(ret$scale, diag = TRUE)))
+ }
```

and can now estimate the mean and Cholesky factor of the covariance matrix. Before we start, we check if the gradient, evaluated at the sample maximum-likelihood estimates, is actually zero.

```
> ### don't start at the solution
> start <- round(c(c(iris_mvn$mean),
+   Lower_tri(iris_mvn$scale, diag = TRUE)), 2)
> llim <- rep(-Inf, J + J * (J + 1) / 2)
> llim[J + c(diagonals(ltMatrices(seq_len(J * (J + 1) / 2), diag = TRUE)))] <- 1e-4
> op <- optim(start, fn = ll, gr = sc, method = "L-BFGS-B",
+   lower = llim,
+   control = list(trace = FALSE,
+   factr = 1e-6)) ### noLD machines
> Chat <- ltMatrices(op$par[-(1:J)], diag = TRUE, names = vars)
> ML <- mvnorm(mean = op$par[1:J], chol = Chat)
```

Quite unsurprisingly, the results are practically equivalent to the analytically available maximum-likelihood estimators in this case

```
> ### covariance
> chol2cov(ML$scale)
```

```
, , 1
```

```
      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length      0.68112    -0.04215      1.2658      0.5128
Sepal.Width       -0.04215     0.18871     -0.3275     -0.1208
Petal.Length      1.26582    -0.32746      3.0955      1.2870
Petal.Width       0.51283    -0.12083      1.2870      0.5771
```

```
> V
```

```
      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length      0.68112    -0.04215      1.2658      0.5128
Sepal.Width       -0.04215     0.18871     -0.3275     -0.1208
Petal.Length      1.26582    -0.32746      3.0955      1.2870
Petal.Width       0.51283    -0.12083      1.2870      0.5771
```

```
> ### mean
```

```
> ML$mean[, , drop = TRUE]
```

```
      Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.843         3.057         3.758         1.199
```

```
> m
```

```
      Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.843         3.057         3.758         1.199
```

We now check if we can obtain corresponding results when jointly optimising the scaled mean ν and the inverse Cholesky factor L :

```
> ll <- function(parm, logLik = TRUE) {
+   L <- ltMatrices(parm[-(1:J)], diag = TRUE, names = vars)
+   x <- mvnorm(invcholmean = parm[1:J], invchol = L)
+   if (!logLik) return(x)
+   -logLik(object = x, obs = obs)
+ }
> sc <- function(parm) {
+   x <- ll(parm, logLik = FALSE)
+   ret <- llgrad(object = x, obs = obs)
+   -c(rowSums(ret$invcholmean), rowSums(Lower_tri(ret$scale, diag = TRUE)))
+ }
> ### note: This is a convex problem now, so (here incorrect)
> ### starting values shouldn't matter
> opL <- optim(start, fn = ll, gr = sc, method = "L-BFGS-B",
+             lower = llim, control = list(trace = FALSE, factr = 1e-6))
> MLL <- ll(opL$par, logLik = FALSE)
```

where the comparison to the analytic estimates is

```
> ## IGNORE_RDIF_BEGIN
```

```
> ### log-likelihood
```

```
> op$value
```

```
[1] 379.9
```

```
> opL$value
```

```

[1] 385.4

> ### covariance
> invchol2cov(MLL$scale)

, , 1

      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length  0.64644   -0.04582    1.1395    0.4411
Sepal.Width   -0.04582    0.21906   -0.4221   -0.1640
Petal.Length  1.13949   -0.42213    2.9581    1.2070
Petal.Width   0.44105   -0.16399    1.2070    0.5330

> V

      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length  0.68112   -0.04215    1.2658    0.5128
Sepal.Width   -0.04215    0.18871   -0.3275   -0.1208
Petal.Length  1.26582   -0.32746    3.0955    1.2870
Petal.Width   0.51283   -0.12083    1.2870    0.5771

> ### mean
> MLL$mean[, ,drop = TRUE]

Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.848      3.054      3.786      1.215

> m

Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.843      3.057      3.758      1.199

```

Now, this was a lot of work to replace `mean` and `var` with something more fancy, and we would of course not go down this way in real life. But how about a more complex situation where one (or more) variables are only known up to intervals? Let's present the first variable is such a case

```

> v1 <- vars[1]
> q1 <- quantile(iris[[v1]], probs = 1:4 / 5)
> head(f1 <- cut(iris[[v1]], breaks = c(-Inf, q1, Inf)))

[1] (5,5.6] (-Inf,5] (-Inf,5] (-Inf,5] (-Inf,5] (5,5.6]
Levels: (-Inf,5] (5,5.6] (5.6,6.1] (6.1,6.52] (6.52, Inf]

> lower <- matrix(c(-Inf, q1)[f1], nrow = 1)
> upper <- matrix(c(q1, Inf)[f1], nrow = 1)
> rownames(lower) <- rownames(upper) <- v1
> obs <- obs[!rownames(obs) %in% v1, ,drop = FALSE]

```

The only necessary modification to our code is the specification of `lower` and `upper` bounds for these intervals, and the removal of the first variable from the “exact continuous” observations `obs`. The rest of the machinery *doesn't need any update at all*. Note that the mean and covariance parameters are no longer orthogonal (as in the toy example above), so we need to optimise over both sets of parameters simultaneously and therefore opt for the parameterisation in terms of ν and \mathbf{L} because this leads to a convex optimisation problem also for interval-censored observations as mentioned in Chapter 7.

One additional tweak is necessary for dealing with extreme parameter values assigning a likelihood of numerically zero to some observations. In this case, the score function is not defined (because we divide the derivative by the likelihood). The corresponding values should be NA and removed from the summation.

```

> ll <- function(parm, logLik = TRUE) {
+   L <- ltMatrices(parm[-(1:J)], diag = TRUE, names = vars)
+   x <- mvnorm(invcholmean = parm[1:J], invchol = L)
+   if (!logLik) return(x)
+   -logLik(object = x, obs = obs, lower = lower, upper = upper,
+           tol = 1e-6) ### probs < tol are considered 0
+ }
> sc <- function(parm) {
+   x <- ll(parm, logLik = FALSE)
+   ret <- llgrad(object = x, obs = obs, lower = lower, upper = upper,
+                 tol = 1e-6) ### probs < tol are considered 0
+   -c(rowSums(ret$invcholmean, na.rm = TRUE),
+       rowSums(Lower_tri(ret$scale, diag = TRUE), na.rm = TRUE))
+ }
> start <- round(opL$par, 2)
> if (require("numDeriv", quietly = TRUE))
+   chk(grad(ll, start), sc(start), check.attributes = FALSE)
> opi <- optim(start, fn = ll, gr = sc, method = "L-BFGS-B",
+             lower = llim, control = list(trace = FALSE, factr = 1e-6))
> MLi <- ll(opi$par, logLik = FALSE)

```

Because the likelihood is a product of a continuous density and a conditional probability as introduced in Chapter 5, the two in-sample log-likelihoods are not comparable. However, the parameters of the two estimated normal distributions can be compared directly (and are rather close in our case)

```

> ### covariance
> invchol2cov(MLi$scale)
, , 1
      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length  0.71817   -0.02607      1.261      0.5167
Sepal.Width   -0.02607    0.18862     -0.327     -0.1211
Petal.Length  1.26064   -0.32698      3.079      1.2794
Petal.Width   0.51668   -0.12107      1.279      0.5739
> invchol2cov(MLL$scale)
, , 1
      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length  0.64644   -0.04582      1.1395     0.4411
Sepal.Width   -0.04582    0.21906     -0.4221    -0.1640
Petal.Length  1.13949   -0.42213      2.9581     1.2070
Petal.Width   0.44105   -0.16399      1.2070     0.5330
> chol2cov(ML$scale)
, , 1
      Sepal.Length Sepal.Width Petal.Length Petal.Width
Sepal.Length  0.68112   -0.04215      1.2658     0.5128
Sepal.Width   -0.04215    0.18871     -0.3275    -0.1208
Petal.Length  1.26582   -0.32746      3.0955     1.2870
Petal.Width   0.51283   -0.12083      1.2870     0.5771

```

```

> ### mean
> MLI$mean[, ,drop = TRUE]

Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.760      3.058      3.759      1.200

> MLL$mean[, ,drop = TRUE]

Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.848      3.054      3.786      1.215

> ML$mean[, ,drop = TRUE]

Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.843      3.057      3.758      1.199

```

Again, we can extract the regression coefficients from these multivariate normal distributions. We start with the μ and C parameterisation:

```

> cdstr <- condDist(ML, which_given = vars[1:3], given = diag(3))
> ### least-squares coefficients
> coef(irislm <- lm(Petal.Width ~ Sepal.Length + Sepal.Width + Petal.Length,
+                   data = iris))

(Intercept) Sepal.Length Sepal.Width Petal.Length
      -0.2403      -0.2073      0.2228      0.5241

> cdstr$mean - ML$mean["Petal.Width",]

           [,1] [,2] [,3]
Petal.Width -0.2073 0.2228 0.5241

> ### residual variance
> summary(irislm)$sigma^2

[1] 0.03685

> c(cdstr$scale^2) ### note: "chol" defines the distribution

[1] 0.03587

```

The regression coefficients and the residual variance are almost identical with those obtained from `lm()`. The same exercise, now with the ν and L parameterisation for exact and interval-censored observations, gives slightly different results for the latter, due to censoring:

```

> ### nu, L for exact observations
> cdstr <- condDist(MLL, which_given = vars[1:3], given = diag(3))
> ### least-squares coefficients
> cdstr$mean - ML$mean["Petal.Width",]

           [,1] [,2] [,3]
Petal.Width -0.231 0.2485 0.5516

> ### residual variance
> c(1 / cdstr$scale^2) ### note: "invchol" defines the distribution

[1] 0.0327

```

```

> ### nu, L for censored observations
> cdstr <- condDist(MLi, which_given = vars[1:3], given = diag(3))
> ### least-squares coefficients
> cdstr$mean - ML$mean["Petal.Width",]

           [,1] [,2] [,3]
Petal.Width -0.1381 0.1942 0.4937

> ### residual variance
> c(1 / cdstr$scale^2)

[1] 0.03844

> ## IGNORE_RDIF_END

```

The log-likelihood and score function automatically marginalise over dimensions where all observations are $(-\infty, \infty)$. We can simply omit these dimensions from the matrices specified as `obs`, `lower`, and `upper` arguments. Let's say we have four dimensions called A to D and three observations. All observations have $A = (-1, 1)$ and $B = (-\infty, \infty)$, so in fact, the likelihood is given by the marginal distribution of A, C, D .

```

> N <- 3
> J <- 4
> L <- ltMatrices(runif(J * (J + 1) / 2), diag = TRUE, names = LETTERS[1:J])
> Z <- matrix(rnorm(J * N), nrow = J)
> Y <- solve(L, Z)
> lwrA <- matrix(-1, nrow = 1, ncol = N)
> uprA <- matrix(1, nrow = 1, ncol = N)
> rownames(lwrA) <- rownames(uprA) <- "A"
> lwrB <- matrix(-Inf, nrow = 1, ncol = N)
> uprB <- matrix(Inf, nrow = 1, ncol = N)
> rownames(lwrB) <- rownames(uprB) <- "B"
> lwr <- rbind(lwrA, lwrB)
> upr <- rbind(uprA, uprB)
> obs <- Y[rev(LETTERS[3:J]),]    ### change order of dimensions

```

With this data, we first compute the log-likelihood and score functions for the complete data, that is, including the infinite intervals for B .

```

> w <- matrix(runif(1000), nrow = 1, byrow = TRUE)
> lABCD <- logLik(mvnorm(invchol = L), obs = obs, lower = lwr, upper = upr, w = w)
> sABCD <- llgrad(mvnorm(invchol = L), obs = obs, lower = lwr, upper = upr, w = w)

```

This is (almost) the same as omitting dimension B from the data, but of course not from the model

```

> lACD <- logLik(mvnorm(invchol = L), obs = obs, lower = lwrA, upper = uprA)
> sACD <- llgrad(mvnorm(invchol = L), obs = obs, lower = lwrA, upper = uprA)

```

We can compare the results

```

> chk(lABCD, lACD)
> nm <- names(sABCD)
> nm <- nm[!nm %in% c("lower", "upper")]
> chk(sABCD[nm], sACD[nm])

```

noting that the scores with respect to the B data in `lower` and `upper` are missing from `sACD`

```

> chk(sABCD$lower["A",,drop = FALSE], sACD$lower)
> chk(sABCD$upper["A",,drop = FALSE], sACD$upper)
> sABCD$lower["B",]      ### zero

[1] 0 0 0

> sABCD$upper["B",]     ### zero

[1] 0 0 0

```

Omitting dimensions might be important because `lpmvnorm` introduced in Chapter 3 does not check if both `lower` and `upper` are infinite and omission thus reduces the dimensionality of the integral we evaluate numerically.

We close this chapter with a word of warning: If more than one variable is censored, the `M` and `w` arguments to `lpmvnorm` and `slpmvnorm` have to be specified in `logLik` and `lLgrad` as additional arguments (...) *AND MUST BE IDENTICAL* in both calls.

Chapter 9

Reduced Rank Covariance Matrices

We sometimes can write the $J \times J$ covariance matrix as $\Sigma = \mathbf{B}\mathbf{B}^\top + \mathbf{D}$ where \mathbf{B} is $J \times K$ with $K < J$ and \mathbf{D} is a J -dimensional diagonal matrix. Marsaglia (1963) and Genz and Bretz (2009), in their Chapter 2.3.1, demonstrated that the probability $\mathbb{P}(\mathbf{a} < \mathbf{Y} \leq \mathbf{b} \mid \Sigma)$ can be written as

$$\mathbb{P}(\mathbf{a} < \mathbf{Y} \leq \mathbf{b} \mid \Sigma) = \int_{[0,1]^K} \prod_{j=1}^J \left(\Phi \left(\frac{b_j - \mathbf{B}_{jk} \Phi^{-1}(w_k)}{\sqrt{\mathbf{D}_{jj}}} \right) - \Phi \left(\frac{a_j - \mathbf{B}_{jk} \Phi^{-1}(w_k)}{\sqrt{\mathbf{D}_{jj}}} \right) \right) d\mathbf{w}$$

where the integration takes place with respect to $\mathbf{w} = (w_1, \dots, w_K)^\top$ from the K - (and not J -) dimensional unit hypercube.

We start implementing low-level functionality for computing log-probabilities for such structures with some book keeping

< RR input B D 140a > ≡

```
stopifnot(!missing(B))
if (!is.matrix(B)) B <- matrix(B, ncol = 1)
J <- nrow(B)
K <- ncol(B)
Dsqrtd <- sqrt(D)
◇
```

Fragment referenced in 142, 143.

We use $Z = \Phi^{-1}(w)$ and optional weights and compute the products $\mathbf{B}_{jk} \Phi^{-1}(w_k)$ for all j and k with standardisation by the diagonal elements of \mathbf{D} .

< RR input Z, weights 140b > ≡

```
stopifnot(nrow(Z) == K)
stopifnot(length(weights) == 1 || length(weights) == ncol(Z))
BZ <- (B / Dsqrtd) %*% Z
◇
```

Fragment referenced in 142, 143.

The limits \mathbf{a}

< RR input lower 141a > ≡

```
if (missing(lower)) {
  pl <- 0
} else {
  stopifnot(length(lower) == J)
  lower <- c(lower)
  lower <- (lower - mean) / Dsqrt
  pl <- pnorm(lBZ <- lower - BZ)
}
◇
```

Fragment referenced in [142](#), [143](#).

and **b**

< RR input upper 141b > ≡

```
if (missing(upper)) {
  pl <- 0
} else {
  stopifnot(length(upper) == J)
  upper <- c(upper)
  upper <- (upper - mean) / Dsqrt
  pu <- pnorm(uBZ <- upper - BZ)
}
◇
```

Fragment referenced in [142](#), [143](#).

are processed and we finally compute the integrant, making sure to avoid negative values. We first compute log-probabilities, compute the sums and exponentiate before summing up. In contrast to `lpmvnorm`, we also allow weights for the summation, such that sparse grids (for example from add-on package **SparseGrid**) can be utilised.

< RR inner 141c > ≡

```
inner <- pu - pl
inner <- pmax(0, inner)
retw <- weights * exp(.colSums(m = J, n = ncol(Z),
                             x = log(inner), na.rm = TRUE))
◇
```

Fragment referenced in [142](#), [143](#).

Finally, we wrap everything up in a function called `lpRR`

$\langle \text{lpRR 142} \rangle \equiv$

```
lpRR <- function(lower, upper, mean = 0, B, D = rep(1, nrow(B)),
                 Z, weights = 1 / ncol(Z), log.p = TRUE) {

   $\langle$  RR input B D 140a  $\rangle$ 
   $\langle$  RR input Z, weights 140b  $\rangle$ 
   $\langle$  RR input lower 141a  $\rangle$ 
   $\langle$  RR input upper 141b  $\rangle$ 

   $\langle$  RR inner 141c  $\rangle$ 
  ret <- sum(retw)
  if (log.p) return(log(max(c(0, ret))))
  return(ret)
}
 $\diamond$ 
```

Fragment referenced in 66.

We test this functionality for dimensions $J = 6$ and $K = 3$. That is, the integration problem reduces from a $J - 1 = 5$ dimensional one in `lpmvnorm` to a three dimensional one in `lpRR`. We first compare the two log-probabilities, computed with high accuracy

```
> J <- 6
> K <- 3
> B <- matrix(rnorm(J * K), nrow = J)
> D <- runif(J)
> S <- tcrossprod(B) + diag(D)
> Linv <- t(chol(S))
> Linv <- ltMatrices(Linv[lower.tri(Linv, diag = TRUE)], diag = TRUE)
> a <- -(2 + runif(J))
> b <- 2 + runif(J)
> M <- 1e6
> dim(w <- matrix(runif((J - 1) * M), nrow = J - 1, byrow = TRUE))

[1]      5 1000000

> lpmvnorm(lower = a, upper = b, chol = Linv, w = w)

[1] -1.348

> dim(Z <- matrix(rnorm(K * M), nrow = K))

[1]      3 1000000

> lpRR(lower = a, upper = b, B = B, D = D, Z = Z)

[1] -1.35
```

The score function with respect to \mathbf{a} , \mathbf{b} , \mathbf{B} , and \mathbf{D} is

$\langle \text{slpRR } 143 \rangle \equiv$

```
slpRR <- function(lower, upper, mean = 0, B, D = rep(1, nrow(B)),
                 Z, weights = 1 / ncol(Z), log.p = TRUE) {
  < RR input B D 140a >
  < RR input Z, weights 140b >
  < RR input lower 141a >
  < RR input upper 141b >

  < RR inner 141c >

  dlBZ <- dnorm(lBZ)
  duBZ <- dnorm(uBZ)

  d <- matrix(retw, nrow = nrow(B), ncol = ncol(Z), byrow = TRUE) / inner
  db <- d * (duBZ - dlBZ)
  tdb <- t(db)
  dB <- -1 * do.call("cbind", lapply(1:nrow(Z),
                                     function(r) colSums(tdb * Z[r,], na.rm = TRUE))) / Dsqrt
  Du <- -.5 / D * uBZ
  Dl <- -.5 / D * lBZ
  dD <- rowSums(d * duBZ * Du, na.rm = TRUE) -
        rowSums(d * dlBZ * Dl, na.rm = TRUE)
  dl <- -rowSums(d * dlBZ, na.rm = TRUE) / Dsqrt
  du <- rowSums(d * duBZ, na.rm = TRUE) / Dsqrt
  dm <- -du - dl # Dinb %*% -rowSums(d * (duBZ - dlBZ))
  fct <- 1
  if (log.p) fct <- 1 / sum(retw)
  list(lower = fct * dl, upper = fct * du, mean = fct * dm,
       B = fct * dB, D = fct * dD)
}
◇
```

Fragment referenced in 66.

We can now compare the gradients for **a**, **b** and the mean in our small example. We cannot expect them to be equal but close

```
> smv <- slpmvnorm(lower = a, upper = b, chol = Linv, w = w)
> sRR <- slpRR(lower = a, upper = b, B = B, D = D, Z = Z)
> chk(c(smv$lower), sRR$lower, tolerance = 1e-2)
> chk(c(smv$upper), sRR$upper, tolerance = 1e-2)
> chk(c(smv$mean), sRR$mean, tolerance = 1e-2 * 2)
```

The gradient with respect to **B** and **D** are finally checked against their numerical approximations

```
> Z <- matrix(rnorm(K * 1000), nrow = K)
> lB <- function(B) lpRR(lower = a, upper = b, B = B, D = D, Z = Z)
> gB <- grad(lB, B)
> sRR <- slpRR(lower = a, upper = b, B = B, D = D, Z = Z)
> chk(gB, c(sRR$B), tolerance = 1e-3)
> lD <- function(D) lpRR(lower = a, upper = b, B = B, D = D, Z = Z)
> gD <- grad(lD, D)
> chk(gD, c(sRR$D), tolerance = 1e-3)
> ### while we are at it, check lower and again
> llwr <- function(a) lpRR(lower = a, upper = b, B = B, D = D, Z = Z)
```

```
> glwr <- grad(llwr, a)
> chk(glwr, c(sRR$lower))
> lupr <- function(b) lpRR(lower = a, upper = b, B = B, D = D, Z = Z)
> gupr <- grad(lupr, b)
> chk(gupr, c(sRR$upper))
```

Chapter 10

Package Infrastructure

< R Header 145 > ≡

```
### Copyright (C) 2022- Torsten Hothorn
###
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###
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###
### You should have received a copy of the GNU General Public License
### along with 'mvtnorm'. If not, see <http://www.gnu.org/licenses/>.
###
### DO NOT EDIT THIS FILE
###
### Edit 'lmvnorm_src.w' and run 'nuweb -r lmvnorm_src.w'
```

◇

Fragment referenced in [2](#), [66](#).

< C Header 146 > ≡

```
/*
  Copyright (C) 2022- Torsten Hothorn

  This file is part of the 'mvtnorm' R add-on package.

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  along with 'mvtnorm'. If not, see <http://www.gnu.org/licenses/>.

  DO NOT EDIT THIS FILE

  Edit 'lmvnorm_src.w' and run 'nuweb -r lmvnorm_src.w'
*/
◇
```

Fragment referenced in [3](#), [67](#).

Appendix

This document uses the following matrix derivatives for vectors $\mathbf{y} \in \mathbb{R}^J$ and matrices $\mathbf{A} \in \mathbb{R}^{J \times J}$, $\mathbf{B} \in \mathbb{R}^{K \times J}$, and $\mathbf{X} \in \mathbb{R}^{J \times K}$:

$$\begin{aligned}
 \frac{\partial \mathbf{A} \mathbf{y}}{\partial \mathbf{y}} &= \mathbf{A} \\
 \frac{\partial \mathbf{A} \mathbf{y}}{\partial \mathbf{A}} &= \mathbf{y} \otimes \mathbf{I}_J \\
 \frac{\partial \mathbf{y}^\top \mathbf{A}^\top \mathbf{A} \mathbf{y}}{\partial \mathbf{A}} &= 2 \mathbf{A} \mathbf{y} \mathbf{y}^\top \\
 \frac{\partial \mathbf{A}^{-1}}{\partial \mathbf{A}} &= -(\mathbf{A}^{-\top} \otimes \mathbf{A}^{-1}) \\
 \frac{\partial \mathbf{A} \mathbf{A}^\top}{\partial \mathbf{A}} &= (\mathbf{A} \otimes \mathbf{I}_J) \frac{\partial \mathbf{A}}{\partial \mathbf{A}} + (\mathbf{I}_J \otimes \mathbf{A}) \frac{\partial \mathbf{A}^\top}{\partial \mathbf{A}} \\
 &= (\mathbf{A} \otimes \mathbf{I}_J) + (\mathbf{I}_J \otimes \mathbf{A}) \frac{\partial \mathbf{A}^\top}{\partial \mathbf{A}} \\
 \frac{\partial \text{diag}(\mathbf{A})}{\partial \mathbf{A}} &= \text{diag}(\text{vec}(\mathbf{I}_J)) \\
 \frac{\partial \mathbf{A}}{\partial \mathbf{A}} &= \text{diag}(\mathbf{I}_{J^2}) \\
 \frac{\partial \mathbf{y}^\top \mathbf{A} \mathbf{y}}{\partial \mathbf{y}} &= \mathbf{y}^\top (\mathbf{A} + \mathbf{A}^\top) \\
 \frac{\partial \mathbf{B} \mathbf{A}}{\partial \mathbf{A}} &= (\mathbf{I}_J \otimes \mathbf{B})
 \end{aligned}$$

and the “vec-trick” $\text{vec}(\mathbf{X})^\top (\mathbf{B} \otimes \mathbf{A}^\top) = \text{vec}(\mathbf{A} \mathbf{X} \mathbf{B})^\top$.

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Bibliography

- Barratt S, Boyd S (2023). “Covariance Prediction via Convex Optimization.” *Optimization and Engineering*, **24**(3), 2045–2078. doi10.1007/s11081-022-09765-w. 115
- Genz A (1992). “Numerical Computation of Multivariate Normal Probabilities.” *Journal of Computational and Graphical Statistics*, **1**(2), 141–149. doi10.1080/10618600.1992.10477010. 1, 67
- Genz A, Bretz F (2002). “Methods for the Computation of Multivariate t Probabilities.” *Journal of Computational and Graphical Statistics*, **11**(4), 950–971. doi10.1198/106186002394. 1, 65
- Genz A, Bretz F (2009). *Computation of Multivariate Normal and t Probabilities*. Lecture Notes in Statistics. Springer-Verlag, Heidelberg, Germany. ISBN 978-3-642-01688-2. 139
- Hothorn T (2024). “On Nonparanormal Likelihoods.” *Technical report*, arXiv 2408.17346. doi10.48550/arXiv.2408.17346. 1
- Klein N, Hothorn T, Barbanti L, Kneib T (2022). “Multivariate Conditional Transformation Models.” *Scandinavian Journal of Statistics*, **49**, 116–142. doi10.1111/sjos.12501. 1
- Marsaglia G (1963). “Expressing the Normal Distribution with Covariance Matrix $A + B$ in Terms of One with Covariance Matrix A .” *Biometrika*, **50**(3–4), 535–538. 139
- Matić I, Radoičić R, Stefanica D (2018). “A Sharp Pólya-Based Approximation to the Normal CDF.” *Applied Mathematics and Computation*, **322**, 111–122. doi10.2139/ssrn.2842681. 71
- Prékopa A (1973). “On Logarithmic Concave Measures and Functions.” *Acta Scientiarum Mathematicarum*, **34**, 335–343. 115