

# Package ‘NOVA’

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**Type** Package

**Title** Neural Output Visualization and Analysis

**Version** 0.1.5

**Description** A comprehensive toolkit for analyzing and visualizing neural data outputs, including Principal Component Analysis (PCA) trajectory plotting, Multi-Electrode Array (MEA) heatmap generation, and variable importance analysis. Provides publication-ready visualizations with flexible customization options for neuroscience research applications.

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aggregate_data	<i>Aggregate Data by Groups</i>
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### Description

Aggregates values within groups using specified method

### Usage

```
aggregate_data(data, group_col, variable_column, value_column, method)
```

### Arguments

data	Data frame to aggregate
group_col	Column name for grouping
variable_column	Column name containing variable identifiers
value_column	Column name containing values to aggregate
method	Aggregation method: "mean", "median", "sum"

### Value

Aggregated data frame

## Examples

```
test_data <- data.frame(  
  Group = rep(c("A", "B"), each = 10),  
  Variable = rep(paste0("V", 1:5), 4),  
  Value = rnorm(20)  
)  
agg <- aggregate_data(test_data, "Group", "Variable", "Value", "mean")
```

---

analyze\_pca\_variable\_importance\_general  
*Analyze and Visualize PCA Variable Importance*

---

## Description

This function performs comprehensive analysis of variable importance in Principal Component Analysis, generating multiple visualization types including loading biplots, importance rankings, PC comparisons, and heatmaps. It extracts variable contributions to specified principal components and creates publication-ready plots with detailed statistical summaries.

## Usage

```
analyze_pca_variable_importance_general(  
  pca_result = NULL,  
  output_dir = tempdir(),  
  experiment_name = "PCA_Analysis",  
  pc_x = "PC1",  
  pc_y = "PC2",  
  color_scheme = "default",  
  top_n = 15,  
  min_loading_threshold = 0.1,  
  save_plots = TRUE,  
  show_labels = TRUE,  
  verbose = TRUE  
)
```

## Arguments

pca_result	A PCA result object. Can be either a prcomp object directly, or a list containing a PCA object in fields named 'pca_result', 'pca', 'result', or 'prcomp'.
output_dir	Character string specifying the directory for saving plots and results (default: "pca_plots").
experiment_name	Character string used as a prefix for output files and plot titles (default: "PCA_Analysis").
pc_x	Character string specifying the principal component for x-axis analysis (default: "PC1").

<code>pc_y</code>	Character string specifying the principal component for y-axis analysis (default: "PC2").
<code>color_scheme</code>	Character string specifying the color palette. Options: "default", "viridis", "colorbrewer" (default: "default").
<code>top_n</code>	Numeric value specifying the number of top variables to focus on in detailed analyses (default: 15).
<code>min_loading_threshold</code>	Numeric value specifying the minimum loading threshold for importance filtering (default: 0.1).
<code>save_plots</code>	Logical indicating whether to save plots and results to disk (default: TRUE).
<code>show_labels</code>	Logical indicating whether to show variable labels on the biplot (default: TRUE).
<code>verbose</code>	Logical indicating whether to print detailed progress messages (default: TRUE).

## Details

The function calculates multiple importance metrics for each variable:

- **PC loadings:** Direct loading values for specified principal components
- **Combined importance:** Euclidean distance combining both PC loadings
- **Contribution percentages:** Percent contribution to each PC's total variance
- **Ranking:** Variables ranked by combined importance score

Four visualization types are generated:

- **Loading Biplot:** Scatter plot showing variable loadings on both PCs with size indicating importance
- **Importance Bar Chart:** Ranked bar chart of top variables by combined importance
- **PC Comparison:** Side-by-side comparison of absolute loadings for both PCs
- **Loading Heatmap:** Color-coded matrix showing loading values and directions

The function automatically:

- Validates input PCA objects from various sources
- Calculates variance explained by each principal component
- Creates publication-ready plots with consistent theming
- Exports detailed CSV files with variable rankings and analysis summaries
- Provides comprehensive statistical summaries

Color schemes provide different aesthetic options:

- `default`: Blue/red palette suitable for most publications
- `viridis`: Colorblind-friendly viridis color scale
- `colorbrewer`: ColorBrewer palettes optimized for scientific visualization

View top variables using `head(results$selected_variables)`

**Value**

A list containing:

- plots** Named list of ggplot objects: 'biplot', 'importance\_bar', 'pc\_comparison', 'heatmap'
- variable\_importance** Data frame with comprehensive variable importance metrics for all variables
- selected\_variables** Data frame containing the top N most important variables with detailed statistics
- analysis\_summary** List with key analysis metrics and variance explained information
- config\_used** List documenting all parameters used in the analysis

**Output Files**

When `save_plots = TRUE`, the function creates files in the specified output directory (default: "pca\_plots"). For CRAN compliance, use `tempdir()` for the output directory:

- PNG files for each visualization type
- CSV file with complete variable importance rankings
- CSV file with selected top variables and detailed metrics
- CSV file with analysis summary and metadata

**See Also**

[prcomp](#) for PCA computation, [biplot](#) for basic PCA plotting

---

apply\_scaling\_enhanced

*Apply Enhanced Scaling Methods*

---

**Description**

Applies various scaling methods to matrix data for heatmap visualization

**Usage**

```
apply_scaling_enhanced(matrix_data, scale_method, verbose = FALSE)
```

**Arguments**

<code>matrix_data</code>	Numeric matrix to scale
<code>scale_method</code>	Scaling method: "variable_0_10", "robust", "row", "column", "none"
<code>verbose</code>	Whether to print scaling information

**Value**

Scaled matrix

---

clean\_heatmap\_matrix *Clean Heatmap Matrix*

---

**Description**

Removes rows and columns with insufficient finite values from matrix

**Usage**

```
clean_heatmap_matrix(matrix_data, min_finite = 2, verbose = FALSE)
```

**Arguments**

matrix_data	Numeric matrix to clean
min_finite	Minimum number of finite values required per row/column
verbose	Whether to print cleaning information

**Value**

Cleaned matrix or NULL if insufficient data

---

create\_annotations\_enhanced  
*Create Enhanced Annotations for Heatmaps*

---

**Description**

Creates annotation data frames and color schemes for heatmap visualization

**Usage**

```
create_annotations_enhanced(rownames_vector, factor_cols)
```

**Arguments**

rownames_vector	Vector of combined row names to parse
factor_cols	Vector of factor column names

**Value**

List containing annotations data frame and color schemes

---

create\_color\_palette\_enhanced  
*Create Enhanced Color Palettes*

---

### Description

Creates color palettes and breaks for heatmap visualization

### Usage

```
create_color_palette_enhanced(  
  palette_name = "yellow_purple",  
  custom_colors = NULL,  
  data_matrix = NULL  
)
```

### Arguments

palette_name	Name of color palette to use
custom_colors	Vector of custom colors (optional)
data_matrix	Data matrix to determine color range

### Value

List containing colors and breaks

---

create\_mea\_heatmaps\_enhanced  
*Create Enhanced Heatmaps for Multi-Electrode Array (MEA) Data Analysis*

---

### Description

This function generates comprehensive heatmap visualizations for MEA data analysis, including individual grouping variable heatmaps, combined interaction heatmaps, and variable correlation matrices. It provides flexible scaling, clustering, and customization options with automatic quality filtering and missing data handling.

**Usage**

```
create_mea_heatmaps_enhanced(  
  data = NULL,  
  processing_result = NULL,  
  config = NULL,  
  value_column = "Normalized_Value",  
  variable_column = "Variable",  
  grouping_columns = c("Treatment", "Genotype"),  
  sample_id_columns = c("Well"),  
  timepoint_column = "Timepoint",  
  scale_method = "z_score",  
  aggregation_method = "mean",  
  missing_value_handling = "remove",  
  cluster_method = "euclidean",  
  cluster_rows = TRUE,  
  cluster_cols = TRUE,  
  create_individual_heatmaps = TRUE,  
  create_combined_heatmap = TRUE,  
  create_variable_correlation = TRUE,  
  output_dir = NULL,  
  save_plots = FALSE,  
  plot_format = "png",  
  plot_width = 10,  
  plot_height = 8,  
  dpi = 300,  
  fontsize = 10,  
  angle_col = 45,  
  show_rownames = TRUE,  
  show_colnames = TRUE,  
  return_data = TRUE,  
  verbose = TRUE,  
  quality_threshold = 0.8,  
  min_observations = 3,  
  use_raw = FALSE,  
  filter_timepoints = NULL,  
  filter_treatments = NULL,  
  filter_genotypes = NULL,  
  split_by = NULL  
)
```

**Arguments**

<code>data</code>	A data frame containing MEA measurement data. If NULL, must provide <code>processing_result</code> .
<code>processing_result</code>	A list object from MEA data processing containing <code>normalized_data</code> or <code>raw_data</code> components. Takes precedence over the <code>data</code> parameter if provided.

config	Configuration list from MEA processing. If NULL and processing_result is provided, will attempt to use config from processing_result\$config_used.
value_column	Character string specifying the column containing measurement values (default: "Normalized_Value").
variable_column	Character string specifying the column containing variable names (default: "Variable").
grouping_columns	Character vector of column names to use for grouping (default: c("Treatment", "Genotype")). Function will auto-detect which columns are available.
sample_id_columns	Character vector of columns identifying individual samples (default: c("Well")).
timepoint_column	Character string specifying the timepoint column (default: "Timepoint").
scale_method	Character string specifying scaling method. Options: "z_score" (default), "min_max", "robust", "none".
aggregation_method	Character string specifying how to aggregate multiple measurements. Options: "mean" (default), "median", "sum".
missing_value_handling	Character string specifying how to handle missing values. Options: "remove" (default), "impute_mean", "impute_zero".
cluster_method	Character string specifying clustering distance method. Options: "euclidean" (default), "correlation", "manhattan".
cluster_rows	Logical indicating whether to cluster rows (default: TRUE).
cluster_cols	Logical indicating whether to cluster columns (default: TRUE).
create_individual_heatmaps	Logical indicating whether to create separate heatmaps for each grouping variable (default: TRUE).
create_combined_heatmap	Logical indicating whether to create interaction heatmap when multiple grouping variables are present (default: TRUE).
create_variable_correlation	Logical indicating whether to create variable correlation heatmap (default: TRUE).
output_dir	Character string specifying output directory (default: NULL, no files saved)
save_plots	Logical indicating whether to save plots to disk (default: FALSE)
plot_format	Character string specifying file format for saved plots (default: "png").
plot_width	Numeric value specifying plot width in inches (default: 10).
plot_height	Numeric value specifying plot height in inches (default: 8).
dpi	Numeric value specifying resolution for saved plots (default: 300).
fontsize	Numeric value specifying font size for heatmap labels (default: 10).
angle_col	Numeric value specifying angle for column labels in degrees (default: 45).
show_rownames	Logical indicating whether to show row names (default: TRUE).

<code>show_colnames</code>	Logical indicating whether to show column names (default: TRUE).
<code>return_data</code>	Logical indicating whether to return processed data matrices (default: TRUE).
<code>verbose</code>	Logical indicating whether to print progress messages (default: TRUE).
<code>quality_threshold</code>	Numeric value between 0-1 specifying minimum data completeness per variable (default: 0.8).
<code>min_observations</code>	Numeric value specifying minimum observations required per group (default: 3).
<code>use_raw</code>	Logical. If TRUE, plot raw electrode values instead of normalized values. Default FALSE.
<code>filter_timepoints</code>	Character vector of timepoint names to include. NULL (default) includes all timepoints.
<code>filter_treatments</code>	Character vector of treatment names to include. NULL (default) includes all treatments.
<code>filter_genotypes</code>	Character vector of genotype names to include. NULL (default) includes all genotypes.
<code>split_by</code>	Character string controlling plot splitting. Use "combination" to render a single heatmap of all wells annotated by both Treatment and Genotype strips. Pass any column name (e.g. "Treatment" or "Genotype") to produce one heatmap per level of that column. NULL (default) produces a single combined heatmap.

## Details

The function performs several key operations:

- Quality filtering: Removes variables with insufficient data completeness
- Missing value handling: Multiple strategies for dealing with NA values
- Data aggregation: Combines multiple measurements per group using specified method
- Scaling: Applies normalization methods appropriate for heatmap visualization
- Clustering: Hierarchical clustering of rows and/or columns using specified distance metrics
- Visualization: Creates publication-ready heatmaps with proper color schemes and annotations

For scaling methods:

- `z_score`: Centers data around mean with unit variance (best for comparing relative changes)
- `min_max`: Scales to 0-1 range (best for absolute comparisons)
- `robust`: Uses median and MAD for outlier-resistant scaling
- `none`: No scaling applied

The function automatically adjusts plot dimensions based on data size and uses optimized color palettes appropriate for the scaling method chosen (diverging palettes for `z_score`/`robust`, sequential palettes for `min_max`).

**Value**

A list containing:

**individual\_heatmaps** Named list of heatmap objects for each grouping variable

**combined\_heatmap** Heatmap object for grouping variable interactions (if applicable)

**variable\_correlation** List with correlation heatmap and correlation matrix

**metadata** List containing processing information and parameters used

Each heatmap object contains: heatmap (pheatmap object), scaled\_data (processed matrix), raw\_data (aggregated input data), annotation (row annotations), annotation\_colors (color schemes), and scaling\_info (scaling parameters).

---

discover\_mea\_structure

*Discover MEA Data Structure*

---

**Description**

This function scans a directory containing MEA (Multi-Electrode Array) experiment folders and analyzes the structure of CSV files to identify experiments, timepoints, measured variables, treatments, and genotypes. It provides a comprehensive overview of the data organization without loading all files into memory.

**Usage**

```
discover_mea_structure(
  main_dir,
  experiment_pattern = "MEA\\d+",
  file_pattern = "\\*.csv$",
  verbose = TRUE
)
```

**Arguments**

main_dir	Character. Path to the main directory containing experiment folders
experiment_pattern	Character. Regex pattern to identify experiment directories (default: "MEA\\d+")
file_pattern	Character. Regex pattern to identify data files (default: "\\*.csv\$")
verbose	Logical. Whether to print progress messages (default: TRUE)

**Details**

The function expects MEA CSV files with standard format: - Row 121: Well identifiers (A1, A2, B1, etc.) - Row 122: Treatment conditions - Row 123: Genotype information - Row 124: Exclusion flags - Rows 125-168: Variable names and measurements

Discover structure of MEA data (requires data directory)

**Value**

A list containing: - experiments: List of experiment info (directories, files, timepoints, metadata) - all\_timepoints: Vector of all unique timepoints found across experiments - all\_variables: Vector of all unique measured variables - potential\_baselines: Timepoints that might serve as baseline conditions - experiment\_count: Total number of experiments found - discovery\_timestamp: When the analysis was performed

---

handle\_missing\_values *Handle Missing Values in MEA Data*

---

**Description**

Handles missing values in MEA datasets using various imputation strategies or removal methods.

**Usage**

```
handle_missing_values(data, value_column, method, verbose)
```

**Arguments**

data	Data frame containing MEA data
value_column	Character string specifying the column with values to process
method	Character string specifying handling method: "remove", "impute_mean", "impute_zero"
verbose	Logical indicating whether to print progress messages

**Value**

Data frame with missing values handled according to specified method

**Examples**

```
test_data <- data.frame(  
  ID = 1:10,  
  Value = c(1.2, NA, 3.4, 2.1, NA, 5.6, 4.3, NA, 2.8, 3.9)  
)  
cleaned <- handle_missing_values(test_data, "Value", "remove", FALSE)
```

---

null_coalesce	<i>Null Coalescing Operator</i>
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---

**Description**

Returns the left-hand side if not NULL, otherwise the right-hand side

**Usage**

```
null_coalesce(lhs, rhs)
```

**Arguments**

lhs	Left-hand side value
rhs	Right-hand side value (default/fallback)

**Value**

lhs if not NULL, otherwise rhs

**Examples**

```
null_coalesce(5, 10)  
null_coalesce(NULL, 10)
```

---

pca_analysis_enhanced	<i>Enhanced PCA Analysis for MEA Data</i>
-----------------------	---

---

**Description**

This function performs Principal Component Analysis (PCA) on MEA data with extensive flexibility for data input sources, parameter configuration, and output options. It handles missing values, applies variance filtering, creates visualization plots, and provides comprehensive results suitable for downstream analysis.

**Usage**

```
pca_analysis_enhanced(  
  normalized_data = NULL,  
  data_path = NULL,  
  config = NULL,  
  processing_result = NULL,  
  min_var = NULL,  
  impute = NULL,  
  scale_data = NULL,
```

```

n_components = NULL,
variance_cutoff = NULL,
grouping_variables = NULL,
sample_id_components = NULL,
value_column = "Normalized_Value",
variable_column = "Variable",
timepoint_column = "Timepoint",
output_path = NULL,
verbose = TRUE
)

```

### Arguments

normalized_data	Data.frame. Pre-loaded MEA data in long format (default: NULL)
data_path	Character. Path to Excel file containing MEA data (default: NULL)
config	List. Configuration object with analysis parameters (default: NULL)
processing_result	List. Output from process_mea_flexible function (default: NULL)
min_var	Numeric. Minimum variance threshold for variable inclusion (default: 0.01)
impute	Logical. Whether to impute missing values (default: TRUE)
scale_data	Logical. Whether to scale variables before PCA (default: TRUE)
n_components	Integer. Number of principal components to extract (default: 2)
variance_cutoff	Numeric. Cumulative variance percentage threshold (default: 70)
grouping_variables	Character vector. Variables for sample grouping (default: c("Treatment", "Genotype"))
sample_id_components	Character vector. Variables to create unique sample IDs (default: c("Well", "Timepoint", "Treatment", "Genotype"))
value_column	Character. Name of column containing values for PCA (default: "Normalized_Value")
variable_column	Character. Name of column containing variable names (default: "Variable")
timepoint_column	Character. Name of column containing timepoint information (default: "Timepoint")
output_path	Character. Optional path to save elbow plot (default: NULL, no file saved)
verbose	Logical. Whether to print detailed progress messages (default: TRUE)

### Details

The function provides three flexible data input methods: 1. **processing\_result**: Direct output from process\_mea\_flexible function 2. **data\_path**: Path to Excel file with normalized\_data sheet 3. **normalized\_data**: Pre-loaded data frame in long format

Data processing includes: - Automatic detection of available columns - Flexible sample ID creation from specified components - Missing value imputation (mean, median, or zero) - Variance-based variable filtering - Automatic scaling option - Creation of elbow plot for component selection

The function handles common MEA data challenges: - Missing timepoint or treatment information - Inconsistent column naming - Mixed data types and missing values - Variable numbers of experiments and conditions

Method 1: Use output from MEA processing function `process_mea_flexible("/path/to/data", baseline_timepoint = "baseline")` `pca_analysis_enhanced(processing_result = mea_result)`

Method 2: Load from saved Excel file `pca_analysis_enhanced(data_path = "/path/to/processed_data.xlsx")`

Method 3: Use pre-loaded data with custom parameters `normalized_data = my_data`

## Value

A list containing: - `pca_result`: Complete `prcomp()` object with PCA results - `plot_data`: Data frame ready for plotting with PC scores and metadata - `variance_explained`: Vector of variance explained by each component - `cumulative_variance`: Vector of cumulative variance explained - `elbow_plot`: `ggplot2` object showing variance explained by components - `elbow_data`: Data frame underlying the elbow plot - `components_needed`: Number of components needed for various variance thresholds - `count_summary`: Summary of sample counts by groups (if applicable) - `data_info`: Information about data processing steps - `config_used`: Configuration parameters actually used - `processing_source`: Source of input data ("`processing_result`", "`excel_file`", or "`direct_data`")

---

`pca_plots_enhanced`      *Enhanced PCA Plotting for Neural and Omics Data*

---

## Description

Creates publication-ready PCA plots with scientific color palettes, flexible aesthetic mapping, and multiple visualization options. Designed specifically for neural activity and omics datasets with support for complex experimental designs including treatments, genotypes, and timepoints.

## Usage

```
pca_plots_enhanced(
  pca_output = NULL,
  plot_data = NULL,
  pca_result = NULL,
  output_dir = NULL,
  processing_result = NULL,
  experiment_name = NULL,
  grouping_variables = NULL,
  color_variable = "Treatment",
  shape_variable = "Genotype",
  secondary_shape_variable = "Timepoint",
  pannels_var = NULL,
  components = c(1, 2),
```

```

    gray_color_value = NULL,
    save_plots = FALSE,
    plot_width = 12,
    plot_height = 10,
    dpi = 300,
    verbose = TRUE
)

```

## Arguments

<code>pca_output</code>	List. Complete PCA output object from <code>pca_analysis_enhanced()</code> (optional)
<code>plot_data</code>	Data.frame. Data containing PC coordinates and metadata variables
<code>pca_result</code>	List. PCA result object (e.g., from <code>prcomp()</code> or <code>princomp()</code> )
<code>output_dir</code>	Character. Directory path for saving plots (default: NULL, no files saved)
<code>processing_result</code>	List. Result object from <code>process_mea_flexible()</code> (optional)
<code>experiment_name</code>	Character. Name for the experiment (used in titles and filenames)
<code>grouping_variables</code>	Character vector. Available metadata variables for plotting (default: <code>c("Treatment", "Genotype", "Timepoint")</code> )
<code>color_variable</code>	Character. Variable name for color aesthetic (default: "Treatment")
<code>shape_variable</code>	Character. Variable name for shape aesthetic (default: "Genotype")
<code>secondary_shape_variable</code>	Character. Alternative shape variable (default: "Timepoint")
<code>pannels_var</code>	Character. Variable for panel faceting (default: NULL)
<code>components</code>	Numeric vector. PC components to plot (default: <code>c(1, 2)</code> )
<code>gray_color_value</code>	Character. Specific value of <code>color_variable</code> to display in gray (default: NULL)
<code>save_plots</code>	Logical. Whether to save plots to files (default: FALSE)
<code>plot_width</code>	Numeric. Plot width in inches (default: 12)
<code>plot_height</code>	Numeric. Plot height in inches (default: 10)
<code>dpi</code>	Numeric. Plot resolution (default: 300)
<code>verbose</code>	Logical. Whether to print progress messages (default: TRUE)

## Details

The function creates up to 5 different plot variants. Files are only saved when `save_plots = TRUE` AND `output_dir` is explicitly provided.

**Value**

A list containing:

**plots** Named list of ggplot objects for each plot type

**plot\_data** Data.frame with plotting data and metadata

**variance\_explained** Numeric vector of variance explained by each component

**components\_plotted** Numeric vector of components used in plots

**color\_palette** Named character vector of colors used

**shape\_palette** Named numeric vector of shapes used

**plotting\_config** List of configuration parameters used

**saved\_files** Character vector of saved file paths (if save\_plots = TRUE)

**See Also**

[process\\_mea\\_flexible](#) for MEA data processing, [discover\\_mea\\_structure](#) for automatic data structure detection

---

perform_mea_pca	<i>Perform MEA PCA Analysis</i>
-----------------	---------------------------------

---

**Description**

Template function for performing PCA on MEA data

**Usage**

```
perform_mea_pca(data, variables = NULL, scale = TRUE, center = TRUE, ...)
```

**Arguments**

data	Data frame or tibble with processed MEA data
variables	Character vector. Variables to include in PCA (if NULL, uses all numeric)
scale	Logical. Whether to scale variables before PCA (default: TRUE)
center	Logical. Whether to center variables before PCA (default: TRUE)
...	Additional PCA parameters

**Value**

List containing PCA results (scores, loadings, variance explained, etc.)

Perform PCA analysis (requires processed MEA data)

---

plot\_mea\_metric      *Plot a Single MEA Metric Across Conditions*

---

### Description

Creates a bar (mean + error), box, violin, or line plot for one measured variable from processed MEA data.

### Usage

```
plot_mea_metric(
  data,
  metric,
  x_var = "Timepoint",
  group_by = "Treatment",
  facet_by = NULL,
  filter_timepoints = NULL,
  filter_treatments = NULL,
  filter_genotypes = NULL,
  value_column = NULL,
  error_type = c("sem", "sd", "ci95"),
  plot_type = c("bar", "box", "violin", "line"),
  colors = NULL,
  show_points = TRUE,
  point_alpha = 0.6,
  title = NULL
)
```

### Arguments

data	Data frame - long-format MEA data (must contain 'Variable' column).
metric	Character. Exact name of the variable to plot.
x_var	Character. Column to use as the x-axis (default "Timepoint").
group_by	Character. Column to use for fill/colour grouping (default "Treatment").
facet_by	Character or NULL. Column name for faceting. NULL = no facets.
filter_timepoints	Character vector or NULL. Subset to these timepoints.
filter_treatments	Character vector or NULL. Subset to these treatments.
filter_genotypes	Character vector or NULL. Subset to these genotypes.
value_column	Character. Which column holds the numeric values. Defaults to "Normalized_Value" if present, else "Value".
error_type	Character. "sem" (default), "sd", or "ci95".

plot_type	Character. "bar" (default), "box", "violin", or "line".
colors	Named character vector of colours, or NULL for ggplot2 defaults.
show_points	Logical. Overlay individual data points (default TRUE).
point_alpha	Numeric. Transparency of data points (default 0.6).
title	Character or NULL. Plot title. NULL = metric name.

**Value**

A ggplot object.

**Examples**

```
## Not run:
plot_mea_metric(processed$all_data, "Mean Firing Rate (Hz)")
plot_mea_metric(processed$all_data, "Burst Rate (Hz)",
                plot_type = "violin", facet_by = "Genotype")

## End(Not run)
```

---

plot\_pca\_trajectories\_general

*Plot PCA Trajectories for Time Series Data*

---

**Description**

This function creates comprehensive visualizations of PCA trajectories over time, showing both individual and group-averaged trajectories with optional smoothing.

**Usage**

```
plot_pca_trajectories_general(
  pca_results,
  pc_x = "PC1",
  pc_y = "PC2",
  trajectory_grouping = NULL,
  timepoint_var = "Timepoint",
  timepoint_order = NULL,
  individual_var = "Experiment",
  point_size = 3,
  alpha = 0.7,
  line_size = 2,
  smooth_lines = FALSE,
  color_palette = NULL,
  color_by = "group",
  save_plots = FALSE,
  output_dir = NULL,
```

```

plot_prefix = "PCA_trajectories",
width = 12,
height = 8,
dpi = 150,
return_list = TRUE,
verbose = TRUE
)

```

### Arguments

<code>pca_results</code>	A data frame or list containing PCA results
<code>pc_x</code>	Character string specifying the principal component for x-axis (default: "PC1")
<code>pc_y</code>	Character string specifying the principal component for y-axis (default: "PC2")
<code>trajectory_grouping</code>	Character vector of column names for grouping trajectories
<code>timepoint_var</code>	Character string specifying the timepoint column (default: "Timepoint")
<code>timepoint_order</code>	Character vector specifying the order of timepoints
<code>individual_var</code>	Character string for individual trajectory identification (default: "Experiment")
<code>point_size</code>	Numeric value controlling point size (default: 3)
<code>alpha</code>	Numeric value controlling transparency (default: 0.7)
<code>line_size</code>	Numeric value controlling line thickness (default: 2)
<code>smooth_lines</code>	Logical indicating whether to apply smoothing (default: FALSE)
<code>color_palette</code>	Character vector of colors for groups
<code>color_by</code>	Character string controlling colour mapping. Use "group" (default) to colour by the full trajectory_grouping combination, or "Treatment" to colour by Treatment only with Genotype labels shown at each trajectory's end point via ggrepel.
<code>save_plots</code>	Logical indicating whether to save plots (default: FALSE)
<code>output_dir</code>	Character string specifying output directory (default: NULL)
<code>plot_prefix</code>	Character string prefix for filenames (default: "PCA_trajectories")
<code>width</code>	Numeric plot width in inches (default: 12)
<code>height</code>	Numeric plot height in inches (default: 8)
<code>dpi</code>	Numeric plot resolution (default: 150)
<code>return_list</code>	Logical indicating whether to return results as list (default: TRUE)
<code>verbose</code>	Logical indicating whether to print messages (default: TRUE)

### Value

A list containing plots, trajectories, and metadata

---

`print_detailed_summary`*Print Detailed PCA Variable Summary*

---

**Description**

Prints formatted summary of PCA variable importance analysis

**Usage**

```
print_detailed_summary(  
  top_vars,  
  pc_x_top,  
  pc_y_top,  
  high_both,  
  pc_x,  
  pc_y,  
  top_n,  
  min_loading_threshold  
)
```

**Arguments**

<code>top_vars</code>	Data frame of top variables by combined importance
<code>pc_x_top</code>	Data frame of top variables for first PC
<code>pc_y_top</code>	Data frame of top variables for second PC
<code>high_both</code>	Data frame of variables important in both PCs
<code>pc_x</code>	Name of first principal component
<code>pc_y</code>	Name of second principal component
<code>top_n</code>	Number of top variables to display
<code>min_loading_threshold</code>	Minimum loading threshold

**Value**

NULL (prints to console)

---

process\_mea\_flexible *Process MEA Data Flexibly*

---

### Description

This function processes Multi-Electrode Array (MEA) data files by reading CSV files, extracting measurements and metadata, applying filters, and optionally normalizing to baseline conditions. It automatically excludes standard deviation variables and handles exclusion flags to produce clean, analysis-ready datasets.

### Usage

```
process_mea_flexible(
  main_dir,
  selected_experiments = NULL,
  selected_timepoints = NULL,
  grouping_variables = c("Treatment", "Genotype"),
  baseline_timepoint = NULL,
  unique_id_vars = c("Well", "Variable"),
  exclude_std_variables = TRUE,
  experiment_pattern = "MEA\\d+",
  timepoint_fusions = NULL,
  verbose = TRUE,
  output_path = NULL
)
```

### Arguments

main_dir	Character. Path to the main directory containing experiment folders
selected_experiments	Character vector. Experiment names to process (default: NULL = all)
selected_timepoints	Character vector. Timepoints to include (default: NULL = all)
grouping_variables	Character vector. Metadata columns to include ("Treatment", "Genotype")
baseline_timepoint	Character. Timepoint to use for normalization (default: NULL = no normalization)
unique_id_vars	Character vector. Variables that uniquely identify observations for normalization
exclude_std_variables	Logical. Whether to automatically exclude standard deviation variables (default: TRUE)
experiment_pattern	Character. Regex pattern for experiment directories (default: "MEA\\d+")
timepoint_fusions	Timepoint fusions to generate

verbose	Logical. Whether to print progress messages (default: TRUE)
output_path	Character. Optional path for output file (default: NULL saves to main_dir with auto-generated name)

### Details

The function automatically detects and excludes variables containing "Std", "std", or "STD" in their names (e.g., "Number of Spikes - Std") while keeping average/mean variables (e.g., "Number of Spikes - Avg"). Wells marked with "Ex" or "ex" in row 124 are excluded.

By default, no files are written. To save output, provide an explicit output\_path parameter. Normalization creates fold-change values relative to baseline timepoint.

Process data without saving (returns data frames only) Save output by providing explicit path

### Value

A list containing: - raw\_data: Processed data in long format - normalized\_data: Baseline-normalized data (if baseline\_timepoint specified) - processing\_params: List of parameters used for processing - output\_path: Path to saved Excel file (only if output\_path was provided) - experiment\_name: Combined experiment identifier

---

quality_filter	<i>Filter Data by Quality Metrics</i>
----------------	---------------------------------------

---

### Description

Filters variables and groups based on observation counts and data completeness

### Usage

```
quality_filter(
  data,
  variable_column,
  value_column,
  grouping_columns,
  quality_threshold,
  min_observations,
  verbose
)
```

### Arguments

data	Data frame to filter
variable_column	Column name containing variable identifiers
value_column	Column name containing values to assess

grouping\_columns      Vector of column names for grouping  
 quality\_threshold      Minimum data completeness ratio (0-1)  
 min\_observations      Minimum number of observations required  
 verbose                Whether to print filtering results

**Value**

Filtered data frame

**Examples**

```
test_data <- data.frame(
  Variable = rep(paste0("V", 1:5), each = 20),
  Value = rnorm(100),
  Group = rep(c("A", "B"), 50)
)
filtered <- quality_filter(test_data, "Variable", "Value", "Group",
  0.8, 5, FALSE)
```

---

setup\_color\_scheme      *Setup Color Scheme*

---

**Description**

Sets up color schemes for plotting functions

**Usage**

```
setup_color_scheme(color_scheme, custom_colors)
```

**Arguments**

color\_scheme      Name of color scheme to use  
 custom\_colors      Custom color list (optional)

**Value**

List of colors for plotting

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